

# Transfer and Speed-up of Methods to Fused-Core Particle Columns – EPA Method 1694

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# Abstract

EPA Method 1694 is used in the determination of pharmaceuticals and personal care products (PPCPs) in environmental samples. These PPCPs have received considerable press attention in recent years, with growing concerns over the presence of these compounds in the world's drinking waters. This method uses solid phase extraction (SPE) and high performance liquid chromatography combined with tandem mass spectrometry (LC-MS-MS) to quantitate more than 70 PPCPs in a variety of environmental matrices. In this work, the Group 1 compounds of EPA Method 1694 were run on a column containing Fused-Core™ particles.

HPLC columns packed with Fused-Core particles or porous particles in the sub-2  $\mu\text{m}$  range can both enhance resolution and speed by producing either higher efficiency (N) for the same column length or equivalent efficiency with a shorter column length. The use of ultra-high performance liquid chromatography (UHPLC) comes at a price of higher pressure when sub-2  $\mu\text{m}$  particles are employed.

## Abstract (contd.)

As an alternative, Fused-Core silica particles have gained acceptance because of the equivalent performance to particles in the sub-2  $\mu\text{m}$  range. These Fused-Core columns employ conventional 2  $\mu\text{m}$  frits and operate ruggedly at much lower pressures that are within the operating limits of conventional HPLC instruments. In this work, Group 1 compounds are shown to exhibit good resolution with MS-MS detection indicating these Fused-Core columns may be used in this EPA Method.

# Experimental

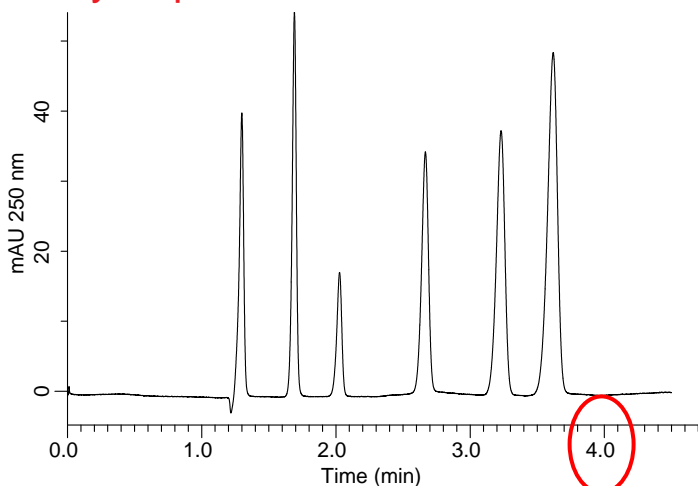
## Example 1. Hydrophobic Test Mix

Initially, separation on a 5  $\mu\text{m}$  C18 column is scaled to Ascentis<sup>®</sup> Express (FC column) to provide equivalent efficiency and retention for latest eluting peak, naphthalene. This involves changes in column length, mobile phase, and flow.

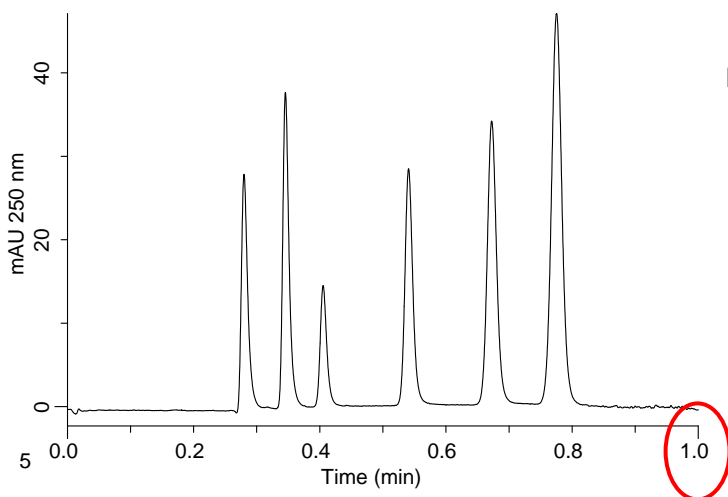
Runtime can be further shortened while still maintaining baseline resolution of critical pair.

# Scaling Method to Fast LC, Moderate Pressure

## Hydrophobic Test Mix



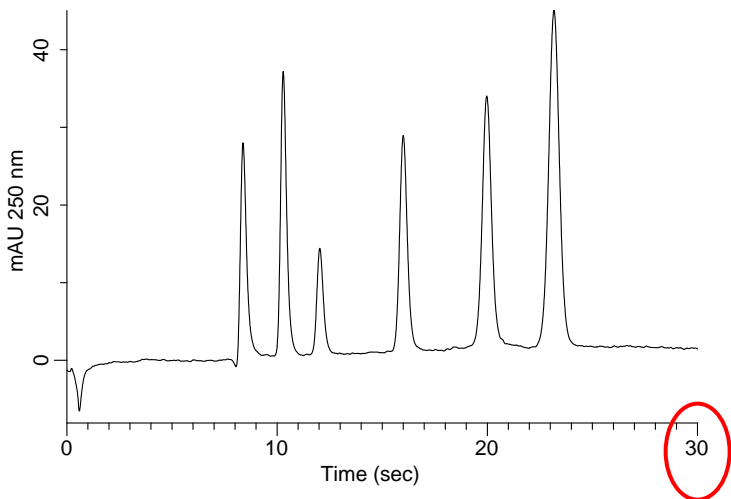
column: C18, 15 cm x 3 mm I.D., 5  $\mu$ m  
mobile phase: 20:80, water:acetonitrile  
flow rate: 0.4 mL/min.  
temp.: 35  $^{\circ}$ C  
injection: 1.5  $\mu$ L  
pressure: 885 psi (61 bar)  
 $N_{(\text{naphthalene})}$ : ~11000  
 $k_{(\text{naphthalene})}$ : 1.78  
elution order: uracil, phenol, acetophenone,  
benzene, toluene, naphthalene



column: Ascentis Express C18, 5 cm x 3 mm I.D., 2.7  $\mu$ m  
mobile phase: 31:69, water:acetonitrile  
flow rate: 0.6 mL/min.  
temp.: 35  $^{\circ}$ C  
injection: 0.5  $\mu$ L  
pressure: 1750 psi (121 bar)  
 $N_{(\text{naphthalene})}$ : ~11000  
 $k_{(\text{naphthalene})}$ : 1.75  
 $R_s_{(\text{acetophenone})}$ : 3.2

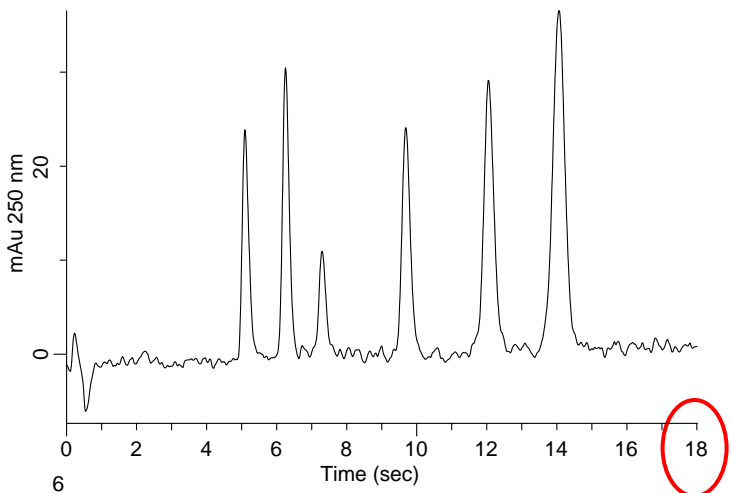
**At least a 4-fold increase in throughput.**

# How fast do you want to go? What system pressure limits exist?



column: Ascentis Express C18, 5 cm x 3 mm I.D., 2.7  $\mu\text{m}$   
mobile phase: 31:69, water:acetonitrile  
flow rate: 1.2 mL/min  
temp.: 35° C  
injection: 0.5  $\mu\text{L}$   
pressure: 3700 psi (255 bar)  
 $R_{s(\text{acetophenone})}$ : 3.1

**9-fold increase in throughput compared to the original method on 5  $\mu\text{m}$  particle.**



column: Ascentis Express C18, 5 cm x 3 mm I.D., 2.7  $\mu\text{m}$   
mobile phase: 31:69, water:acetonitrile  
flow rate: 2.0 mL/min  
temp.: 35° C  
injection: 0.5  $\mu\text{L}$   
pressure: 6400 psi (441 bar)  
 $R_{s(\text{acetophenone})}$ : 2.8

**15-fold increase in throughput compared to the original method on 5  $\mu\text{m}$  particle.**

# Experimental (contd.)

## Example 2. EPA Method 1694

Pharmaceuticals in the water supply represent a growing public concern, as represented by stories in many newspapers nationwide.



Current research at EPA includes development of testing methodologies for these compounds:

- Method 1694: Pharmaceutical and Personal Care Products in Water, Soil, Sediment, and Biosolids
- Method 1698: Steroids and Hormones in Water, Soil, Sediment, and Biosolids

# Experimental (contd.)

## LC-MS/MS Conditions

<b>Instrument</b>	Applied Biosystems 3200QT																																				
<b>Column</b>	Ascentis Express C18, 10 cm x 2.1 mm, I.D., 2.7 $\mu$ m																																				
<b>Mobile Phase</b>	(A): 0.1% formic acid and 0.1% ammonium formate in water; (B): 50:50 methanol:acetonitrile																																				
<b>Gradient and Flow</b>	<table border="1"><thead><tr><th>Time</th><th>Flow <math>\mu</math>L/min</th><th>%A</th><th>%B</th></tr></thead><tbody><tr><td>0.00</td><td>150</td><td>95.0</td><td>5.0</td></tr><tr><td>4.00</td><td>250</td><td>95.0</td><td>5.0</td></tr><tr><td>22.50</td><td>300</td><td>12.0</td><td>88.0</td></tr><tr><td>23.00</td><td>300</td><td>0.0</td><td>100.0</td></tr><tr><td>26.00</td><td>300</td><td>0.0</td><td>100.0</td></tr><tr><td>26.50</td><td>150</td><td>95.0</td><td>5.0</td></tr><tr><td>33.00</td><td>150</td><td>95.0</td><td>5.0</td></tr><tr><td>43.00</td><td>150</td><td>95.0</td><td>5.0</td></tr></tbody></table>	Time	Flow $\mu$ L/min	%A	%B	0.00	150	95.0	5.0	4.00	250	95.0	5.0	22.50	300	12.0	88.0	23.00	300	0.0	100.0	26.00	300	0.0	100.0	26.50	150	95.0	5.0	33.00	150	95.0	5.0	43.00	150	95.0	5.0
Time	Flow $\mu$ L/min	%A	%B																																		
0.00	150	95.0	5.0																																		
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23.00	300	0.0	100.0																																		
26.00	300	0.0	100.0																																		
26.50	150	95.0	5.0																																		
33.00	150	95.0	5.0																																		
43.00	150	95.0	5.0																																		
<b>Temperature</b>	40 ° C																																				
<b>Injection Volume</b>	5.0 $\mu$ L																																				
<b>Source Conditions</b>	Turbo ion spray ESI +, MRM																																				
<b>MS-MS Transitions</b>	See Table 1																																				
<b>Dwell time</b>	50 msec																																				

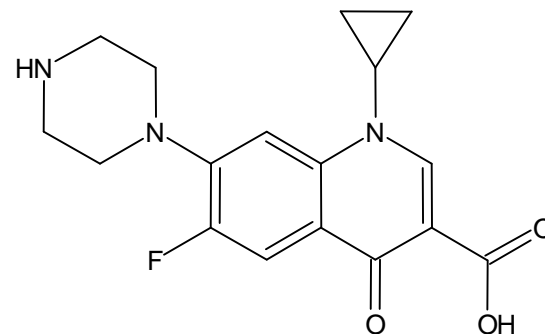
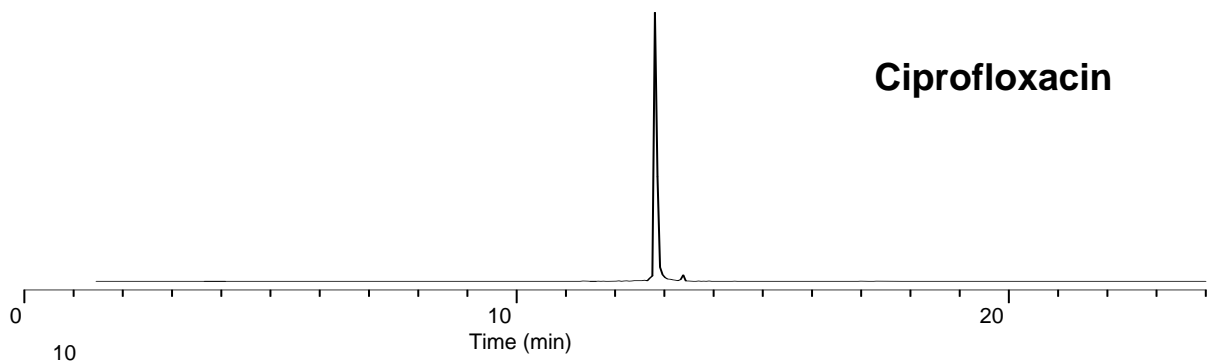
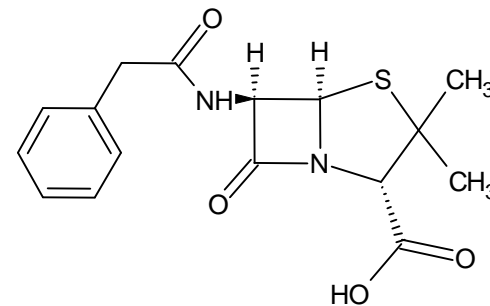
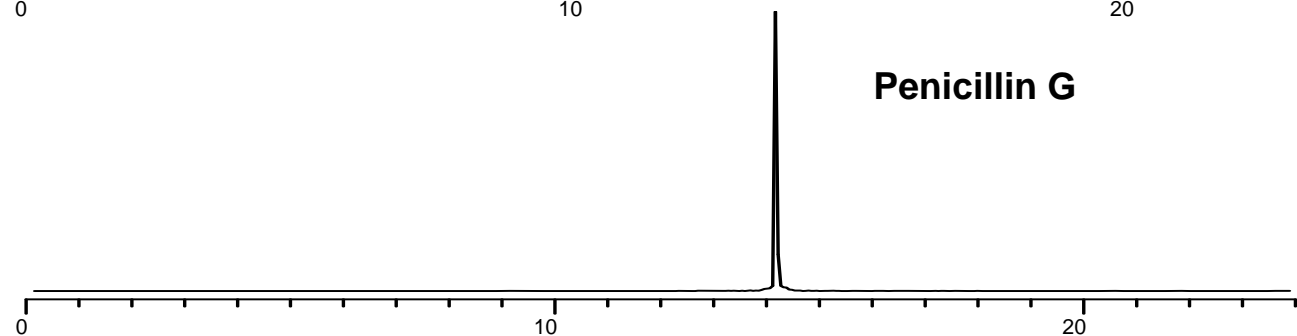
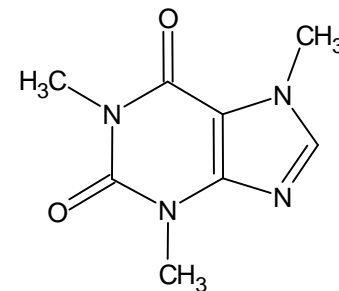
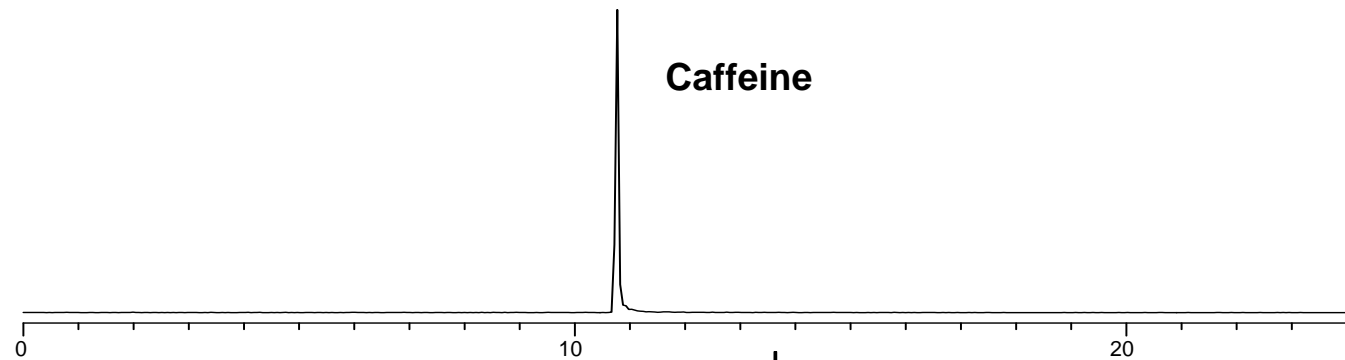


## Experimental (contd.)

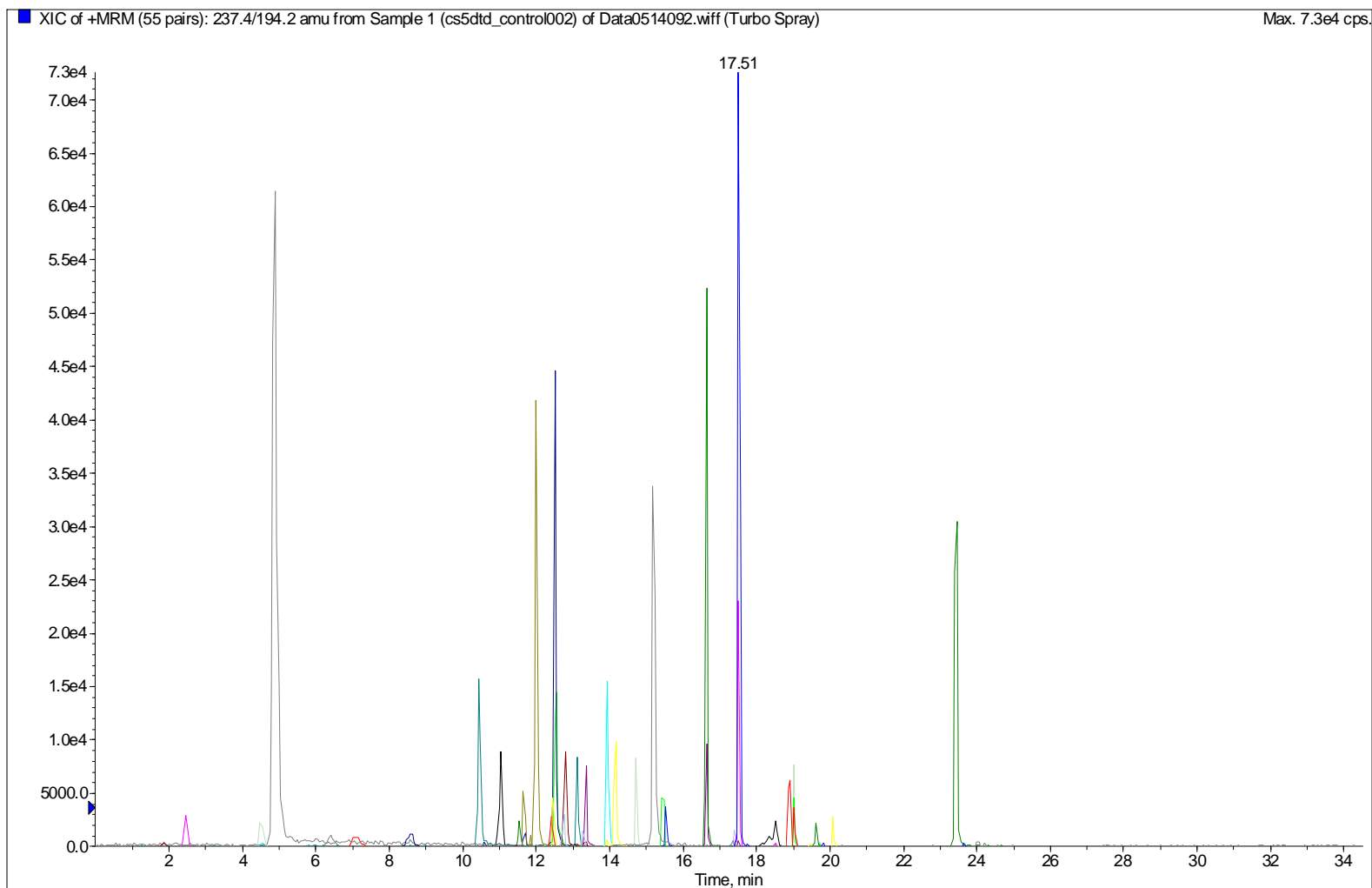
### SPE Extraction Method

<b>Extraction Cartridge</b>	Supelco Select HLB SPE Tube, 500 mg/6 mL
<b>Sample</b>	500 mL of drinking water spiked with Group 1 compounds, adjusted to pH=4 with 6M HCl
<b>Tube Conditioning</b>	20 mL of methanol, 6 mL of water, 6 mL of water at pH 2 (with 6 M HCl)
<b>Sample extraction</b>	10 mL/min through tube
<b>Dry time</b>	5 min
<b>Elution</b>	12 mL, 50:50 methanol:acetonitrile
<b>Dry down</b>	40 ° C, under nitrogen stream
<b>Reconstitution</b>	To final volume of 2 mL using mobile phase

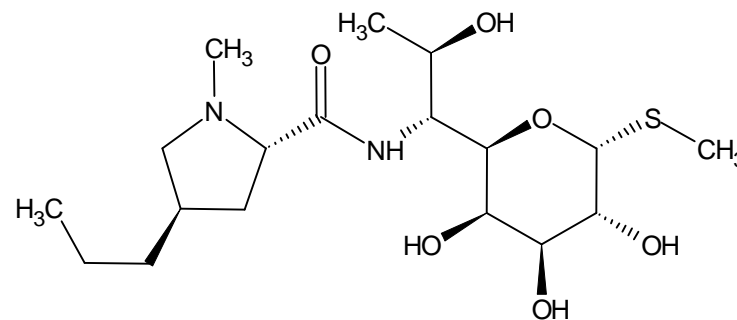
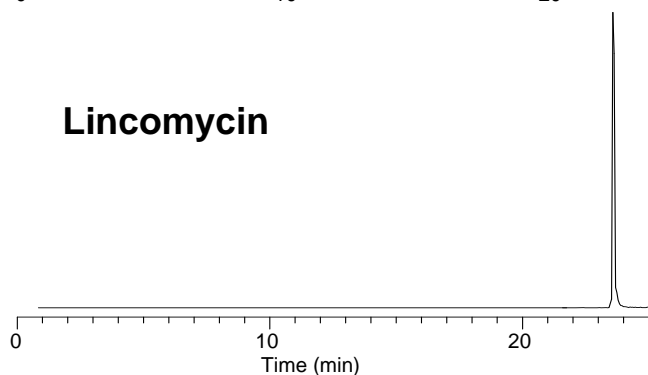
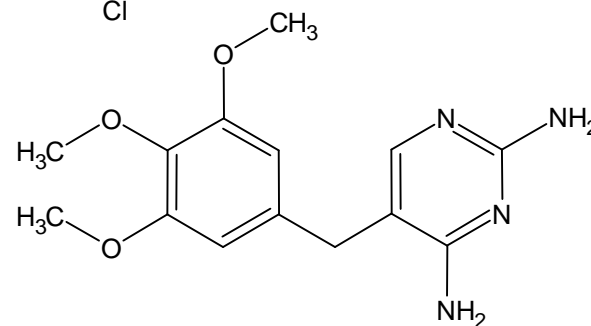
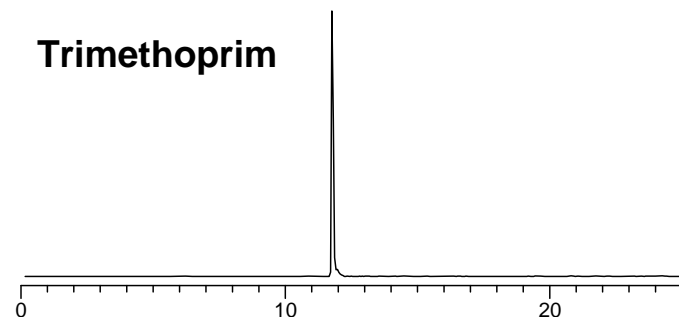
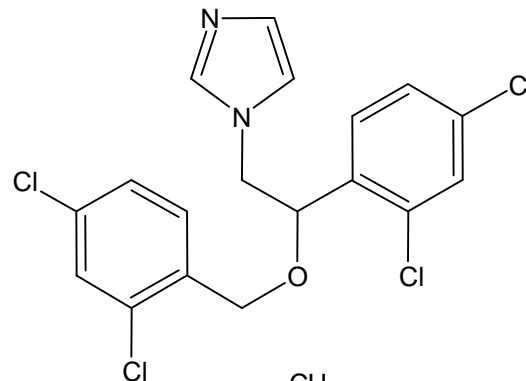
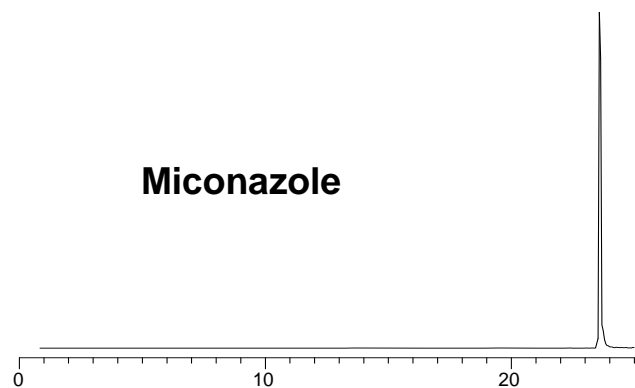
# Figure 2. EPA Method 1694 Select MRM Transitions from Figure 1



# Figure 1. LC-MS-MS Chromatogram of Group 1 Compounds



# Figure 2. EPA Method 1694 Select MRM Transitions from Figure 1 (contd.)



# Table 1. Group 1 Compounds from EPA Method 1694

<b>Compound</b>	<b>MRM Transition</b>
Acetaminophen	152.2-126.2
Ampicillin	350.3-106.1
Azithromycin	749.9-591.6
Caffeine	195.0-138.0
Carbadox	263.2-231.2
Carbamazepine	237.4-194.2
Cefotaxime	456.4-396.1
Ciprofloxacin	332.2-314.2
Clinafloxacin	366.3-348.1
Cloxacillin	469.1-160.1
Codeine	300.0-152.0
Cotinine	177.0-98.0
Dehydronifedipine	345.5-284.1
Digoxigenin	391.2-355.2
Digoxin	781.5-113.1
Diltiazem	415.5-178.0
1,7-Dimethylxanthine	181.2-124.0
Diphenhydramine	256.8-168.1
Enrofloxacin	360.0-316.0
Erythromycin	734.4-158.0
Erythromycin anhydrate	716.4-158.0
Flumequine	262.0-173.7
Fluoxetine	310.3-148.0
Lincomycin	407.5-126.0

# Table 1. Group 1 Compounds from EPA Method 1694 (contd.)

<b>Compound</b>	<b>MRM Transition</b>
Lomefloxacin	352.2-308.1
Miconazole	417.0-161.0
Norfloxacin	320.0-302.0
Norgestimate	370.5-124.0
Ofloxacin	362.2-318.0
Ormetoprim	275.3-259.1
Oxacillin	402.2-160.2
Oxolinic acid	244.1-216.1
Penicillin G	335.1-160.1
Penicillin V	373.2-182.2
Roxithromycin	837.0-158.8
Sarafloxacin	386.0-299.0
Sulfachloropyridazine	285.0-156.0
Sulfadiazine	251.2-156.1
Sulfadimethoxine	311.0-156.0
Sulfamerazine	265.0-156.0
Sulfamethazine	279.0-156.0
Sulfamethizole	271.0-156.0
Sulfamethoxazole	254.0-156.0
Sulfanilamide	173.2-108.2
Sulfathiazole	256.3-156.0
Thiabendazole	202.1-175.1
Trimethoprim	291.0-230.0
Tylosin	916.6-174.2
Virginiamycin	526.3-508.3

# Results

Figure 1 shows the LC-MS-MS chromatogram of a standard mix of the Group 1 compounds at the CS5 level in Method 1694. Good chromatographic resolution was obtained using the Fused-Core LC column.

A few compounds were not detected in the spiked sample. Most of these were –cillin type compounds, and further experiments with the SPE phase are needed to check the retention characteristics on the SPE phase for these compounds.

Finally, a number of differences between the published method MRM transitions and those used here were seen. All transitions used are listed in Table 1.



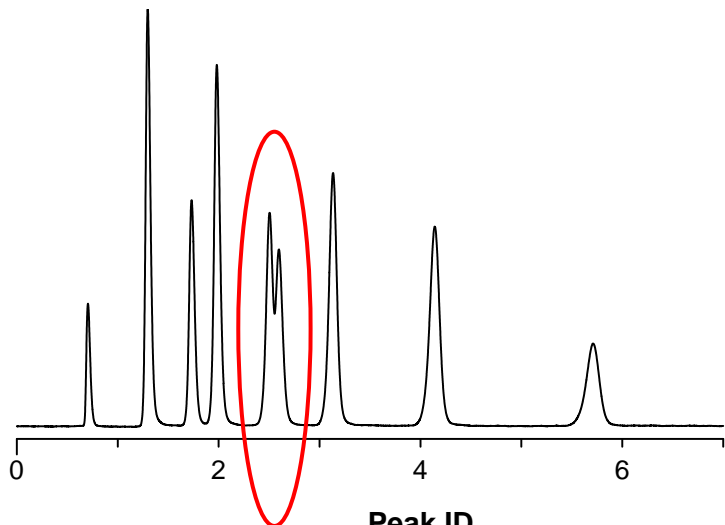
## Example 3. Tricyclic Antidepressants

Initially, drop-in substitution is made with Ascentis Express for 3  $\mu\text{m}$  C18 column, with adjustment in mobile phase composition to yield isoelutropic elution of latest eluting peak.

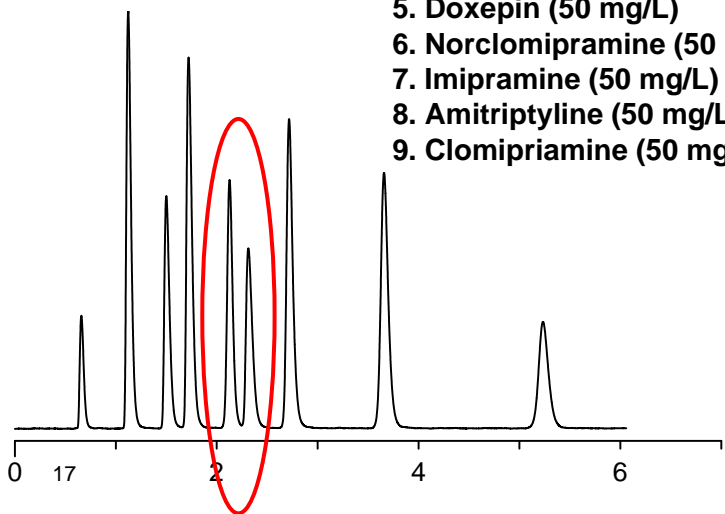
While backpressure is higher, though within operating conditions of a conventional system, resolution of critical pair is achieved.



# Method Transfer with Gains in Resolution



- Peak ID**
1. Uracil
  2. Nordoxepin (50 mg/L)
  3. Desipramine (50 mg/L)
  4. Nortriptyline (50 mg/L)
  5. Doxepin (50 mg/L)
  6. Norclomipramine (50 mg/L)
  7. Imipramine (50 mg/L)
  8. Amitriptyline (50 mg/L)
  9. Clomipramine (50 mg/L)



column: **3  $\mu$ m C18**, 10 cm x 2.1 mm I.D.  
 mobile phase A: 100 mM ammonium acetate, pH 7.0  
 (titrated with ammonium hydroxide)  
 mobile phase B: water  
 mobile phase C: methanol  
 online mixing: A:B:C = 10:25:65  
 flow rate: **0.4 mL/min.**  
 temp.: 55 °C  
 det.: 250 nm  
 injection: 1  $\mu$ L  
 pressure: **1750 psi (121 bar)**  
 $k_{(clomipramine)}$ : **7**  
 $R_{s(norclomipramine)}$ : **0.61**

column: **Ascentis Express C18**, 10 cm x 2.1 mm I.D.  
 mobile phase A: 100 mM ammonium acetate, pH 7.0  
 (titrated with ammonium hydroxide)  
 mobile phase B: water  
 mobile phase C: methanol  
 online mixing: A:B:C = 10:28:62 (**isoelutropic for  
 clomipramine**)  
 flow rate: **0.3 mL/min.**  
 temp.: 55 °C  
 det.: 250 nm  
 injection: 1  $\mu$ L  
 pressure: **2990 psi (206 bar)**  
 $k_{(clomipramine)}$ : **7**  
 $R_{s(norclomipramine)}$ : **1.71**



## Example 4. Comparisons to sub-2 $\mu\text{m}$ columns

At a given column length, Ascentis Express provides similar performance at much lower backpressures. This affords the opportunity for reducing runtime all within the constraints of whatever instrument is available.

At equivalent flow and backpressure, Ascentis Express provides superior performance by virtue of longer column length.

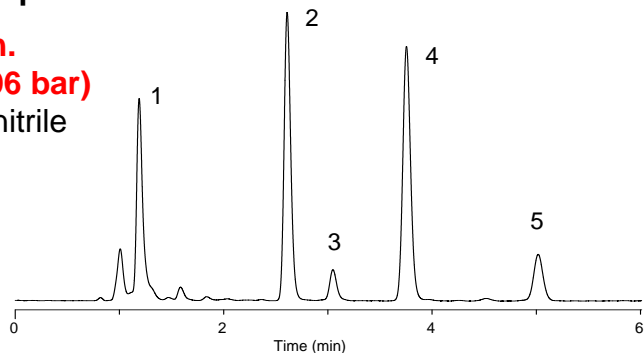
# Fast LC, Moderate Pressure vs sub-2 $\mu\text{m}^1$



innovat

## Ascentis Express C18

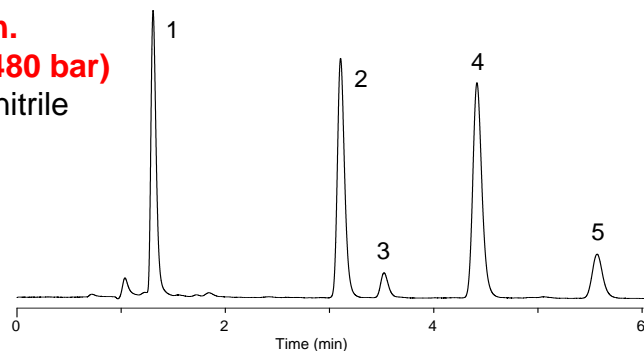
0.2 mL/min.  
1390 psi (96 bar)  
45% acetonitrile



column: 10 cm x 2.1 mm I.D.  
mobile phase: water:acetonitrile; isoelutropic for  $\beta$ -Estradiol  
flow rate: variable  
det.: 200 nm  
injection: 1  $\mu\text{L}$

## Sub-2 $\mu\text{m}$ competitor 2

0.2 mL/min.  
7000 psi (480 bar)  
51% acetonitrile

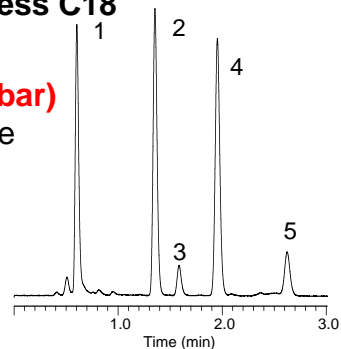


### Peak ID:

1. Estriol
2.  $\beta$ -Estradiol
3. Contaminant
4. Estrone
5. Estrone Degradant

## Ascentis Express C18

0.4 mL/min.  
2880 psi (199 bar)  
45% acetonitrile



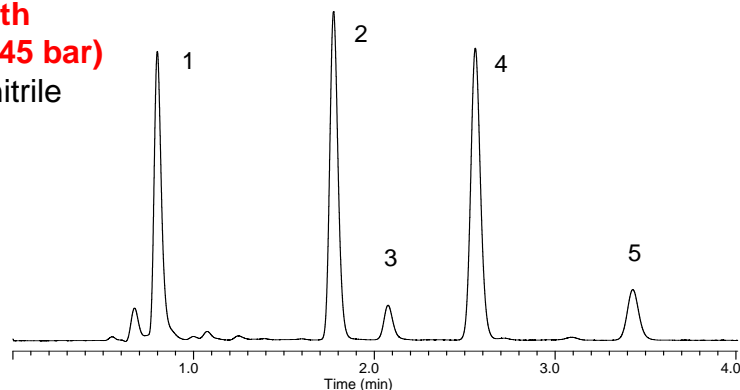
Column	Flow mL/ min.	Peak 1 N	Peak 2 N	Peak 3 N	Peak 3 Rs	Peak 4 N	Peak 5 N
Ascentis Express C18	0.2	2180	7970	9040	3.58	11760	14160
Ascentis Express C18	0.4	1540	5820	6700	3.12	9020	11200
Sub-2 $\mu\text{m}$ competitor	0.2	3150	9590	10430	3.15	12200	14070

1. This estrogen application was performed on a nonoptimized system.

# Fast LC, Moderate Pressure vs sub-2 $\mu\text{m}$

## Ascentis Express C18

10 cm length  
2100 psi (145 bar)  
45% acetonitrile



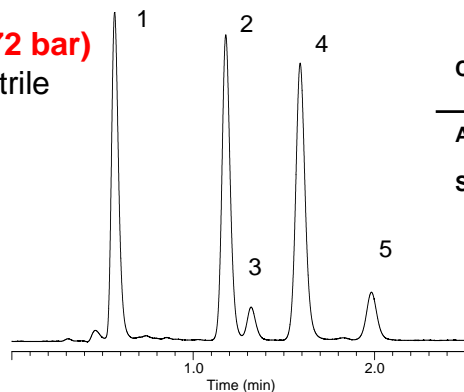
column: 2.1 mm I.D.  
mobile phase: water:acetonitrile; **isoelutropic for  $\beta$ -Estradiol**  
flow rate: **0.3 mL/min**  
det.: 200 nm  
injection: 1  $\mu\text{L}$

### Peak ID:

1. Estriol
2.  $\beta$ -Estradiol
3. Contaminant
4. Estrone
5. Estrone degradant

## Sub-2 $\mu\text{m}$ competitor 1

5 cm length  
2500 psi (172 bar)  
46% acetonitrile



Column	Peak 1 N	Peak 2 N	Peak 3 N	Peak 3 Rs	Peak 4 N	Peak 5 N
Ascentis Express C18	1780	6890	7180	3.28	10360	12500
Sub-2 $\mu\text{m}$ competitor	980	3160	3260	1.58	4330	5210

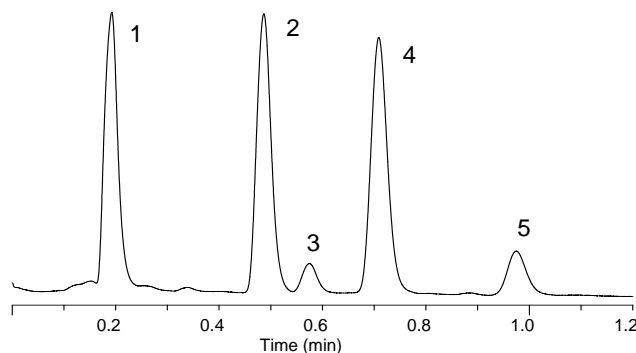
# Fast LC, High Pressure vs sub-2 $\mu\text{m}$

## Ascentis Express C18

**10 cm length**

45% acetonitrile

1.5 mL/min.



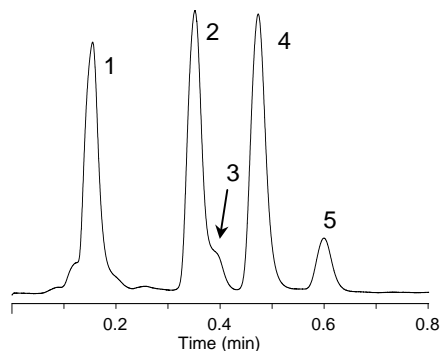
column: 2.1 mm I.D.  
mobile phase: water:acetonitrile; **isoelutropic for  $\beta$ -Estradiol**  
flow rate: **variable**  
det.: 200 nm  
injection: 1  $\mu\text{L}$   
pressure: **11,700 psi (807 bar)**

## Sub-2 $\mu\text{m}$ competitor 1

**5 cm length**

46% acetonitrile

1.38 mL/min.



### Peak ID:

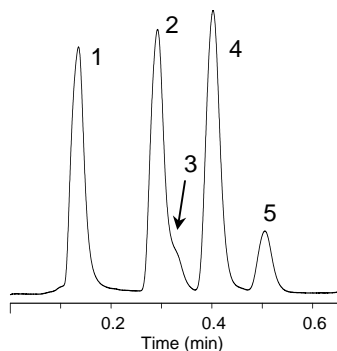
1. Estriol
2.  $\beta$ -Estradiol
3. Contaminant
4. Estrone
5. Estrone degradant

## Sub-2 $\mu\text{m}$ competitor 2

**5 cm length**

51% acetonitrile

1.35 mL/min.



## Conclusions

The relatively low backpressure of Fused-Core columns permits transfer of methods from traditional porous particle columns while realizing shorter run times and/or improved performance, all with conventional HPLC instrumentation.

The extent of improvements in throughput are determined by system limitations and/or objectives of the method.

For a given column length, Fused-Core columns provide similar performance to sub-2  $\mu\text{m}$  columns, but at much lower backpressure.

At constant pressure, improved performance of Fused-Core columns over sub-2  $\mu\text{m}$  columns is realized by use of approximately twice the column length.

# Trademarks

Ascentis, Supel – Sigma-Aldrich Biotechnologies LP  
Fused-Core – Advanced Materials Technology, Inc.