

Experimental Strategies to Improve Drug-target Identification in Mass Spectrometry-based Thermal Stability Assays

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Overview

Purpose: Mass-spectrometry-based thermal stability assays (MS-TSA) enable the on and off-target profiling of drugs^{2,3}, which is a key bottleneck in drug development¹. We investigated a combination of experimental MS-based approaches for the improved qualitative and quantitative performance of MS-TSA.

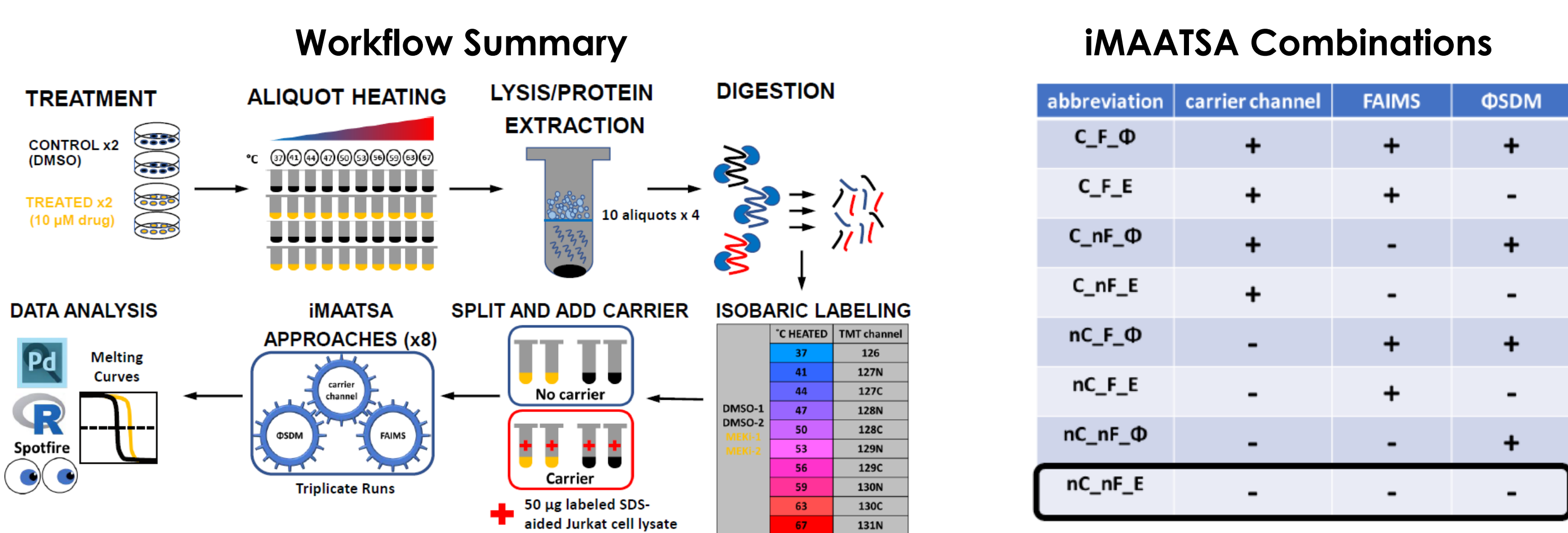
Methods: Jurkat cells were treated with either DMSO (control) or a MEK inhibitor. Melt curves were prepared by heating aliquots to different temperatures, digesting supernatant, and labelling with TMT10 plex. Half of the samples were spiked with an isobarically labelled whole cell digest. Unfractionated samples were evaluated with eight experimental data acquisition approaches.

Preliminary Data: Individually, each of the evaluated experimental approaches demonstrated benefits compared to the control analysis, while the approach implementing all three technologies (Φ SDM, FAIMS, and a carrier channel) produced the most unique high-quality protein melt curve comparisons.

Introduction

Global and targeted mass spectrometry-based thermal stability assays (MS-TSAs) have recently emerged as one of the most promising solutions for the identification of protein-drug interactions^{2,3}. MS-TSAs exploit the phenomenon of ligand-induced thermal stabilization of proteins, whereby modulated protein melting temperatures in drug-treated samples compared to a control set indicate protein-drug binding. We have investigated Phased-constrained Spectral Deconvolution Method^{4,7} (Φ SDM), Field Asymmetric Ion Mobility Spectrometry⁵ (FAIMS), and the implementation of an isobaric carrier channel⁶ individually and in combination as improved MS-based acquisition approaches for thermal stability assays (iMAATSA) for the improved qualitative and quantitative performance of MS-TSA.

Methods

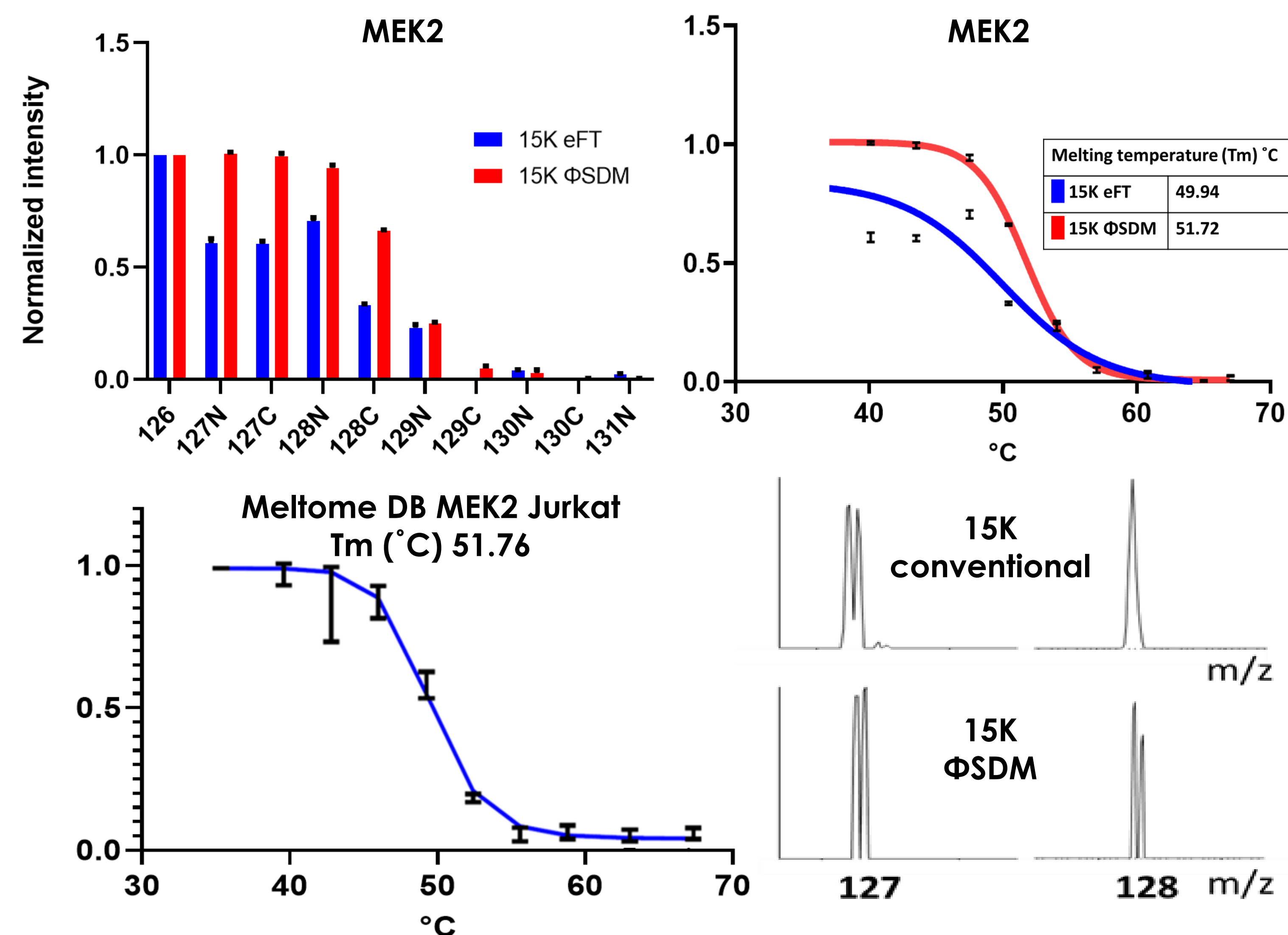


Easy Spray ES803
 90-minute gradient
 FAIMS CV: -35, -50, -65
 eFT: 54 ms max IT
 64 ms transient length
 30K resolution setting
 Φ SDM: 22 ms max IT
 32 ms transient length
 15K resolution setting

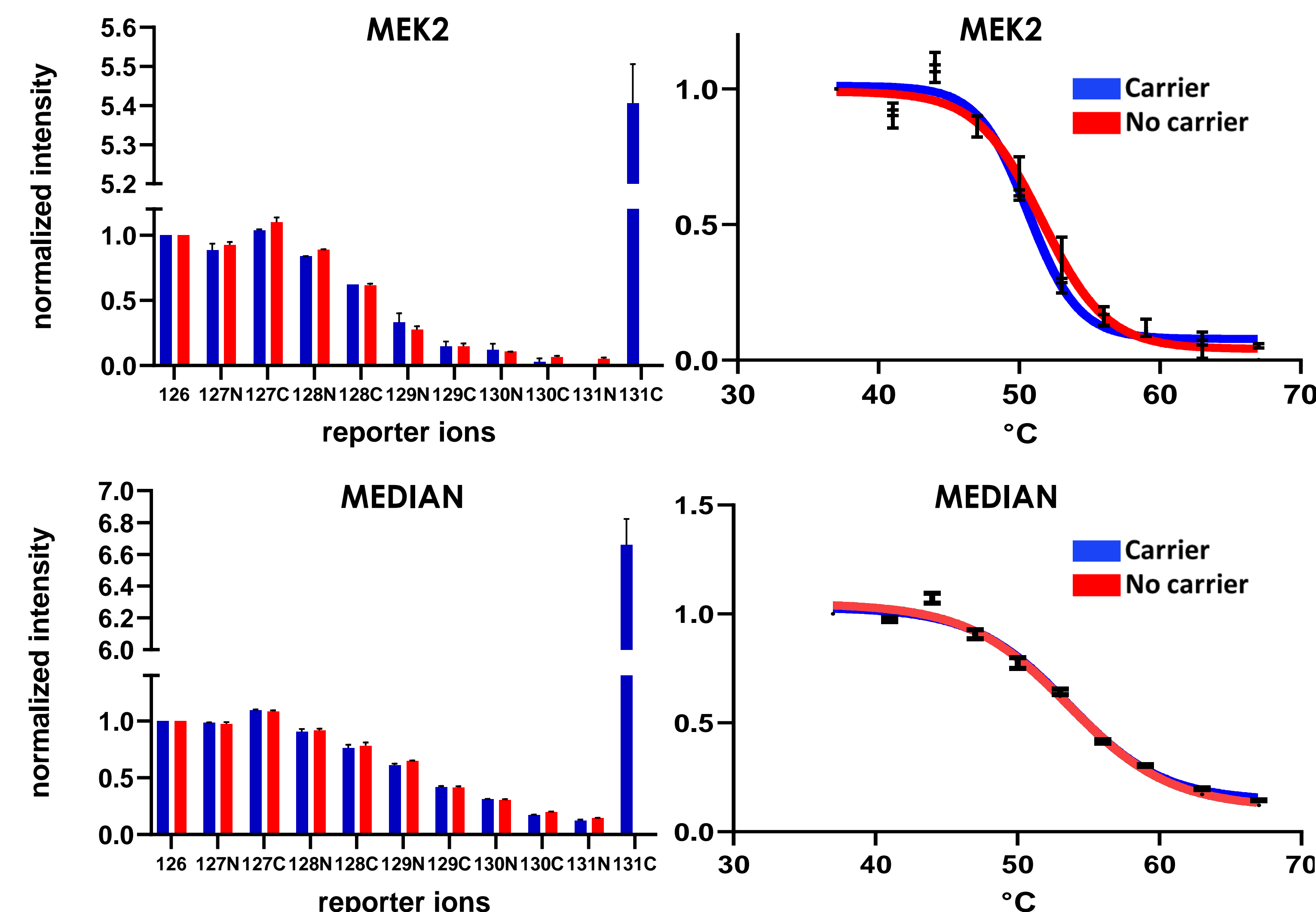
Φ SDM ~ Fourier transform algorithm implemented for increased resolution and speed
 FAIMS ~ Separates ions based on their interaction with a charged field and reduces isolation interference
 isobaric carrier channel ~ Detergent-aided isobarically labeled whole-cell digest used to increase the signal of low intensity peptides

Results

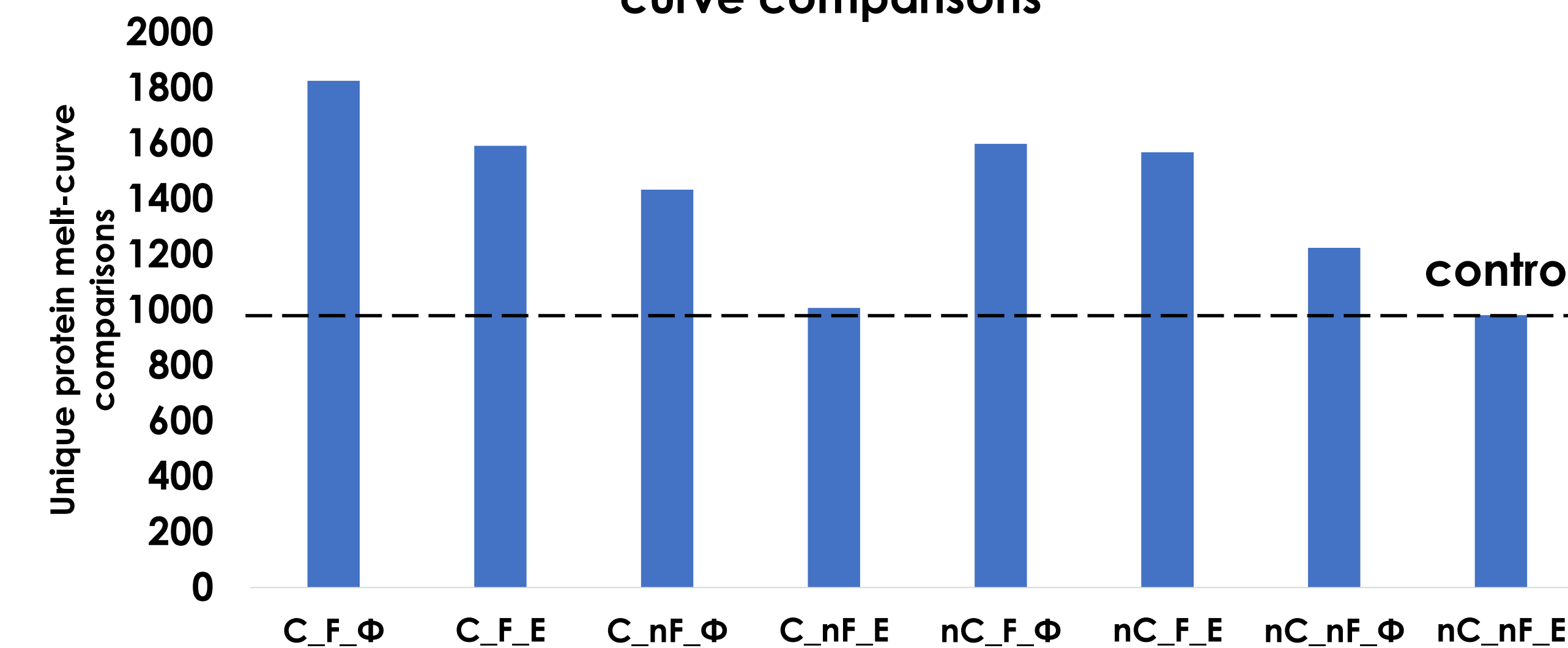
Φ SDM provides sufficient resolving power to separate 15N and 13C TMT reporter ions at a resolution setting of 15K, which resulted in accurate melting curves at high scan speed



The inclusion of a whole-cell labelled digest as a carrier channel (131C) did not significantly alter melt-curve profiles



The addition of each iMAATSA technology appears to have a synergistic effect, with the approach incorporating all three leading to the highest melt-curve comparisons



Conclusions

- Accurate melting curves can be produced using a resolution setting of 15K with Φ SDM
- The use of an isobaric carrier channel does not alter melting-curve profiles or T_m determination
- The use of iMAATSA allowed for significantly more comparisons of high-quality melt-curves
- The use of iMAATSA allowed for significantly more comparisons of high-quality melt-curves

References

1. Seashore-Ludlow, B., Axelsson, H. & Lundböck, T. Perspective on CEISA literature: Toward more quantitative data interpretation. *SLAS DISCOVERY: Advancing the Science of Drug Discovery* 25, 118-126 (2020)
2. Savitski, M. M. et al. Tracking cancer drugs in living cells by thermal profiling of the proteome. *Science (New York, N.Y.)* 346, 1255-784, doi:10.1126/science.1255784 (2014).
3. Molina, D. M. et al. Monitoring Drug Target Engagement in Cells and Tissues Using the Cellular Thermal Shift Assay. *Science* 341, 84, doi:10.1126/science.1233606 (2013)
4. Keltrop, C.; Aizikov, K.; Bath, T.; Kreuzman, A.; Grinfeld, D.; Lange, O.; Mourad, D.; Makarov, A.; Olsen, J. Limits for Resolving Isobaric Tandem Mass Tag Reporter Ions Using Phase-Constrained Spectrum Deconvolution. *Journal of Proteome Research*, 2018, 1535-3893
5. Schweppe, D. K. et al. Characterization and Optimization of Multiplexed Quantitative Analyses Using High-Field Asymmetric-Waveform Ion Mobility Mass Spectrometry. *Analytical Chemistry* 91, 4010-4016, doi:10.1021/acs.analchem.8b05399 (2019)
6. Yi, L. et al. Boosting to Amplify Signal with Isobaric Labeling (BASIL) Strategy for Comprehensive Quantitative Phosphoproteomic Characterization of Small Populations of Cells. *Analytical chemistry* 91, 5794-5801, doi:10.1021/acs.analchem.9b00024 (2019)
7. D. Grinfeld, K.; Aizikov, A.; Kreuzmann, E.; Damoc, A.; Makarov, A. Phase-Constrained Spectrum Deconvolution for Fourier Transform Mass Spectrometry. *Anal. Chem.* 2017 89 (2), 1202-1211
8. Jarzab, A.; Kurzawa, N.; Hopf, T. et al. Meltome atlas—thermal proteome stability across the tree of life. *Nat Methods* 17, 495-503 (2020). <https://doi.org/10.1038/s41592-020-0801-4>

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