

Improving HPLC Column Selection and System Performance

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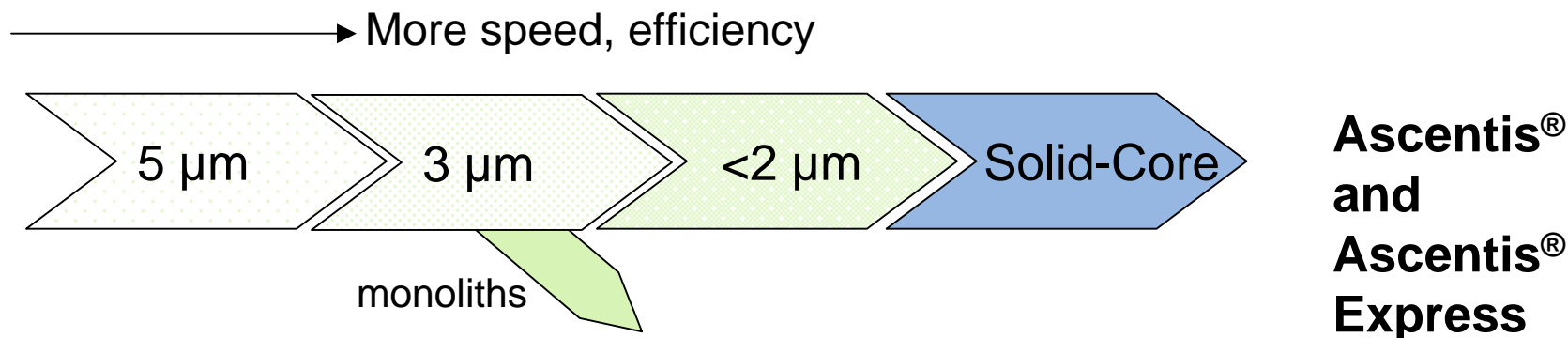


sigma-aldrich.com

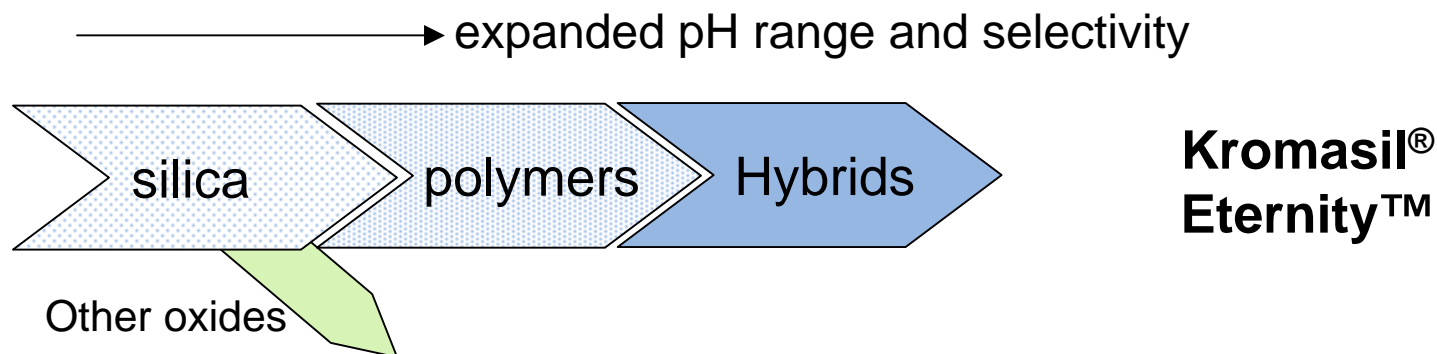
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LC Particle Innovation Leads the Way*

Particle Size and Architecture (porosity)



Particle Composition (and stationary phases)



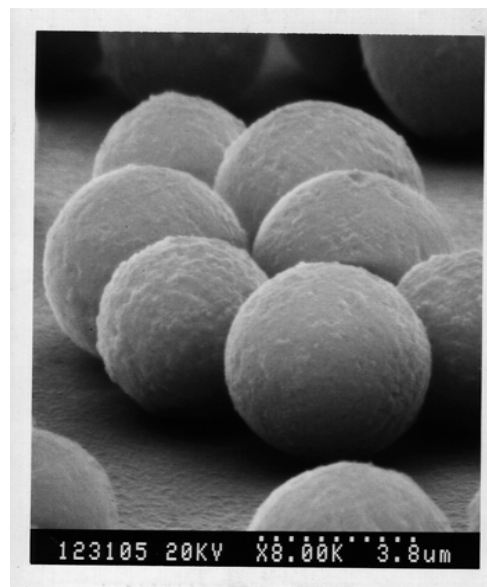
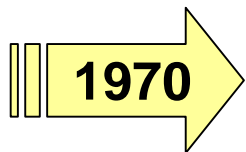
Discovery[®] Zr

* From Frank Michel

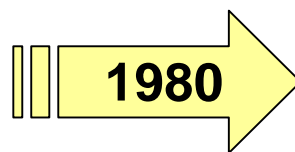
Evolution of HPLC Column Particle Shapes



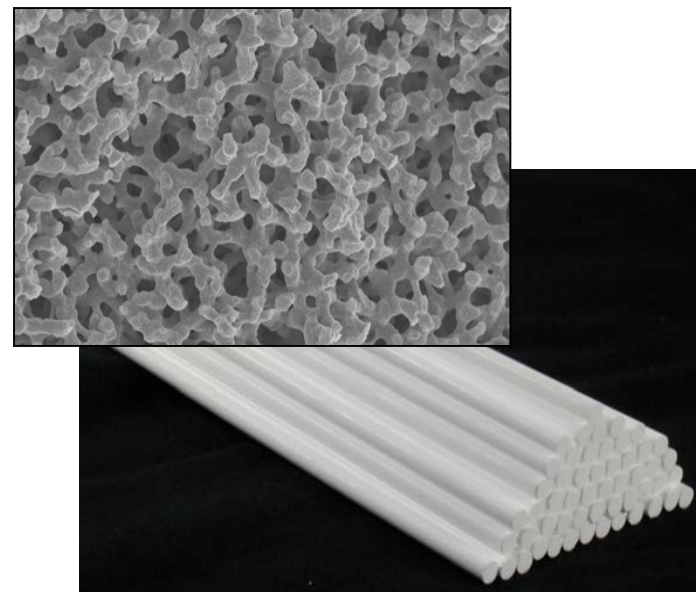
Irregular particles



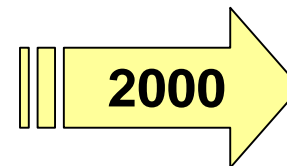
Spherical particles



Remain the workhorse

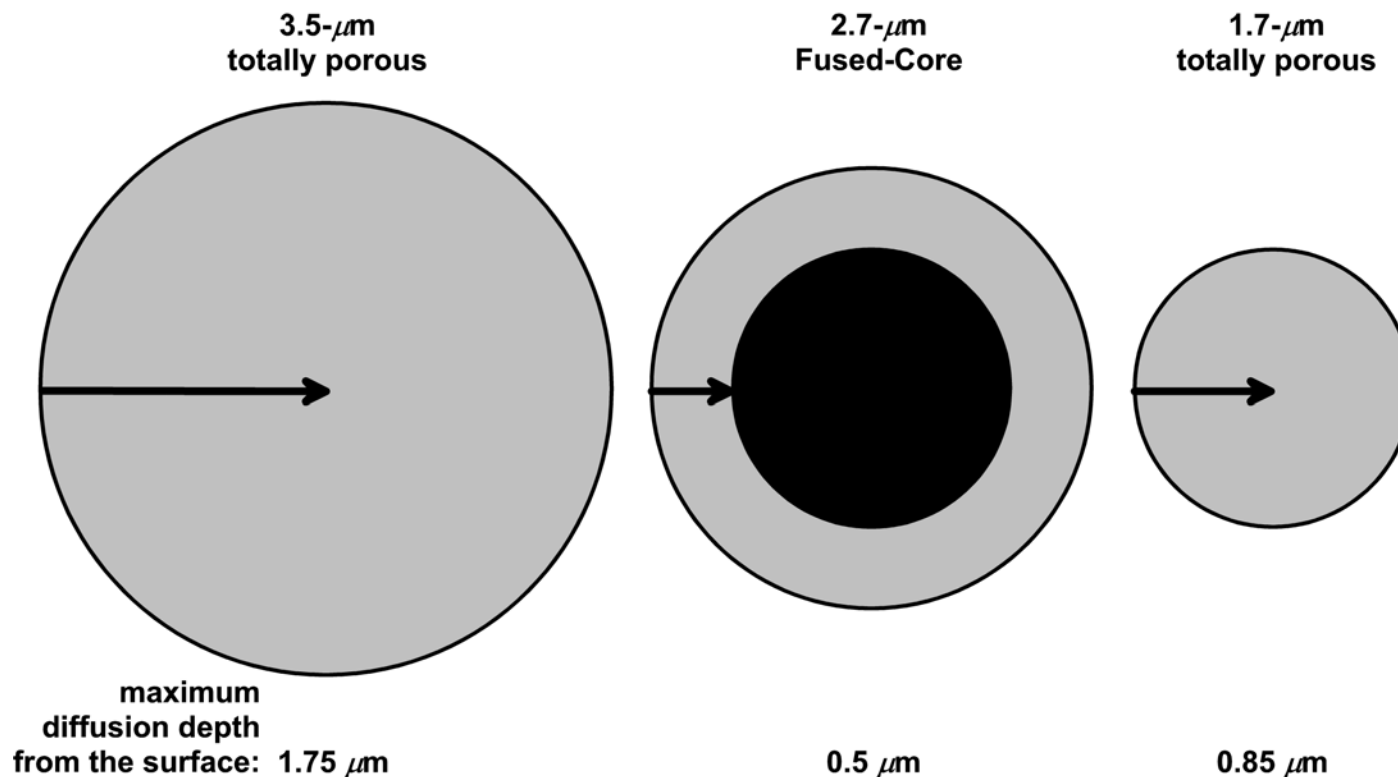


Monoliths



Particle Innovation has Reduced Column Dispersion*

...more efficiency, narrower peaks, higher peak capacity, more sensitivity.

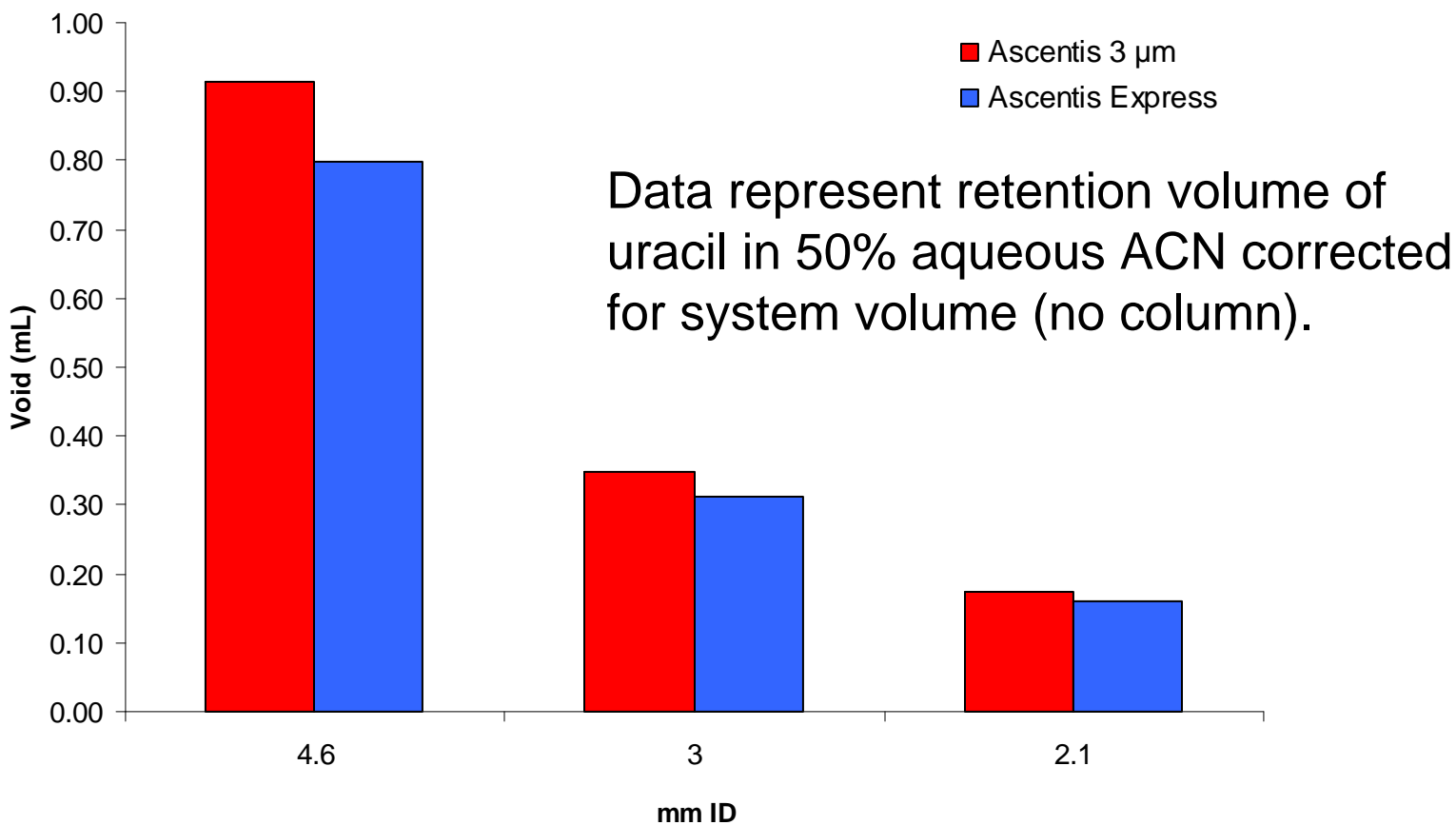


* T. L. Chester, American Lab, Vol. 41 No. 4, pp 11-15, March 2009.

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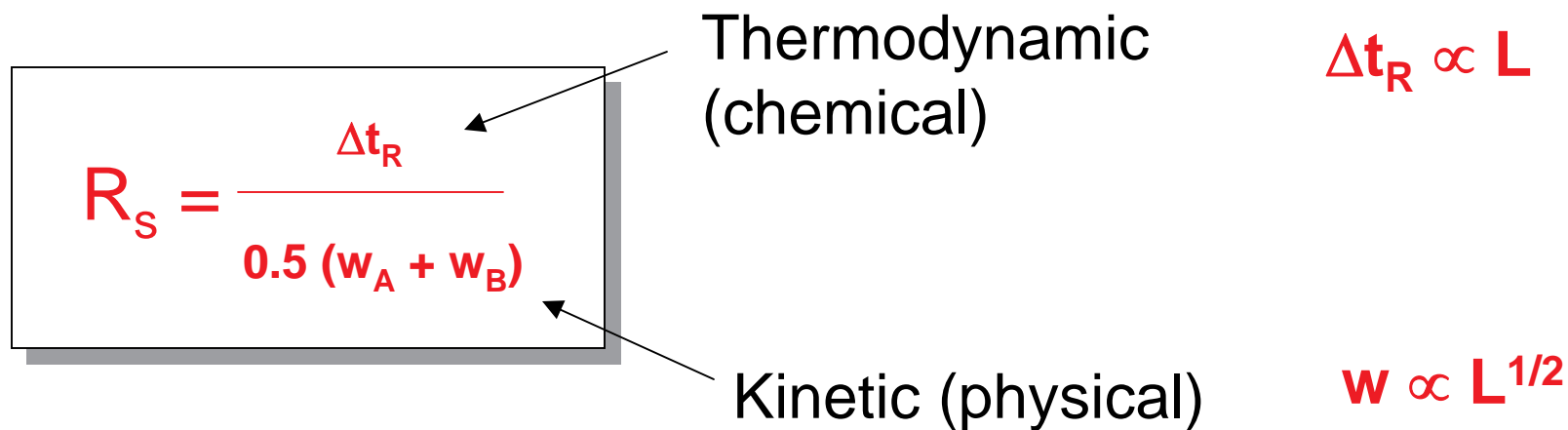
Measured Void Volumes: Solid-Core vs Porous



10cm x 4.6mm C18 Columns

Resolution: Peaks Separate Faster Than They Spread

- Maximize: Differential migration (Δt_R) - selectivity
- Minimize: Band dispersion (w) - efficiency



The diagram shows the resolution equation $R_S = \frac{\Delta t_R}{0.5(w_A + w_B)}$ enclosed in a box. Two arrows point from the text labels to the equation: one from 'Thermodynamic (chemical)' to the numerator Δt_R , and one from 'Kinetic (physical)' to the denominator $0.5(w_A + w_B)$. To the right of the box, the relationships $\Delta t_R \propto L$ and $w \propto L^{1/2}$ are written in red.

$$R_S = \frac{\Delta t_R}{0.5(w_A + w_B)}$$

Thermodynamic (chemical) $\Delta t_R \propto L$

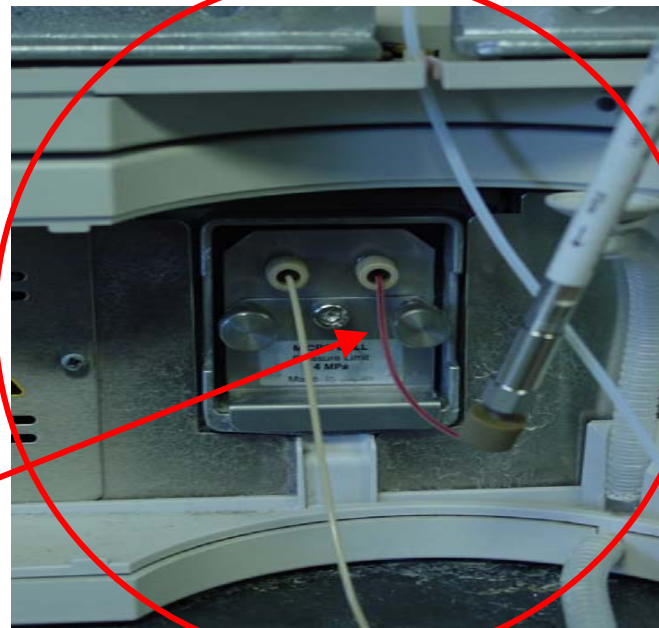
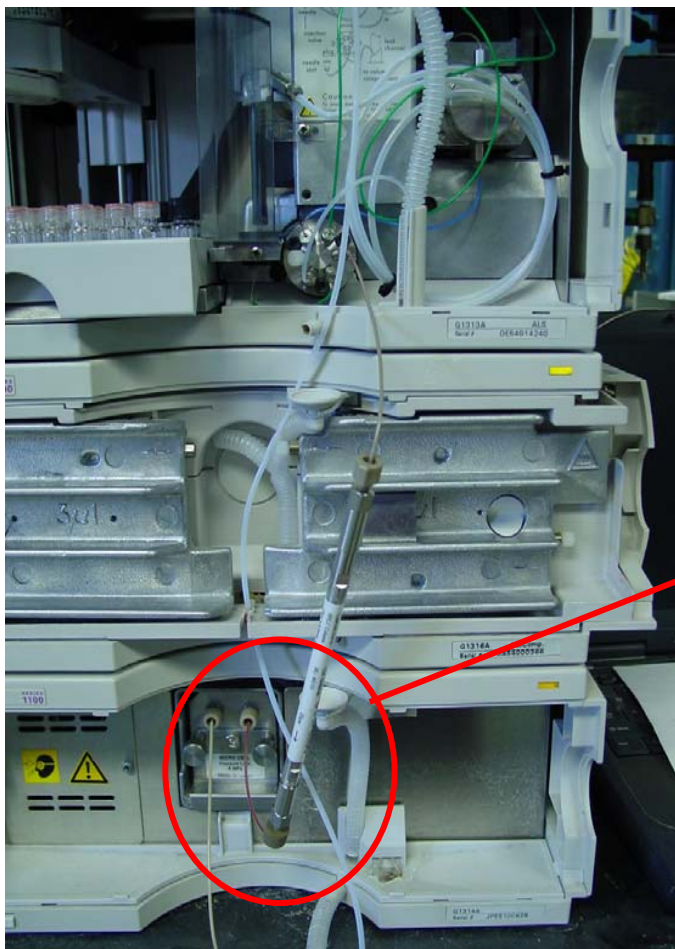
Kinetic (physical) $w \propto L^{1/2}$

Improvements have been made in both areas

Measuring HPLC System Suitability



HPLC System Organization and Optimization



Injector and detector tubing for the Agilent 1100 were 0.007 inches ID. A 10 μ L flow cell was employed. Experiments are underway to examine how tubing, flow cells and other HPLC system variables affect performance.

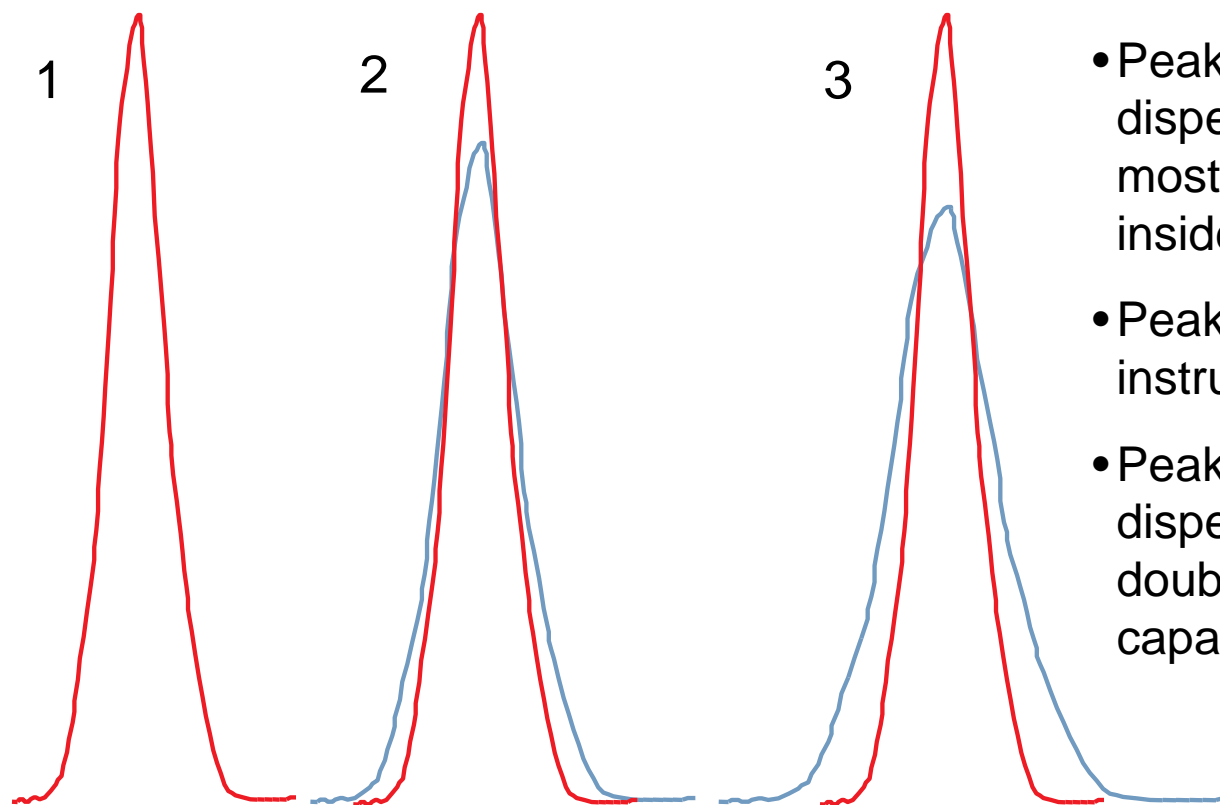
Peak Band Spreading Equation

$$\sigma_{\text{sys}}^2 = \sigma_{\text{col}}^2 + \sigma_{\text{instr}}^2 \quad \sigma_{\text{sys}} = (\sigma_{\text{col}}^2 + \sigma_{\text{instr}}^2)^{1/2}$$

- **Band spreading (dispersion) can occur in both HPLC columns and instruments leading to a *system equation*; instrument dispersion is also referred to as instrument bandwidth (IBW)**
- **New developments in particles have greatly reduced column dispersion; instrument brand, model and configuration now matters; *system* performance can be dramatically different for column and instrument combinations; *can't use a column without an instrument!***
- **New instruments with higher pressure ratings and smaller volume tubing and components have been designed for modern, smaller particle columns; however, traditional instruments may also be suitable for modern columns, especially when optimized by the user; simple tests can qualify instruments for minimum dispersion.**

Illustration of Instrument Dispersion

A peak from the same column in three different instruments.



- Peak 1 shows a low dispersion instrument where most of the spreading occurs inside the column.
- Peak 2 shows moderate instrument dispersion in blue.
- Peak 3 shows high instrument dispersion; peak width has doubled and column peak capacity is halved.

Time spent outside the column destroys system efficiency

Origins of Column Band Spreading* (van Deemter)

$$H = A d_p + B D_m / u + C d_p^2 u / D_m$$

- **A term**- Multipath; bed uniformity; eddy diffusion
- **B term**- Longitudinal diffusion (axial in nature)
- **C term**- Rate of mass transfer from moving phase through stagnant mobile phase into stationary phase (radial in nature)

Instrument impact on H and N is often neglected

Band Spreading Inside the Column Bed

$$\sigma_{col}^2 = V_o^2 (1 + k)^2 / N$$

V_o = mobile phase column volume (μL)
(unretained peak retention volume; void volume)

k = peak capacity factor

N = number of column theoretical plates

- **Small bed geometry, short retention and high efficiency favor low dispersion (dilution) within a packed column.**
- **Instrument bandwidth becomes more harmful to efficiency and resolution for short, small ID columns with low k values and high N .**

Column vs System Band-Spreading

The effect of system band width can be calculated from the additive relationship of variances, where the total variance of the peak is equal to the sum of the true on-column peak variance plus the instrument variance.

$$\sigma^2_{\text{system}} = \sigma^2_{\text{column}} + \sigma^2_{\text{instrument}}$$

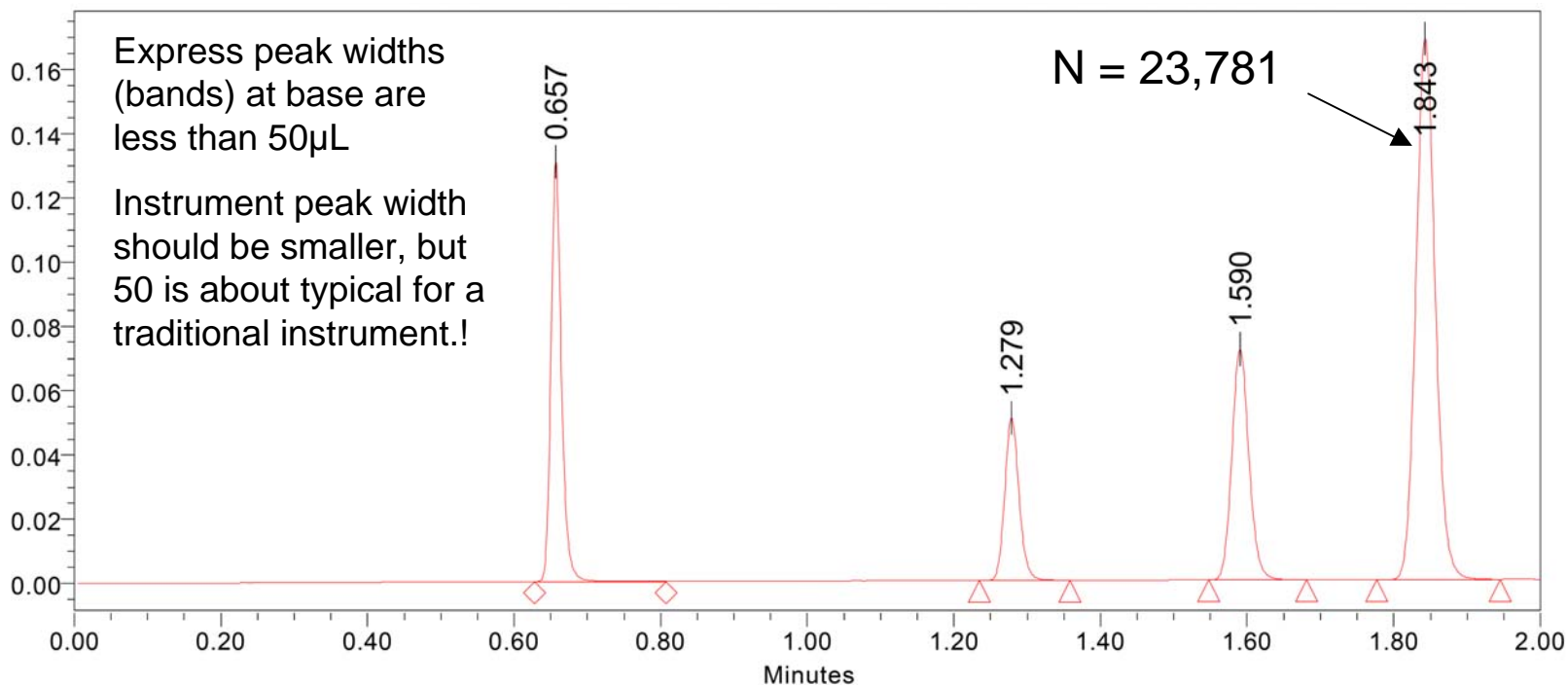
$$\sigma^2_{\text{instrument}} = \sigma^2_{\text{injector}} + \sigma^2_{\text{detector}} + \sigma^2_{\text{connector tubing}}$$

$$\sigma_{\text{system}} = \text{Total peak dispersion in } \underline{\text{volume units}} \text{ (}\mu\text{L)}$$

$$\sigma_{\text{system}} = W_b/4$$

$W_b = 4\sigma$ is an easy way
to estimate dispersion

Test Mix Chromatogram on Ascentis Express Fused-Core C18*



Sample: Uracil, benzene, toluene and anthracene

Column: Ascentis Express C18, 100x4.6mm

Mobile phase: 30/70 water/ACN; Flow: 1.25 mL/min; T = 35°C

Instrument: Waters Acquity, 220 bar (3200 psi)

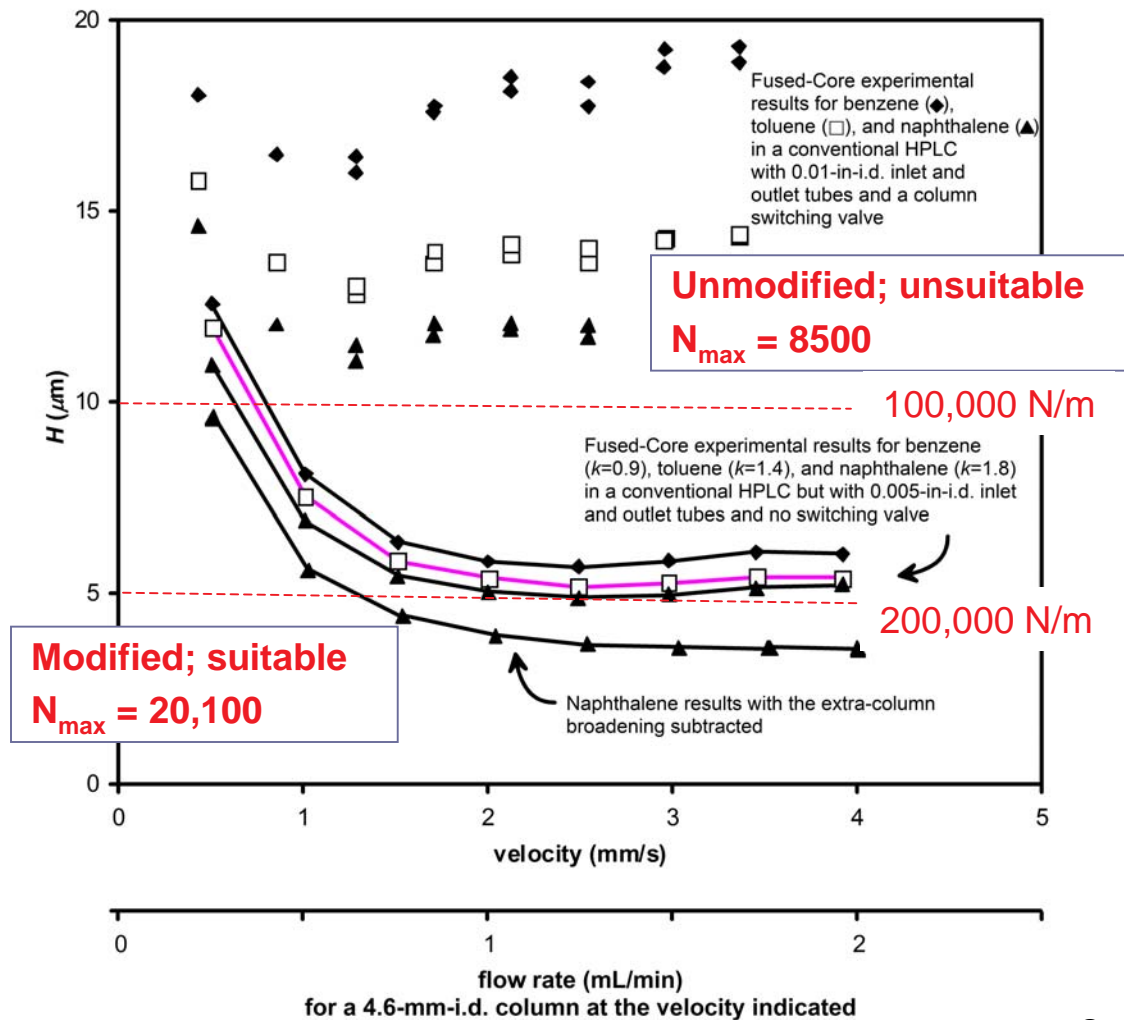
* T. L. Chester, American Lab, Vol. 41 No. 4, pp 11-15, March 2009.

Fused-Core Measured Plate Height for Unmodified and Modified Instrument³

% of Total Dispersion

	Unmodified	Modified
Inj	<1	1
Col	30	72
Conn Tubes	66	8
Det*	4	19

* detector cell had a welded heat exchanger that could not be conveniently replaced (Waters Alliance).

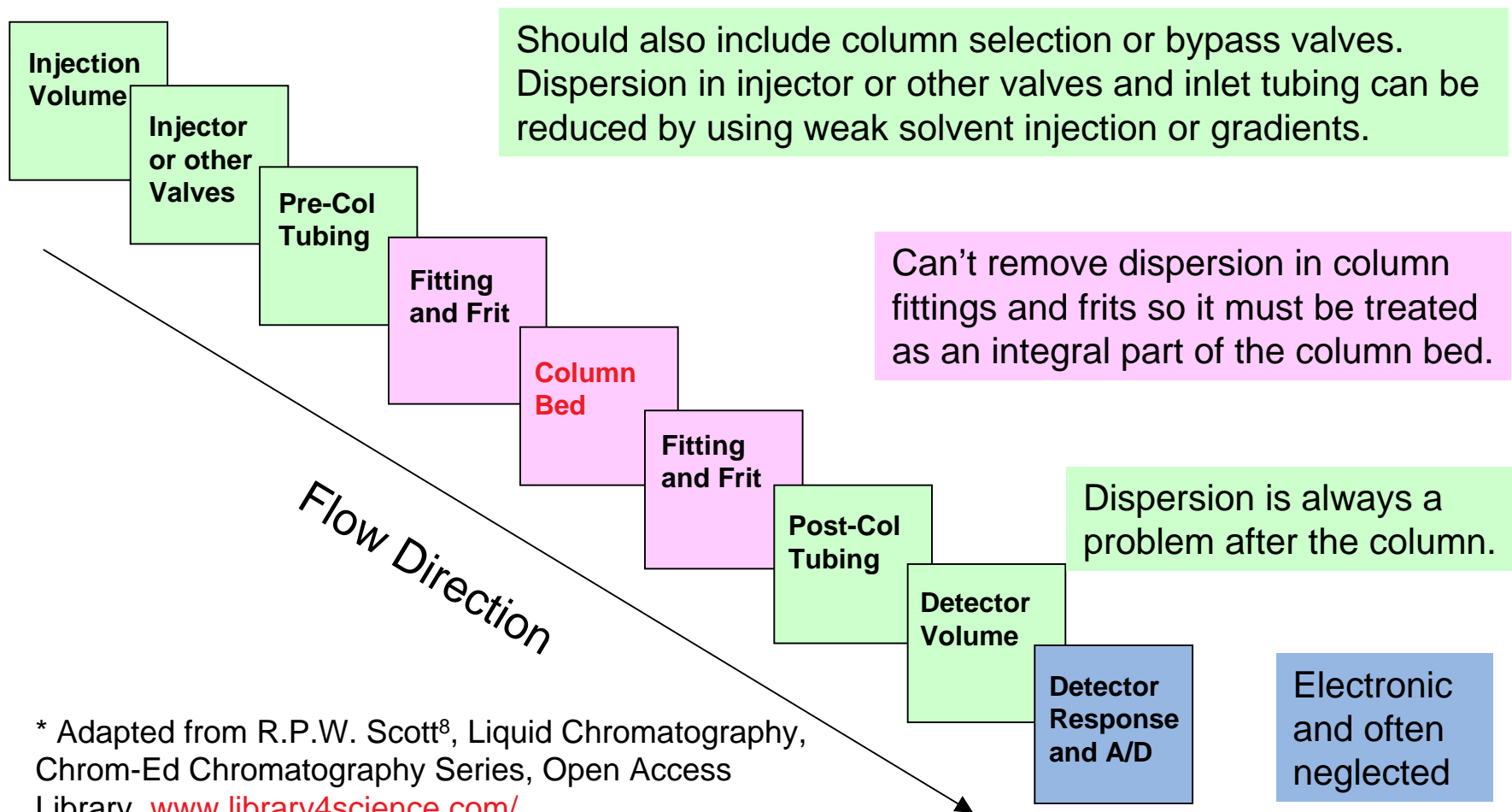


What Causes HPLC Instrument Dispersion?

.... Time sample spends outside the column bed.

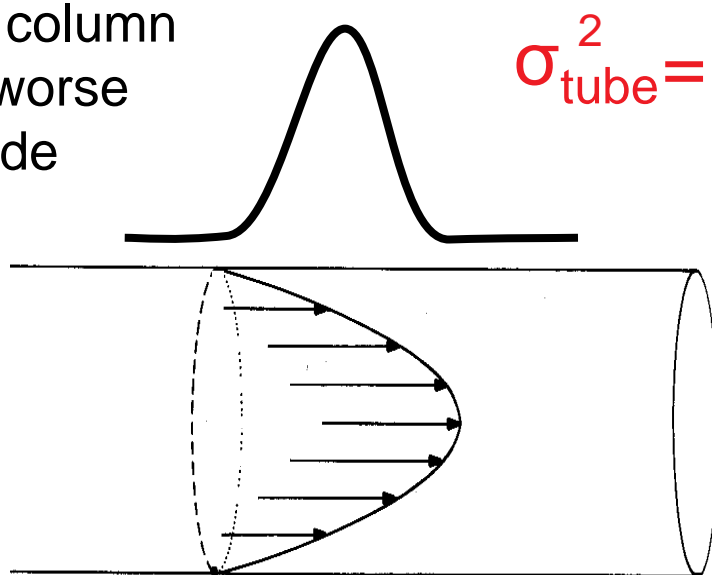


HPLC System Components that Affect Measured Peak Bandwidth*



Dispersion in Open Cylindrical Flow Path

Time outside column bed is much worse than time inside



$$\sigma_{\text{tube}}^2 = 1.36 \times 10^{-3} d_t^4 L_t F/D$$

d = cylinder diameter

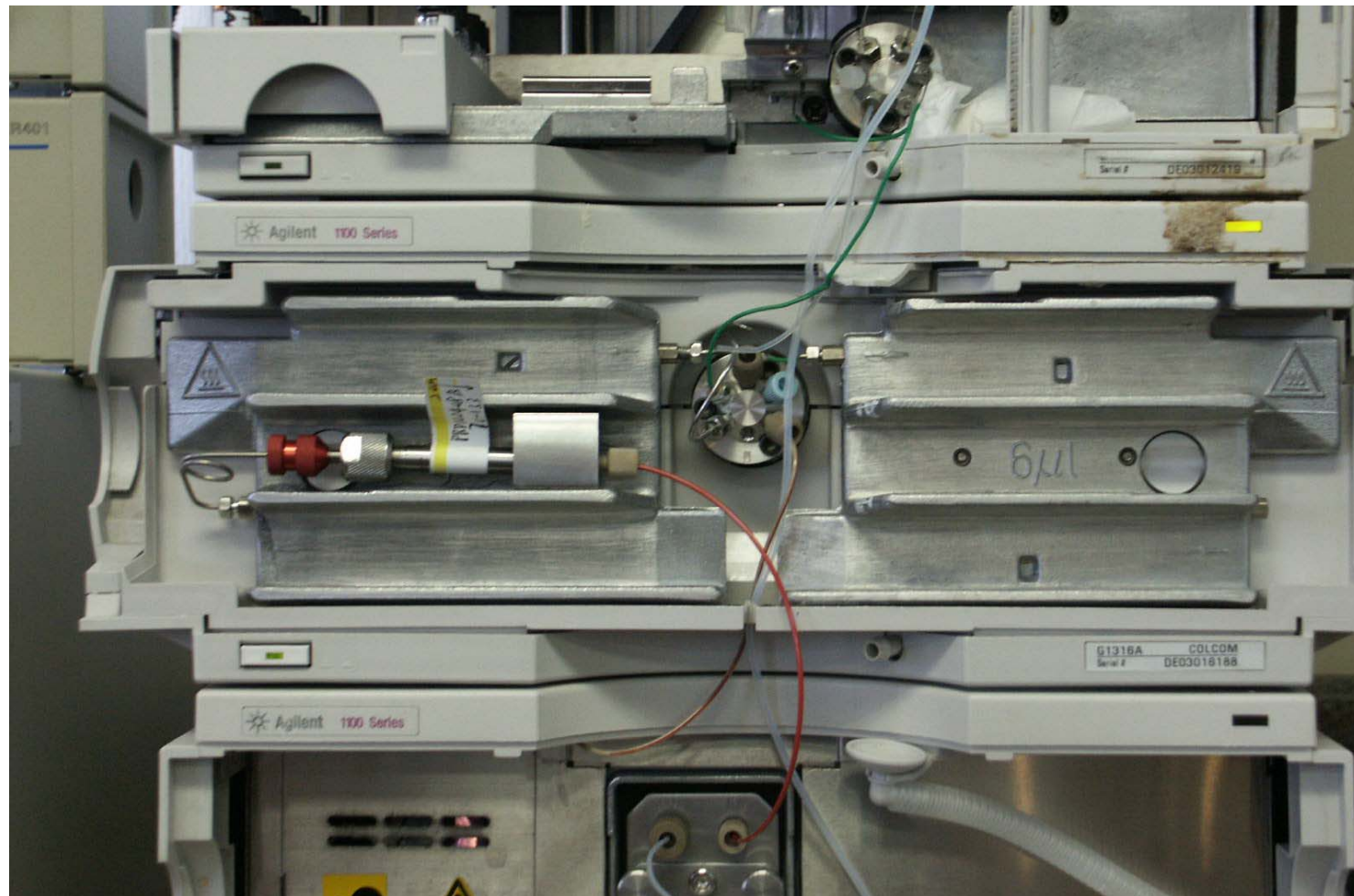
L = cylinder length

F = flow rate

D = diffusion coefficient

- **Dispersion from volume elements is constant for any given flow rate and analyte, but note that dispersion (bandwidth) increases with flow.**
- **Velocity at the wall is essentially zero under laminar flow conditions. Small inside diameter, short length, low flow and fast solute diffusion favor low dispersion in connection tubes and accessories. Larger molecules show greater dispersion (as 1/D) in connectors.**

Optimizing Instrument Configuration for Elevated Temperature Operation*



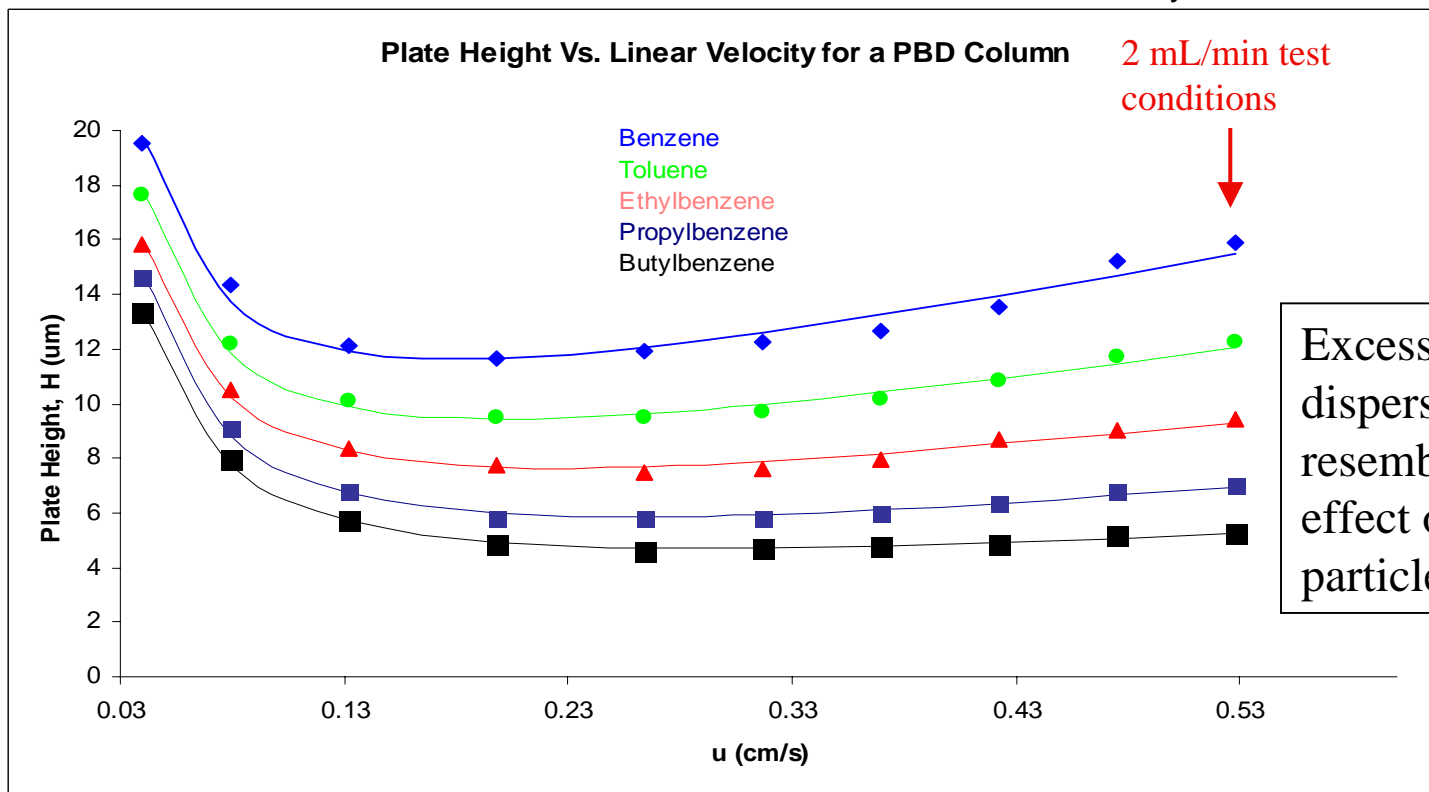
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*Agilent 1100 photo compliments of Dan Nowlan at ZirChrom

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Performance of a Factory Agilent 1100 Instrument with Sub- $2\mu\text{m}$ Zirconia

A family of curves with H_{\min} ranging from $<5\mu\text{m}$ to $>12\mu\text{m}$ indicates strong instrument impact on H_{system}

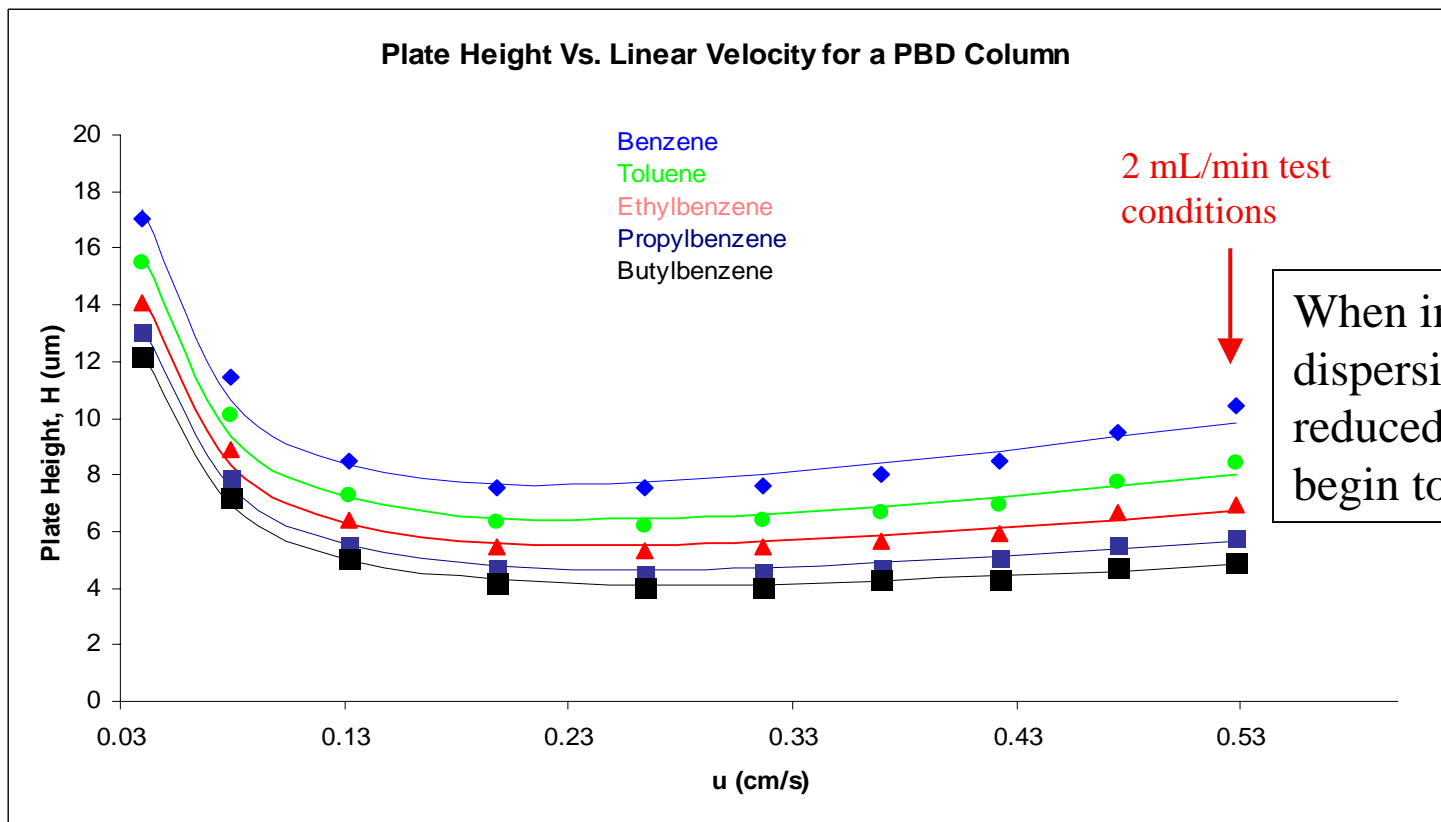


Excess instrument dispersion resembles the effect of larger particles (C-term).

Plate height vs linear velocity, Temperature 30 °C, Mobile phase: 50/50 ACN/water, Column: 50 x 4.6mm, Agilent 1100/UV with Standard Cell and 0.007" i.d. tubing.

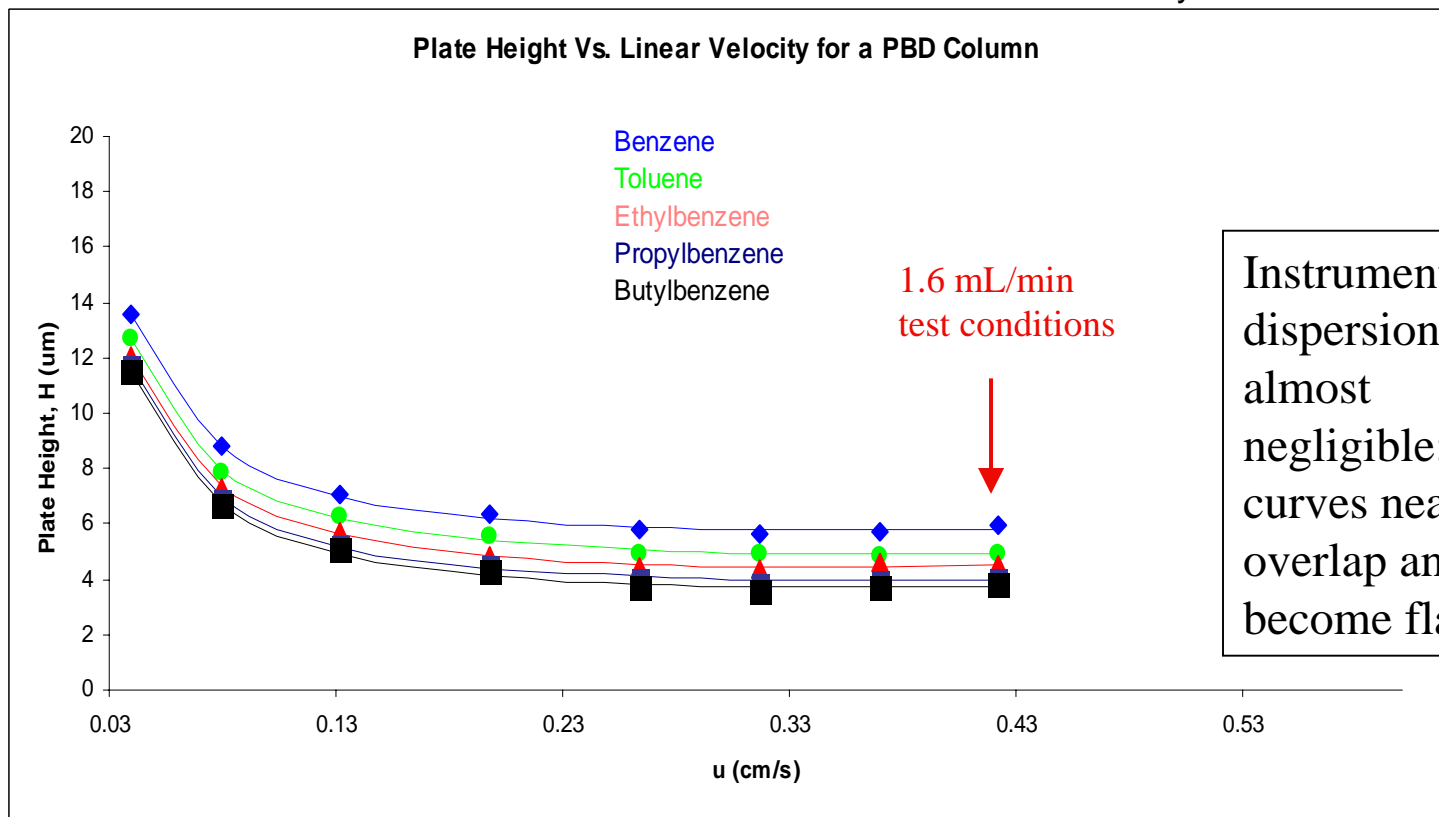
Optimizing Factory Agilent 1100 by Adding Micro Flow Cell

New family of curves with H_{min} ranging from $<5\mu m$ to $8\mu m$ indicates lower instrument impact on H_{system}



Optimized Factory Instrument with Micro Cell + 0.005" ID Tubing

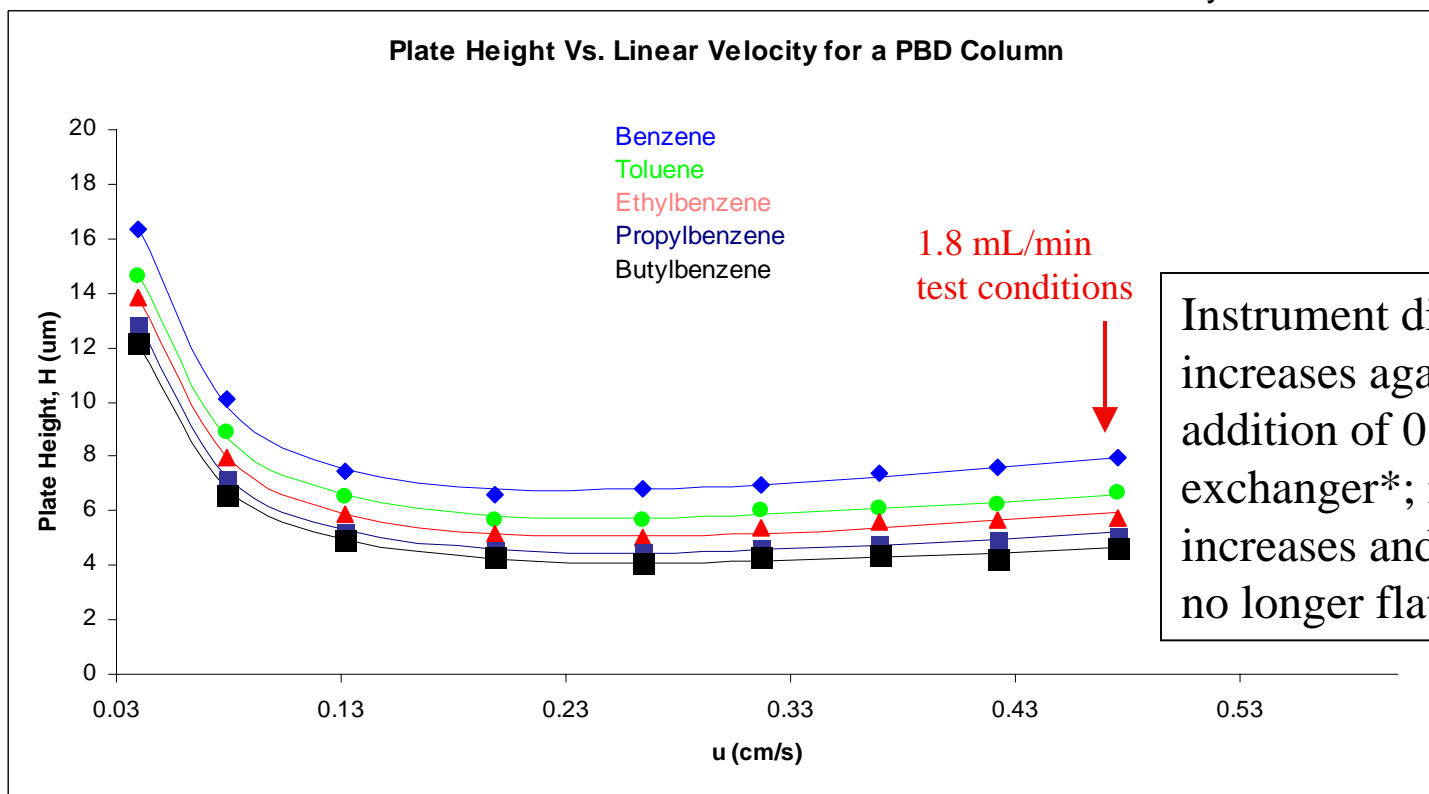
New family of curves with H_{\min} ranging from $4\mu\text{m}$ to $6\mu\text{m}$ indicates very low instrument impact on H_{system}



Instrument dispersion is almost negligible; curves nearly overlap and become flat.

Factory Instrument with Micro Cell + 0.005" ID Tubing + Heat Exchanger

New family of curves with H_{\min} ranging from $<5\mu\text{m}$ to $>7\mu\text{m}$ indicates moderate instrument impact on H_{system}



Instrument dispersion increases again due to addition of 0.007" heat exchanger*; range of H increases and curves are no longer flat.

* An Agilent Model 1200 heat exchanger may be surface mounted to the heater block to maintain low dispersion for heated applications.

Efficiency Plot for Agilent 1100 with Micro-Cell and Factory Tubing

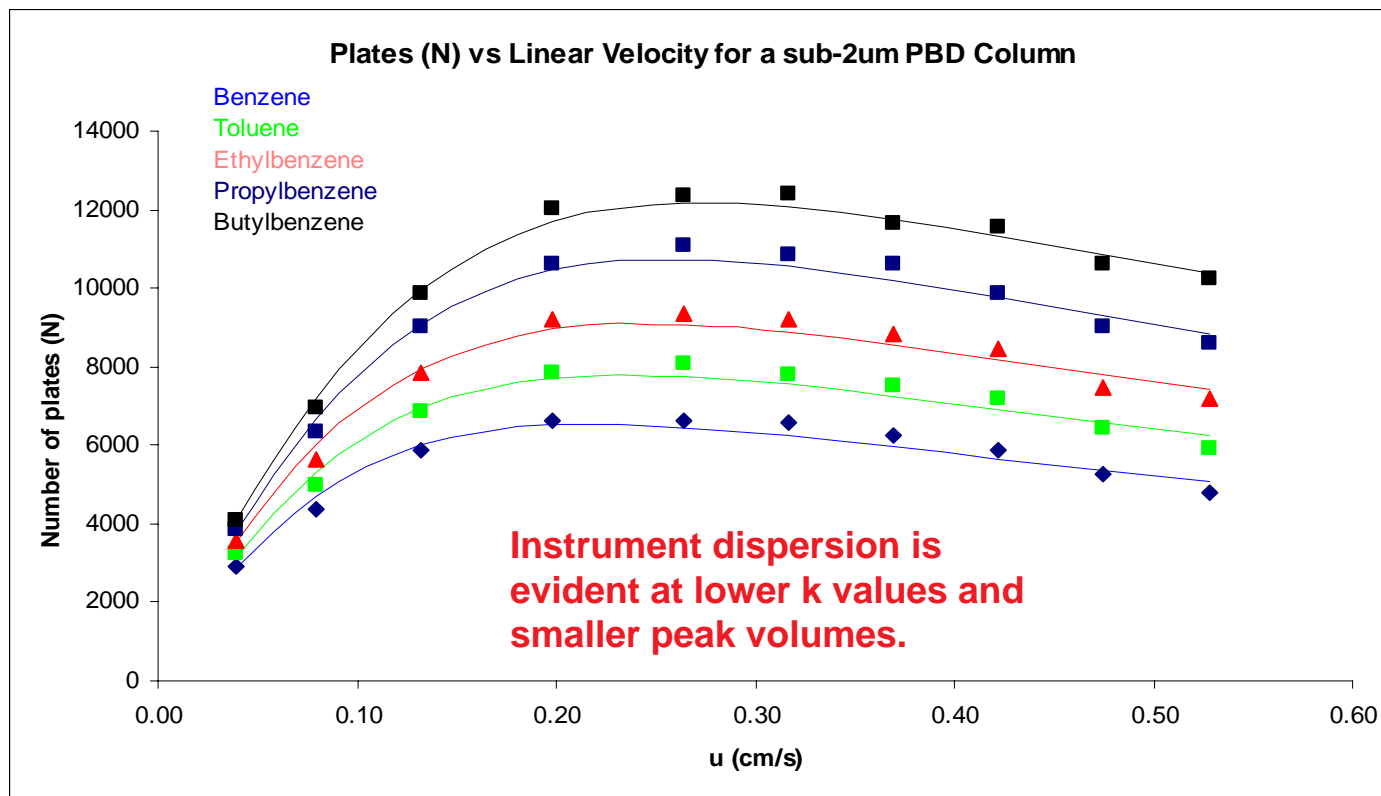


Plate height vs linear velocity for retained solutes: Alkylbenzenes, Temperature 30 °C, Mobile phase: 50/50 ACN/water; Column: 50 x 4.6mm, Agilent 1100/UV with Micro Cell and 0.007" i.d. tubing.

Beta-Blockers on Zr-PBD Sub-2 μ m at 75 °C

Analytes

- 1=Labetalol
- 2=Atenolol
- 3=Acebutolol
- 4=Metoprolol
- 5=Oxprenolol
- 6=Lidocaine
- 7=Quinidine
- 8=Alprenolol
- 9=Propranolol

Zr-PBD

50mm x 4.6mm, sub-2 μ m

22/78 ACN/20mM K₃PO₄ at pH=12

F=2.5 mL/min

UV=254nm

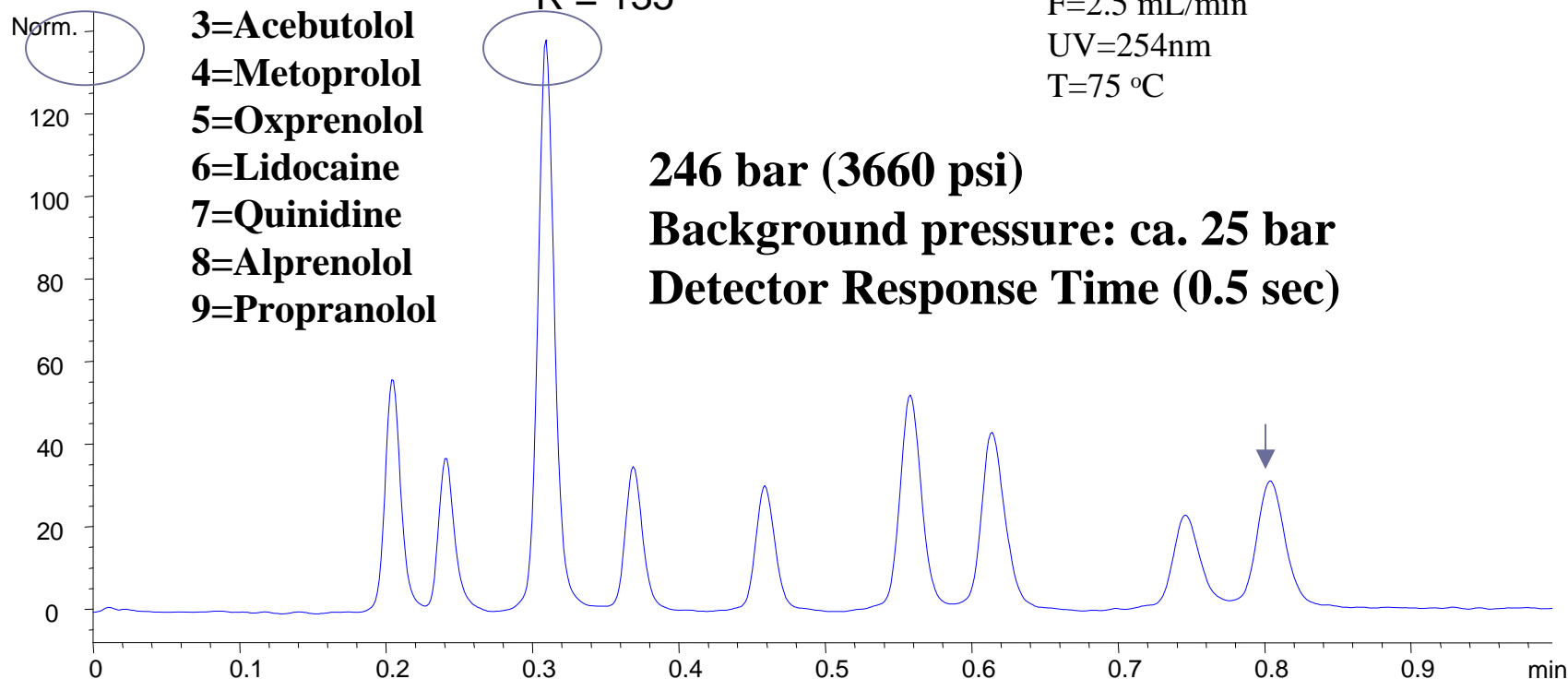
T=75 °C

R = 135

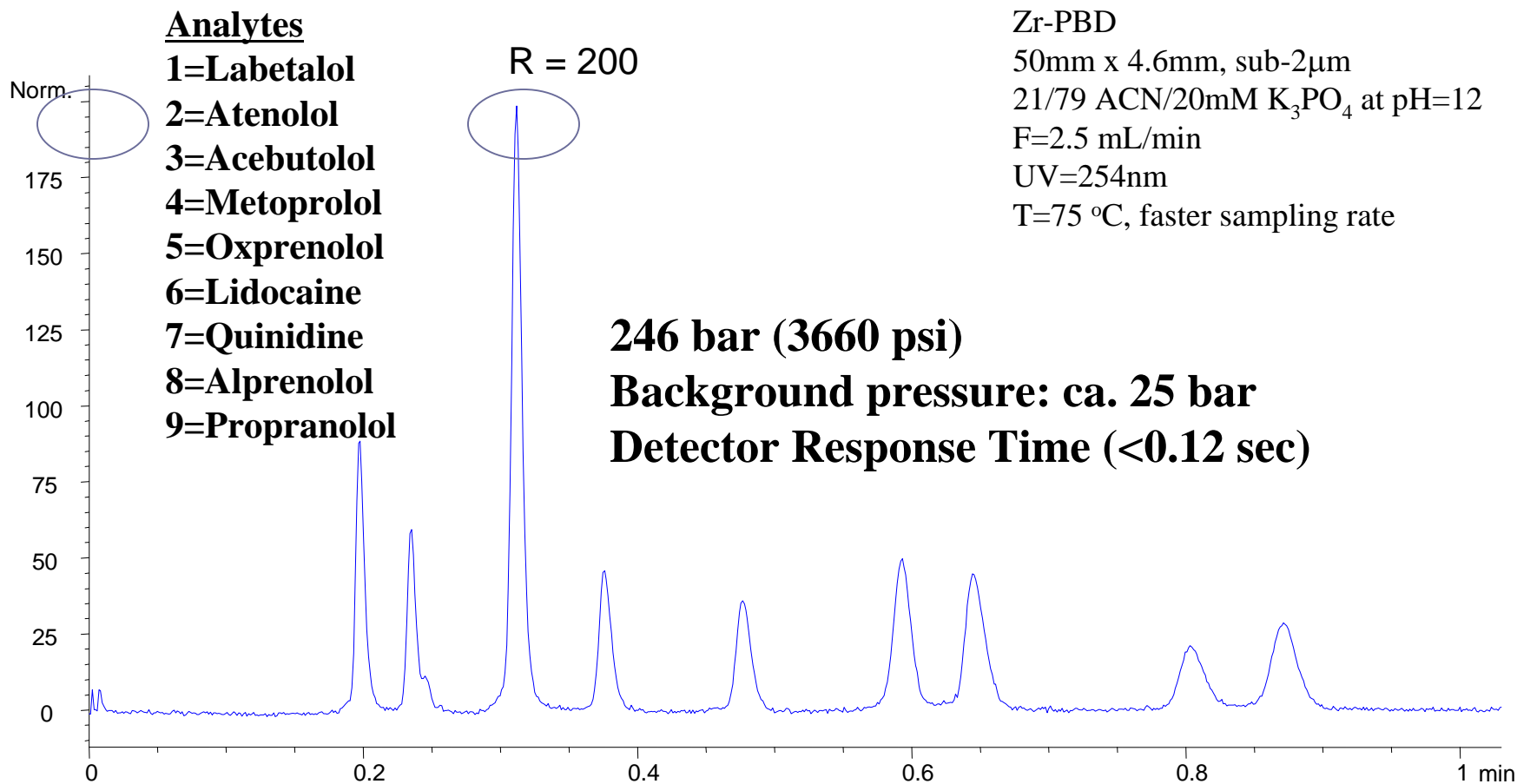
246 bar (3660 psi)

Background pressure: ca. 25 bar

Detector Response Time (0.5 sec)



Beta-Blockers Optimized with Faster Detector Response



Importance of Injection Volume and Gradient Focusing



Performance of Some Factory HPLC Instruments

Column Plates Determined from Toluene ($k = 2$) Half-Height Width
 Isocratic Elution with 60% ACN
 10 μ L Injected in Mobile Phase
 (All instruments in standard configuration with analytical flowcells)

Column	Agilent 1100	Agilent 1200	Waters 2695	Column Supplier Test Data
Ascentis C18 150x4.6mm 5 μ m	16911 (6.7% loss)	18034 (0% loss)	16874 (6.9% loss)	18119
Ascentis Express (Fused-Core) C18 100x4.6mm 2.7 μ m	18649 (34% loss)	22001 (22% loss)	18666 (34% loss)	28164

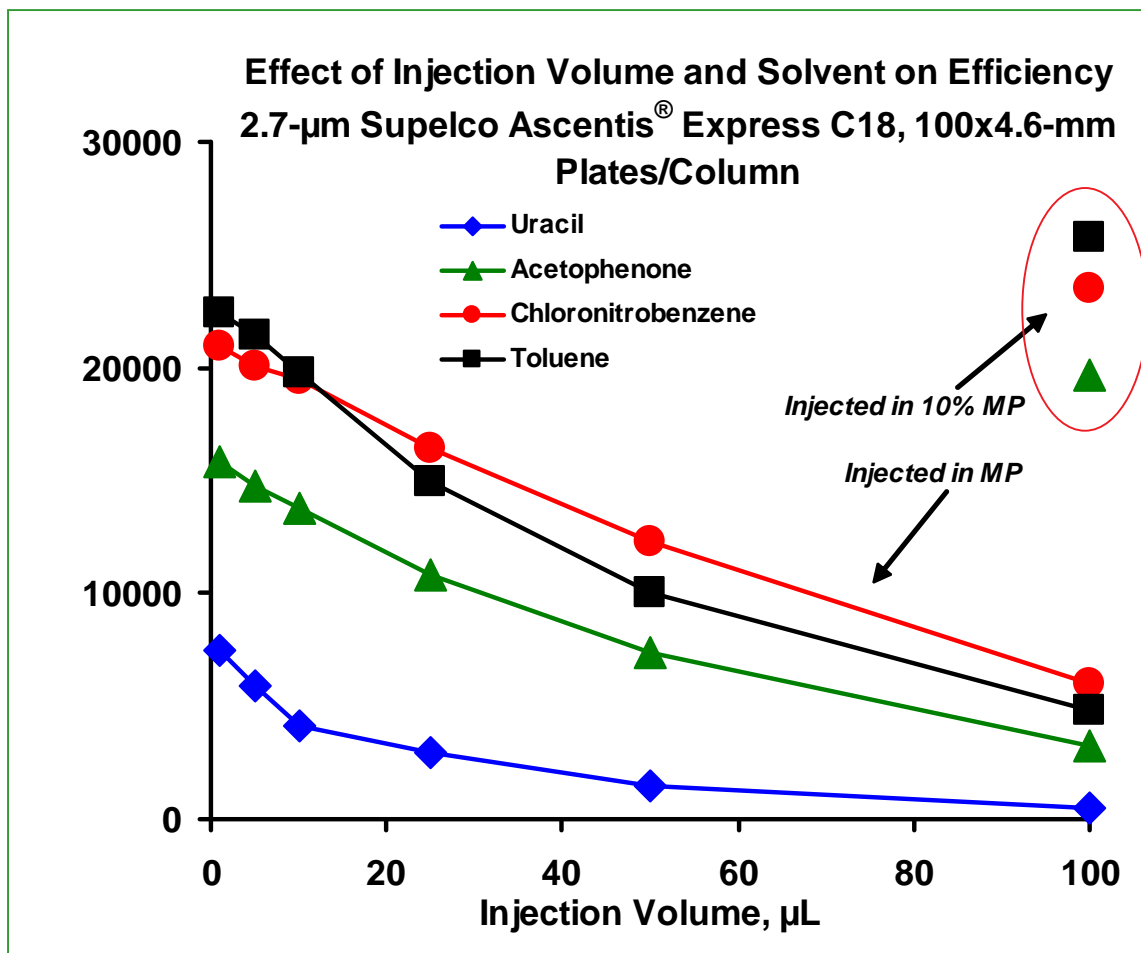
Effect of Variables Using Agilent 1200*

Summary of Toluene Plates from 2.7- μ m Supelco Ascentis[®] Express FCP Column Isocratic Elution with 60% ACN on Agilent 1200 10 μ L Injected in Mobile Phase*

Non-Optimized (10 mm / 13 μ L flowcell)	22001
Change to 6 mm / 5 μ L flowcell	21912
Minimize tubing lengths (don't bother)	22104
*10x sample dilution with 10x volume increase	25738

S. Bannister studies related to ref. 7 (Xcelience Labs, Tampa, FL).

Dispersion Caused by Injections in Mobile Phase



- Sample was diluted 1:10 with water for weak solvent injection.
- Note that efficiency performance was greater with a 100 μL injection that has been diluted with water than the original 1 μL injection in mobile phase.

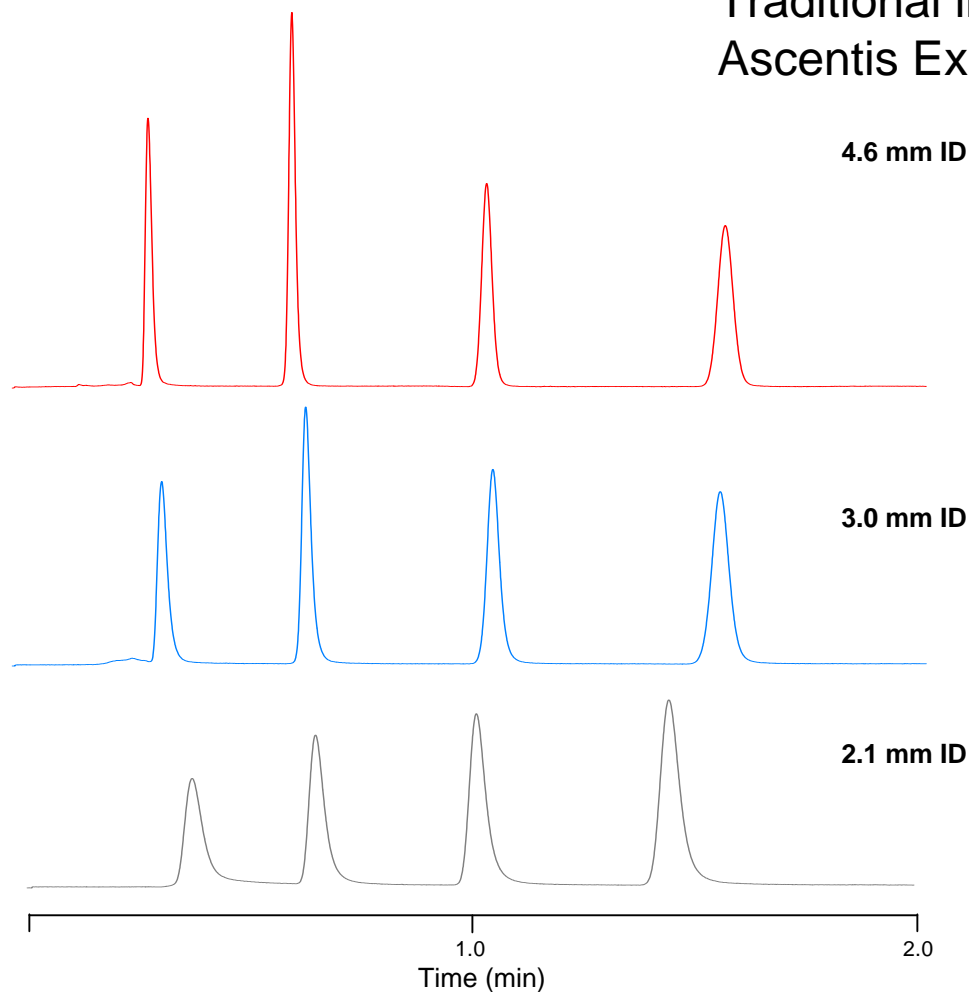
Estimating Instrument Bandwidth



Simple System Suitability (Bandwidth) Test

...one instrument with three columns (three systems)

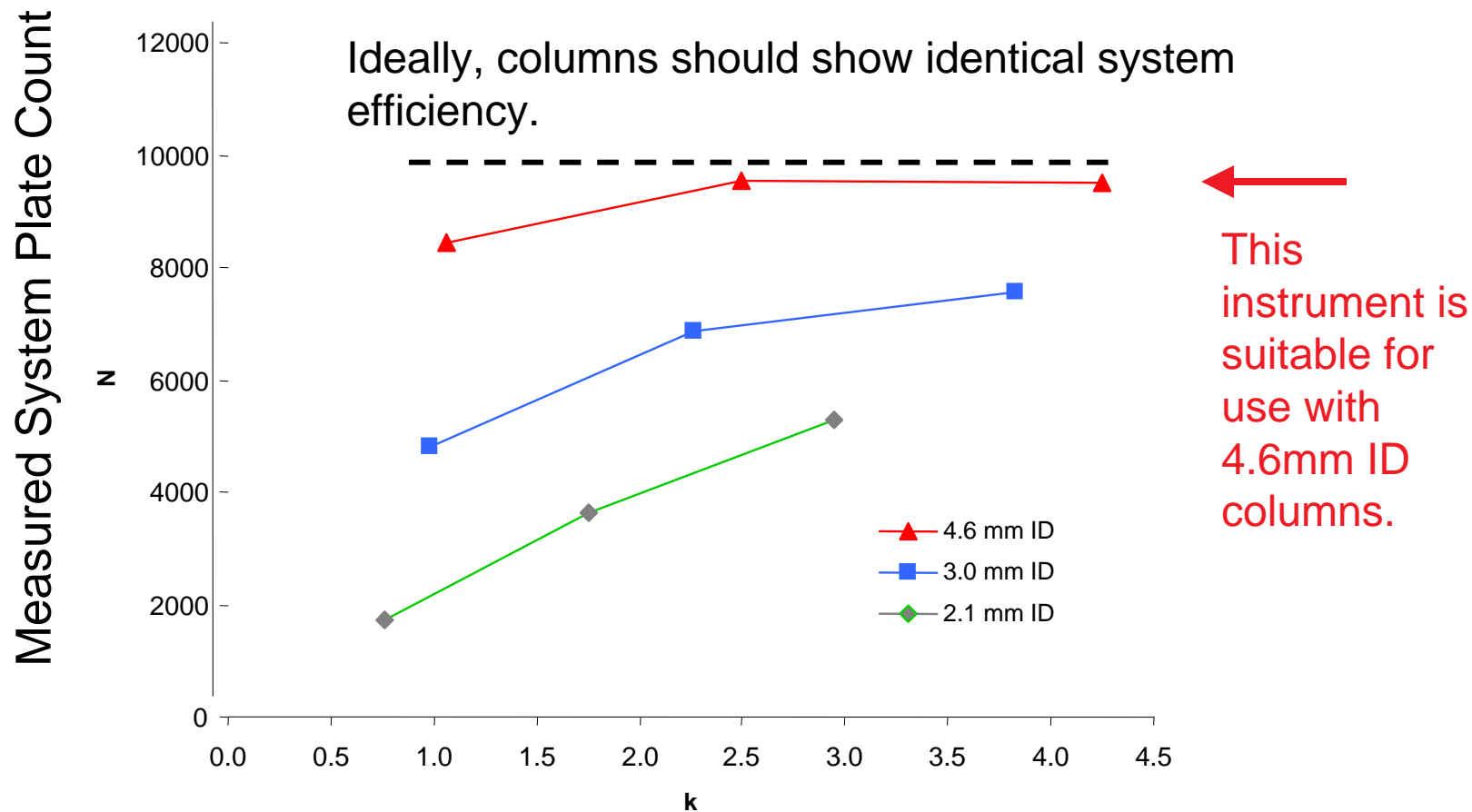
Traditional instrument with 5 cm
Ascentis Express columns



Choose test columns that
can show $>200,000$ N/m
at $k = 2-3$.

4.6mm ID columns
usually look pretty good
due to large void volume
and peak volumes.

Ranking System Performance with Three Columns

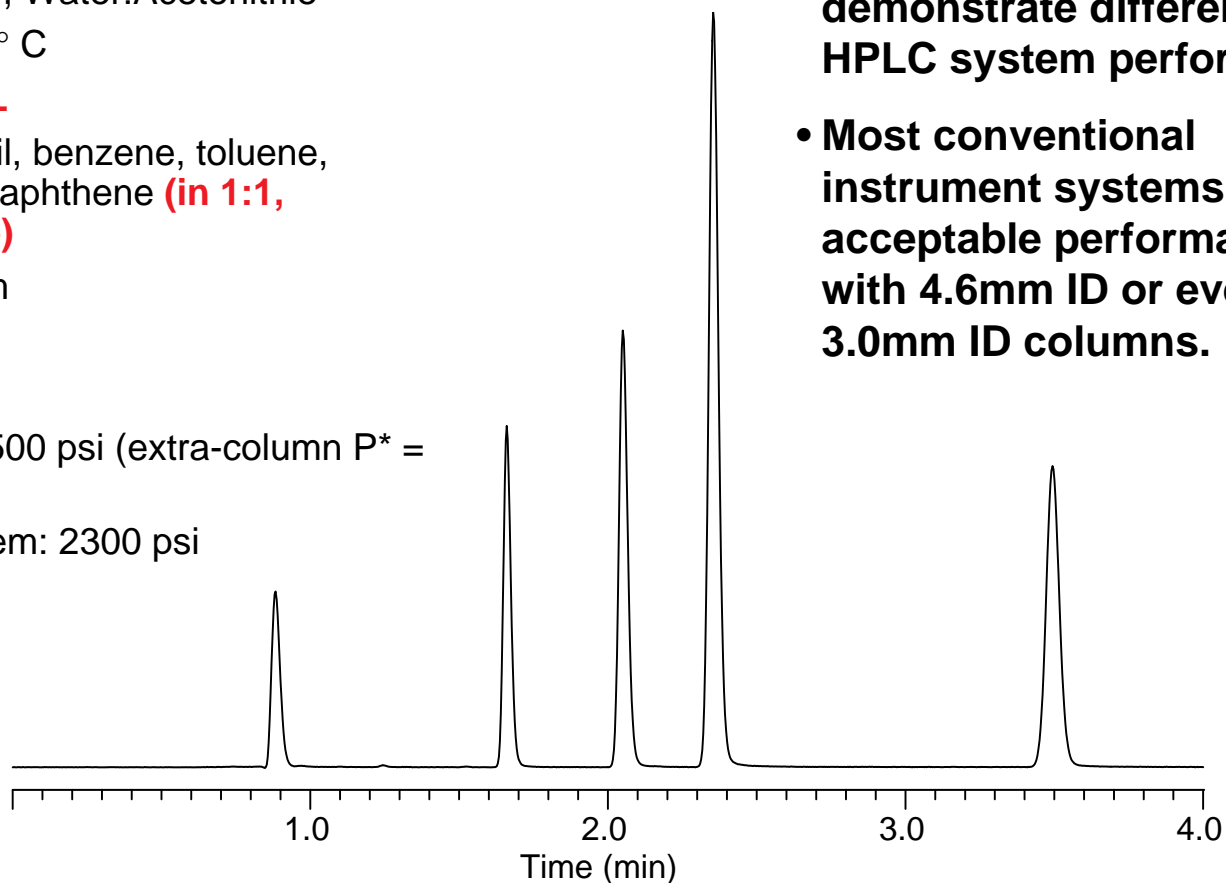


Another System Suitability (Bandwidth) Test

...one column with three instruments (three systems)

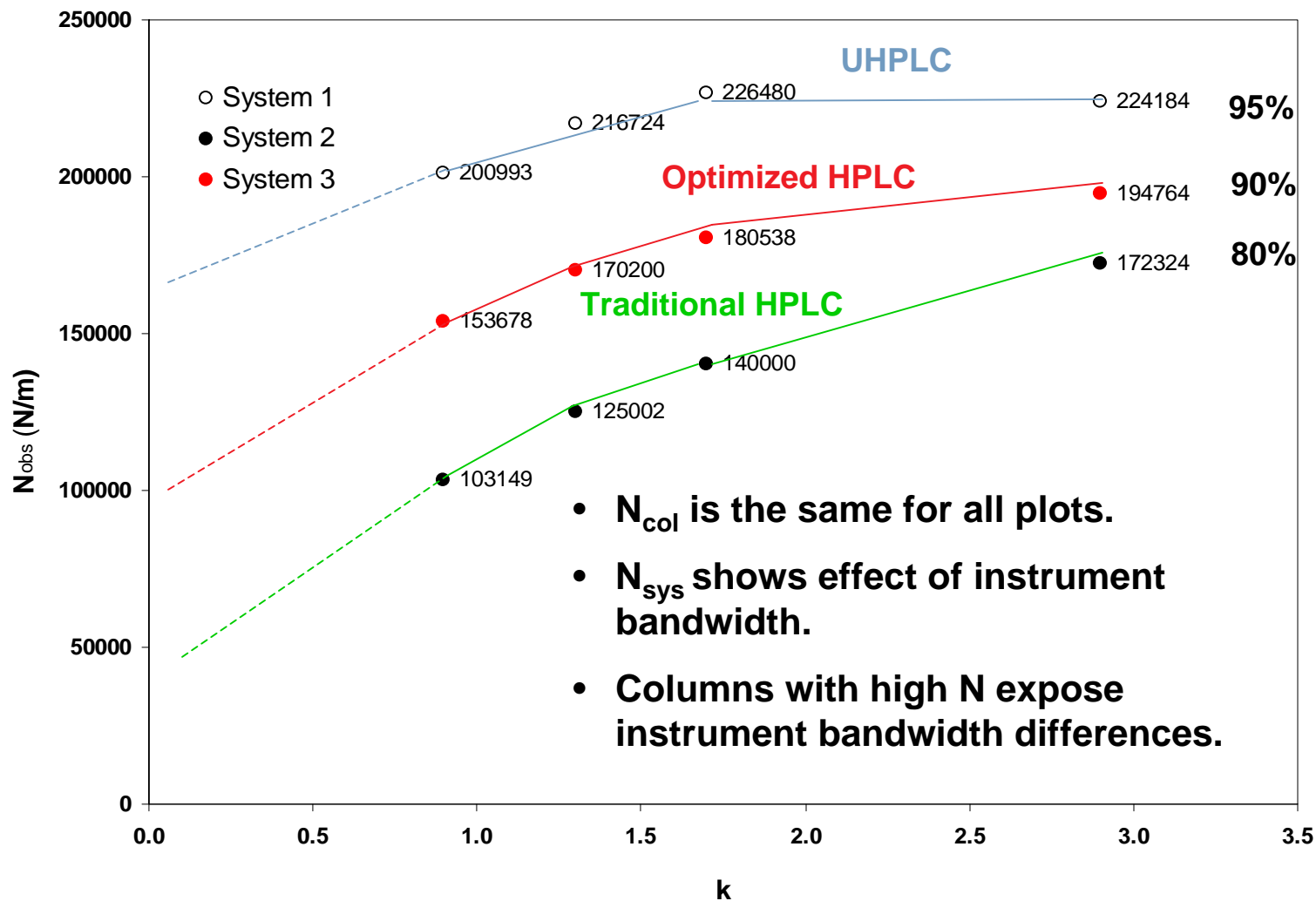
- **Column:** Ascentis Express C18, 2.1 x 150 mm
- **Mobile phase:** 3:7, Water:Acetonitrile
- **Column temp:** 35° C
- **Injection vol:** 1 μ L
- **Sample mix:** uracil, benzene, toluene, naphthalene, acenaphthene (in 1:1, water:acetonitrile)
- **Detection:** 250 nm
- **Flow:** 0.3 mL/min
- **Pressure:**
 - Ultra System: 3500 psi (extra-column P* = 1200 psi)
 - Traditional System: 2300 psi

- 2.1mm ID columns were used in the study to demonstrate differences in HPLC system performance.
- Most conventional instrument systems show acceptable performance with 4.6mm ID or even 3.0mm ID columns.



Ranking System Suitability Performance (Bandwidth)

The blue instrument is suitable (maybe the red one too)



Direct Method for Measuring IBW

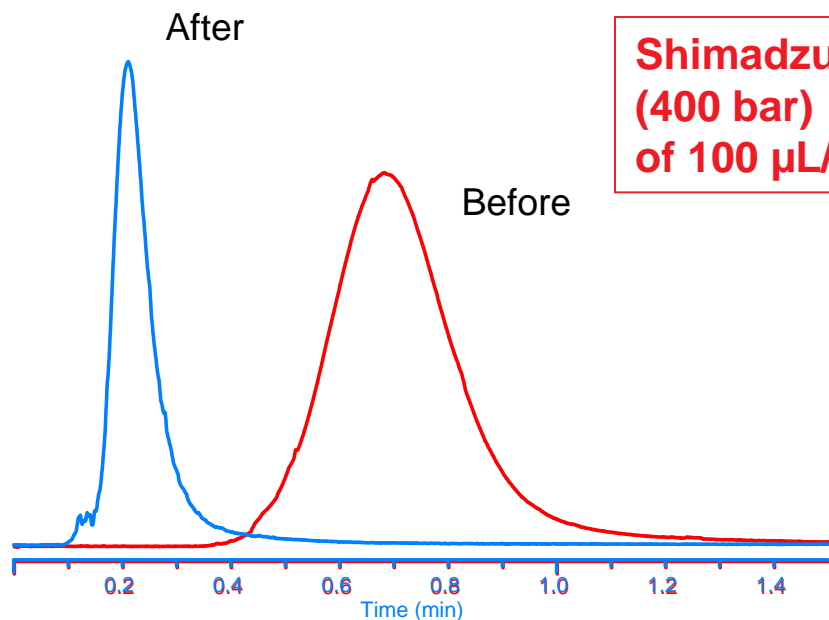
- Connect injector to detector
 - ZDV union
 - shunt
- Inject small volume (μL or less) of chromophore
- Record peak (or retention time and N)
- Calculate IBW (flow in μL) or measure directly from peak retention

$$\sigma = (t_r \times \text{flow}) / \sqrt{N}$$

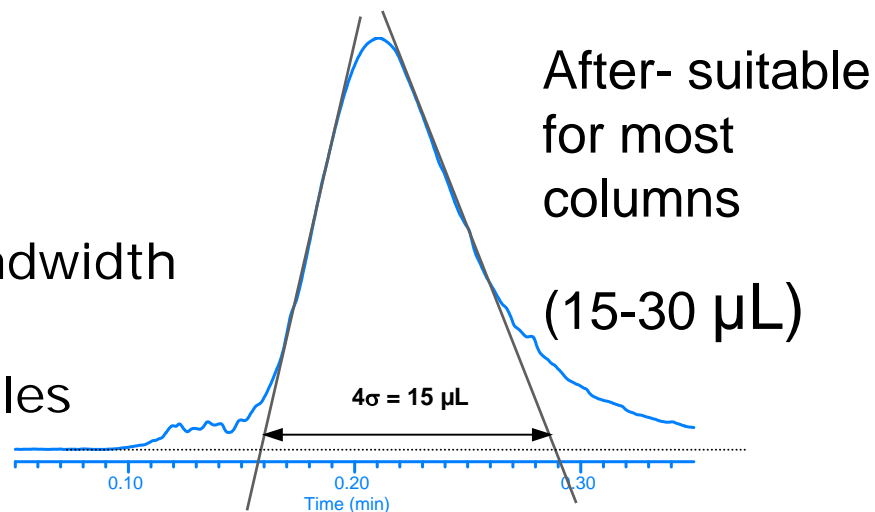
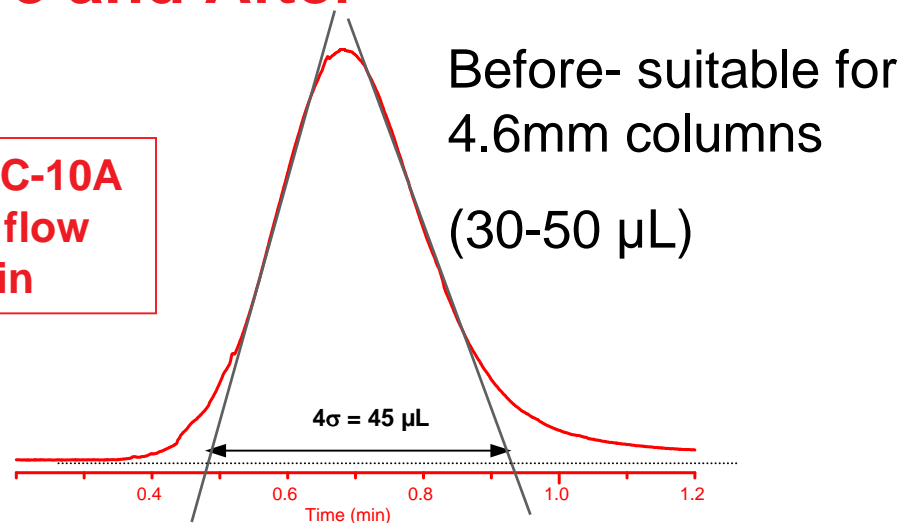
$$\text{IBW} = 4\sigma$$

- Common mistakes
 - data sampling rate too slow
 - detector response time too slow
 - flow rate too fast (or variable)
 - calculation of N

Comparison of IBW Before and After Instrument Optimization

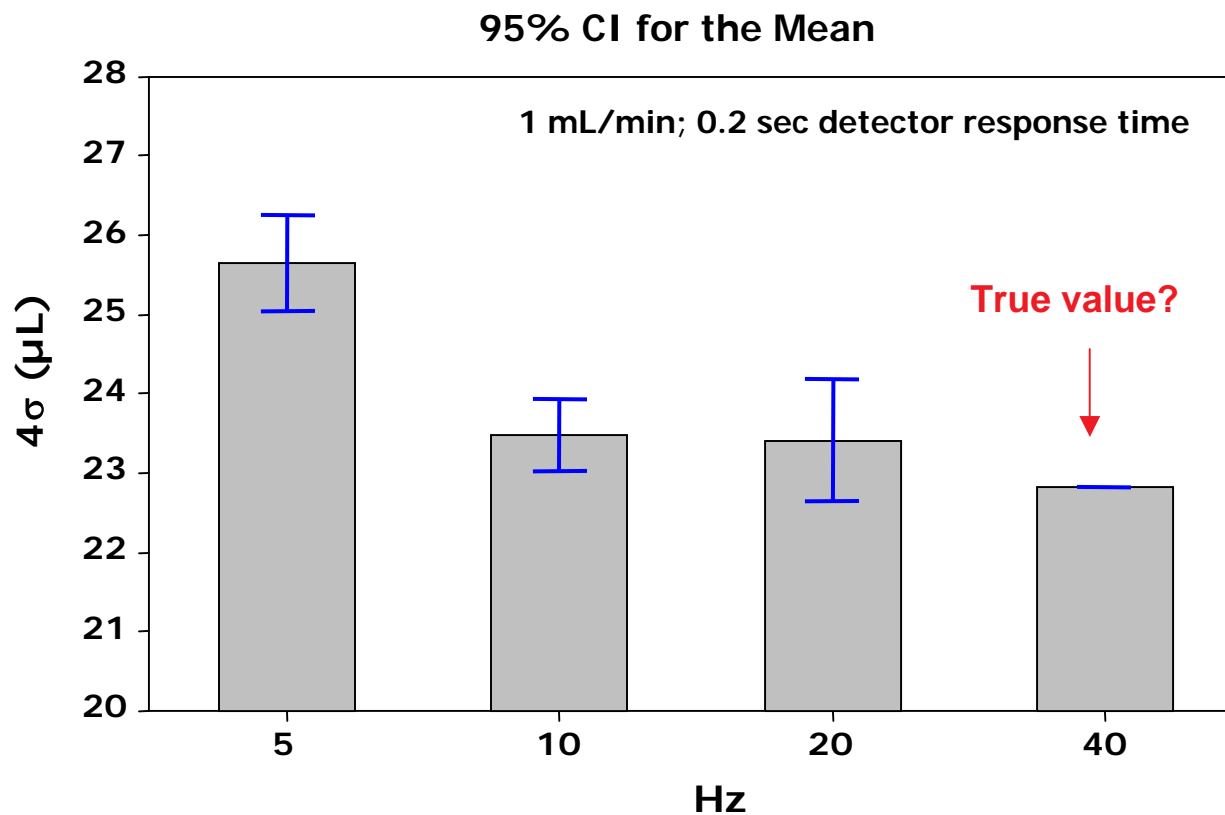


Shimadzu LC-10A
(400 bar) @ flow
of 100 $\mu\text{L}/\text{min}$

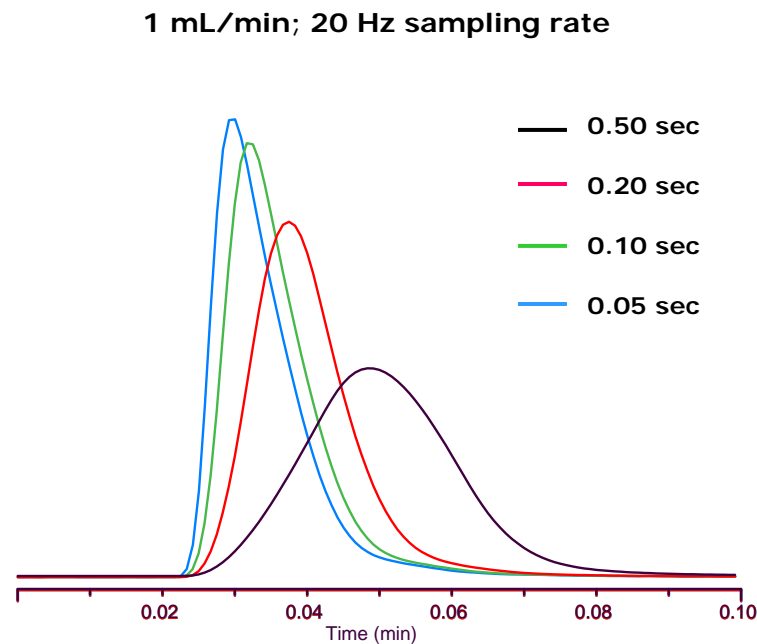
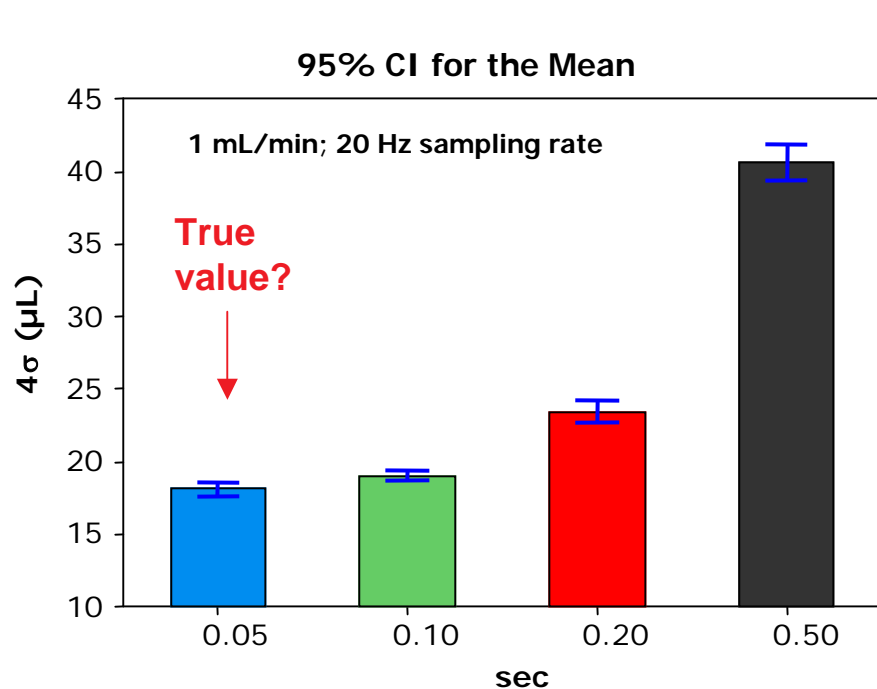


- Remove column and measure bandwidth at base for small volume injection
- Clear contrast between peak profiles before and after optimization

Effect of Data Sampling Rate on Measured IBW

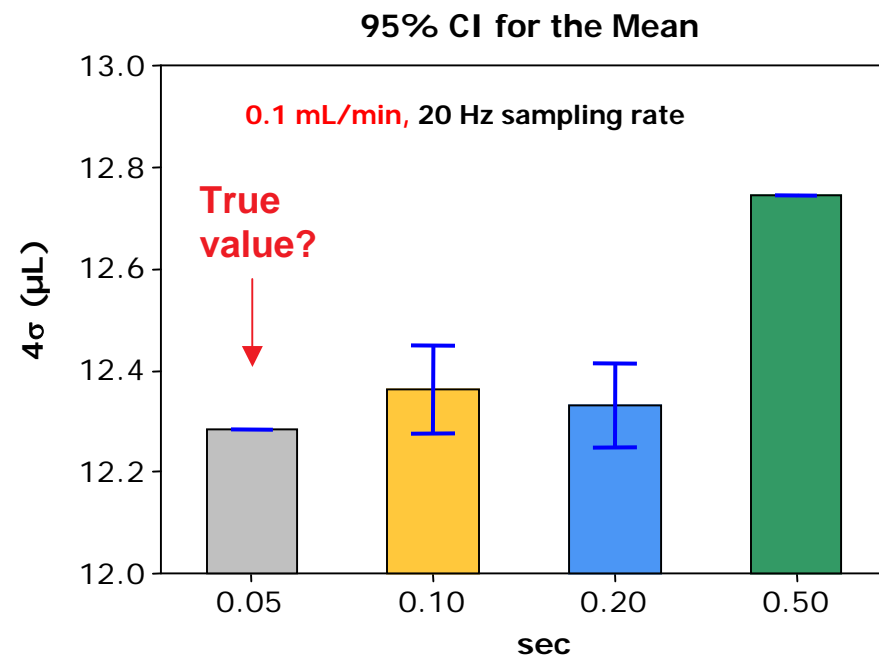
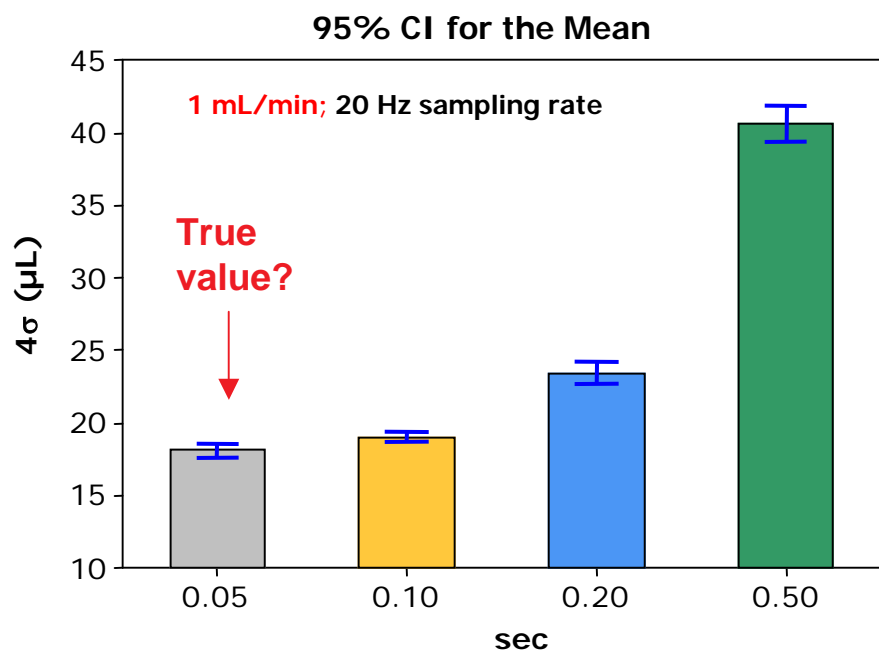


Effect of Detector Response Time on Measured IBW



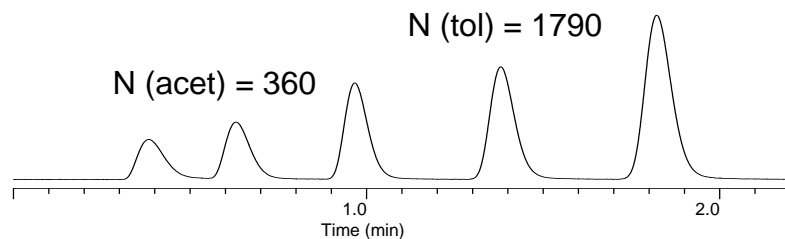
- Slow filter response time adversely effects accurate capture of peak dimensions by data system

Effect of Flow rate on Measured IBW



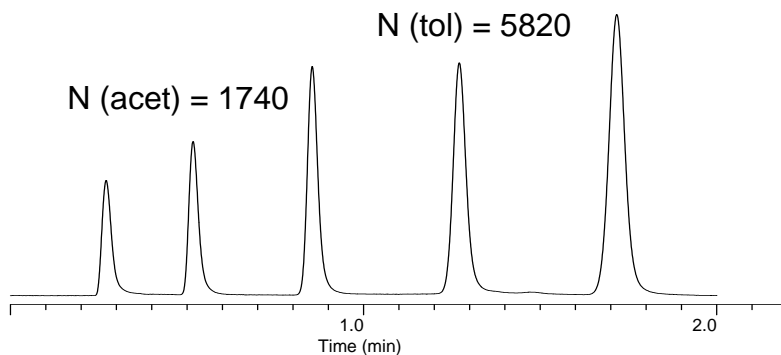
- Flow rate of 0.1 mL/min will inherently yield less dispersion, but also permits accurate capture of peak dimensions well within capabilities of data system

2.1 mm ID x 5 cm, 0.4 mL/min

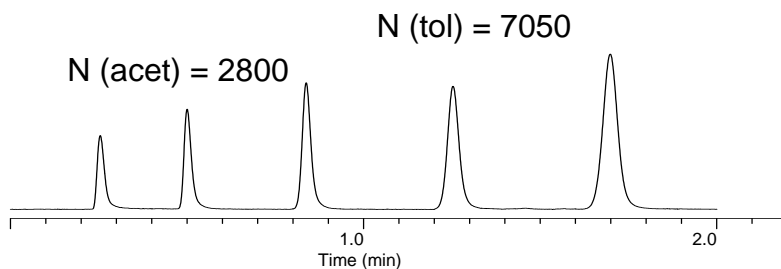


IBW: 42 μ L
Tubing ID: 0.010"
Flow cell: 16 μ L, 10 mm path
Pressure: 1560 psi

True N = >10,000

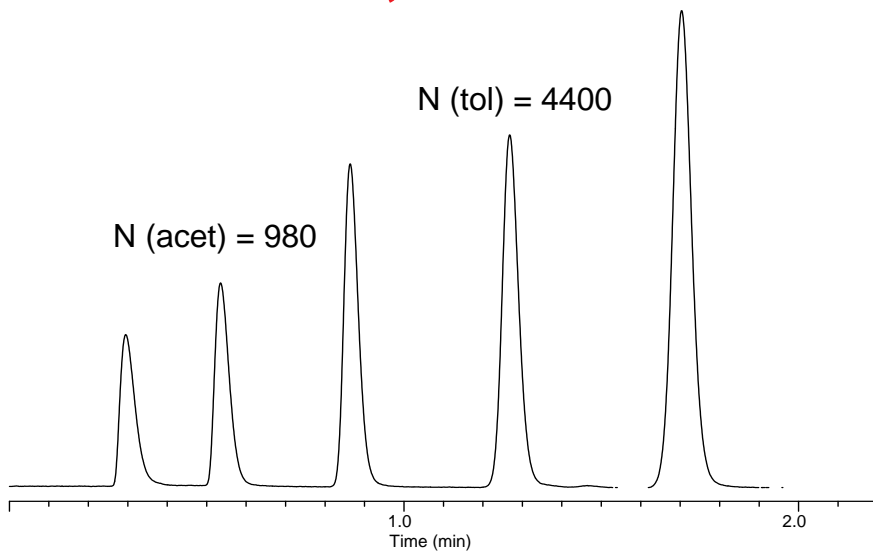


IBW: 17 μ L
Tubing ID: 0.005"
Flow cell: 16 μ L, 10 mm path
Pressure: 1680 psi



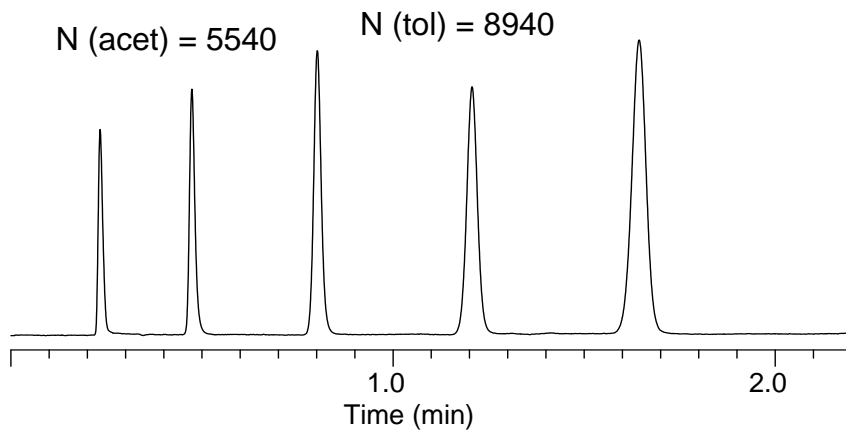
IBW: 12 μ L
Tubing ID: 0.005"
Flow cell: 2.5 μ L, 5 mm path
Pressure: 1680 psi

3 mm ID x 5 cm, 0.8 mL/min



IBW: 42 μL
Tubing ID: 0.010"
Flow cell: 16 μL , 10 mm path
Pressure: 1600 psi

True N = >10,000



IBW: 12 μL
Tubing ID: 0.005"
Flow cell: 2.5 μL , 5 mm path
Pressure: 1640 psi

Fekete Direct Measurements of IBW⁴

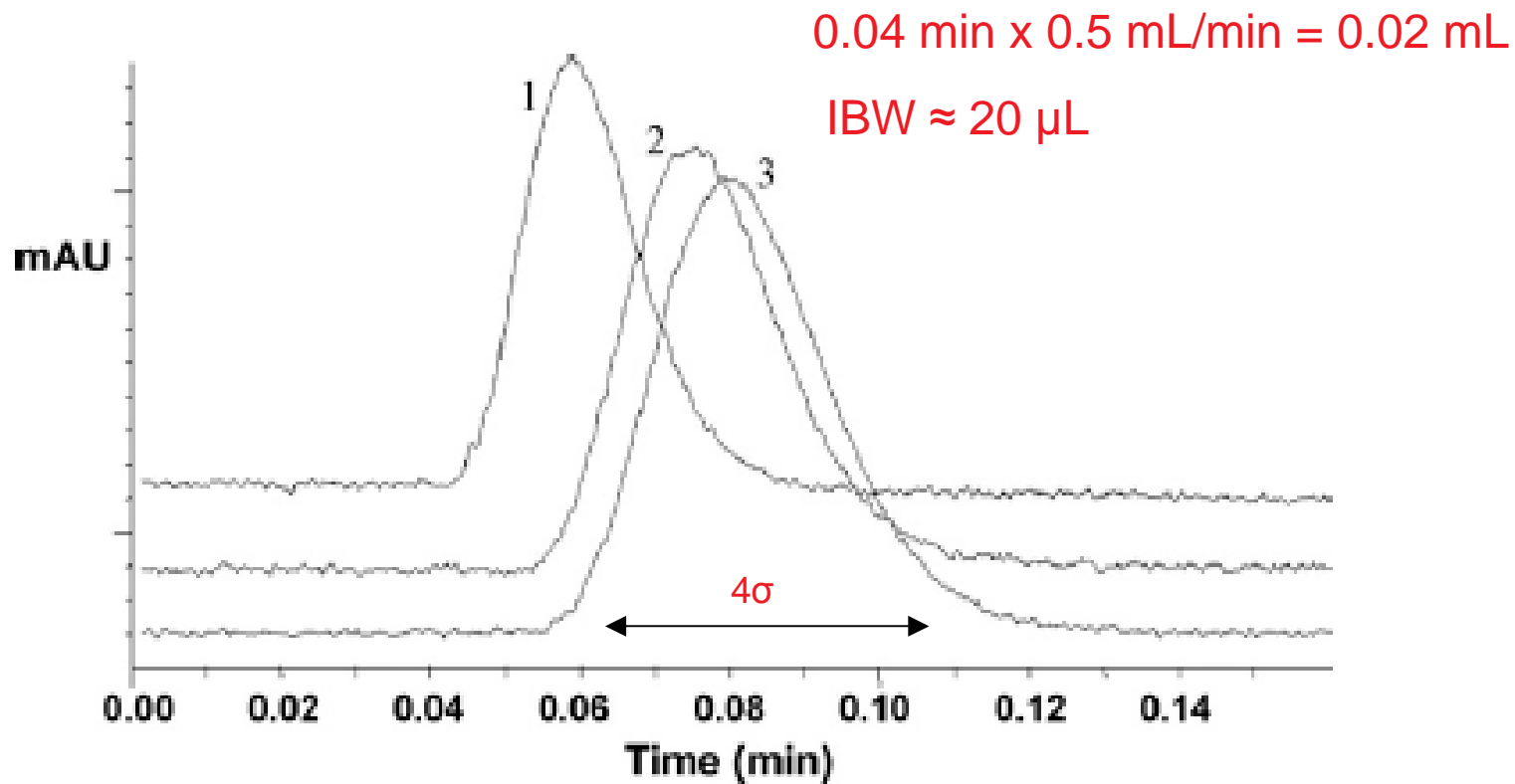


Fig. 9. Extra-column band profiles of the different liquid chromatographic systems. 1: Waters Acquity UPLC; 2: Agilent 1200 RRLC; 3: Shimadzu Prominence UFLC.

Summary of Variables that Affect Instrument Bandwidth and System Suitability

Instrument volume should be small with respect to column internal volume which determines peak volume at base.

- Reduce tubing ID and **volume**
- Reduce tubing length and **volume**
- Reduce detector flow cell **volume**
- Improve detector response **time**
- Match data collection rate to peak width in **time**
 - At least 20-30 points across the peak

Dispersion References

1. R. Majors, *Are you Getting the Most Out of Your HPLC Column?*, LCGC NA, Vol. 21, No. 12, 1124-1133 (December 2003).
2. R. A. Henry and D. S. Bell, *Important Guidelines for Optimizing Speed and Sensitivity in Small Molecule LC-UV and LC-MS*, LCGC NA, Vol. 23, No. 5, 2-7 (May 2005).
3. T. Chester, *Sub-2 μ m Performance with a Conventional Instrument Using 2.7 μ m Fused-CoreTM Particles*, American Lab, Volume 41, No. 4, 11-15 (2009).
4. S. Fekete, et. al., *Shell and Small Particles: Evaluation of New Technology*, J. of Pharmaceutical and Biomedical Analysis, 49, 64-71 (2009).
5. F. Gritti, G. Guiochon, et. al., *Achieving Full Performance of Columns by Optimizing HPLC Instruments*, J. of Chromatogr. A, 1217, 3000-3012 (2010)
6. H. Brandes, unpublished data, Sigma-Supelco Applications Lab Reports and Notebooks (2008-2009).
7. R. A. Henry and D. Nowlan, *Use of Sub-2 μ m Zirconia-PBD at Elevated pH and Temperature*, Oral Paper, EAS 2009, Somerset, NJ.

Thanks for your attention!



R. A. Henry 2010

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