

Applying 'MRM Spectrum Mode' and Library Searching for Enhanced Reporting Confidence in Routine Pesticide Residue Analysis

ASMS 2017 TP-194

David Baker¹, Christopher Titman¹, Neil Loftus¹,
Jonathan Horner²

¹Shimadzu, Manchester, UK

²Concept Life Sciences, Cambridge, UK

Applying 'MRM Spectrum Mode' and Library Searching for Enhanced Reporting Confidence in Routine Pesticide Residue Analysis

Introduction

To help reduce the incidence of false positive and false negative reporting in pesticide residue monitoring routine multiple-reaction monitoring (MRM) methods have been enhanced to monitor a higher number of fragment ion transitions to increase specificity and reporting confidence. In this workflow, typically 6-10 fragment ion transitions were monitored for each target pesticide as opposed to a conventional approach using 2-3 fragment ions. By acquiring a high number of fragment ion transitions, each

target pesticide had a corresponding fragmentation spectra which could be used in routine library searching and compound verification using reference library match scores. This 'MRM Spectrum mode' was applied to quantify and identify 193 pesticides using 1,291 MRM transitions without compromising limits of detection, linearity or repeatability using a high speed data acquisition triple quadrupole MS/MS.

Materials and Methods

Pesticide spiked samples, extracted using established QuEChERS based methods, were provided by Concept Life Sciences, UK. Matrices included turmeric, plum, peppermint, parsnip, cherry, lime, pumpkin, tomato and potato. Final extracts were prepared in acetonitrile without

any dilution and directly injected into the LC-MS/MS. A water co-injection method, performed automatically in the auto-sampler, was used to improve early eluting peak shapes in addition to a sub 2 micron particle size column to improve peak capacity (Table 1) .

Applying 'MRM Spectrum Mode' and Library Searching for Enhanced Reporting Confidence in Routine Pesticide Residue Analysis

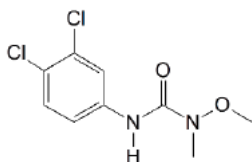
Table 1. LC and MS/MS acquisition parameters used to create the LC-MS/MS method.

Liquid chromatography													
UHPLC	: Nexera LC system												
Analytical column	: HSS T3 (100 x 2.1, 1.7µm)												
Column temperature	: 40°C												
Flow rate	: 0.4mL/minute												
Solvent A	: 5 mmol/L ammonium formate and 0.004% formic acid												
Solvent B	: 5 mmol/L ammonium formate and 0.004% formic acid in methanol												
Binary Gradient	: <table border="1" style="margin-left: 20px;"> <thead> <tr> <th>Time (mins)</th> <th>%B</th> </tr> </thead> <tbody> <tr> <td>1.50</td> <td>35</td> </tr> <tr> <td>11.50</td> <td>100</td> </tr> <tr> <td>13.00</td> <td>100</td> </tr> <tr> <td>13.01</td> <td>3</td> </tr> <tr> <td>15.00</td> <td>Stop</td> </tr> </tbody> </table>	Time (mins)	%B	1.50	35	11.50	100	13.00	100	13.01	3	15.00	Stop
Time (mins)	%B												
1.50	35												
11.50	100												
13.00	100												
13.01	3												
15.00	Stop												
Injection volume	: 0.1 µl (plus 30µl water)												
LC-MS/MS Mass spectrometry	MRM Spectrum Mode : generating library searchable spectra												
Target number of compounds	: 193												
Total number of MRM transitions	: 1,291 transitions (1,229 in ESI+ and 62 in ESI-)												
Pause time/dwell time	: 1 msec/3 msec												
Ionisation mode	: ESI +/-												
Polarity switching time	: 5 msec												
Source temperatures (interface; heat block; DL)	: 350°C; 300°C; 150°C												
Gas flows (nebulising; heating; drying)	: 3L/min; 10 L/min; 10L/min												
LC-MS/MS Mass spectrometry	2 MRM method												
Target number of compounds	: 193												
Total number of MRM transitions	: 386 (374 in ESI+ and 12 in ESI-)												
Pause time/dwell time	: 1 msec/3 msec												
Ionisation mode	: ESI +/-												
Polarity switching time	: 5 msec												
Source temperatures (interface; heat block; DL)	: 350°C; 300°C; 150°C												
Gas flows (nebulising; heating; drying)	: 3L/min; 10 L/min; 10L/min												

Applying 'MRM Spectrum Mode' and Library Searching for Enhanced Reporting Confidence in Routine Pesticide Residue Analysis

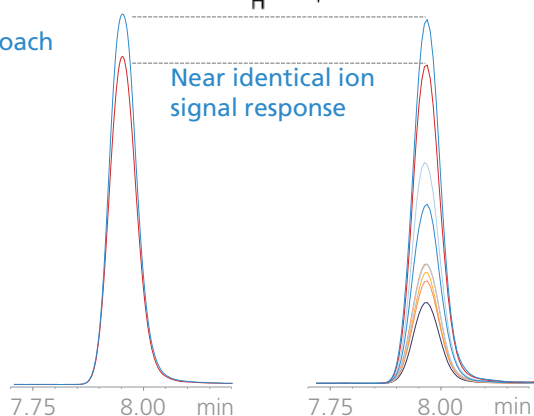
Results

Compound Name Linuron
Formula C₉H₁₀Cl₂N₂O₂
CAS 330-55-2



Higher specificity
Higher reporting confidence
Library searchable fragment data

Conventional approach
2 MRM's
1:248.80>160.00
2:248.80>182.10



MRM Spectrum Mode
9 MRM's
1:248.80>160.00
2:248.80>182.10
3:250.80>162.00
4:248.80>133.10
5:250.80>135.00
6:248.80>161.00
7:250.80>184.10
8:248.80>125.00
9:248.80>153.00

Figure 1. Using a high speed triple quadrupole mass analyser a higher number of fragment ions were acquired in MRM increasing the specificity of detection and reducing false negative and false positive reporting. In the case of linuron, 9 precursor-fragment ion transitions were used to increase confidence in assay specificity. There is no compromise in data quality between methods despite a higher number of fragment ions monitored. Signal intensity, linearity, reproducibility are in good agreement between both methods.

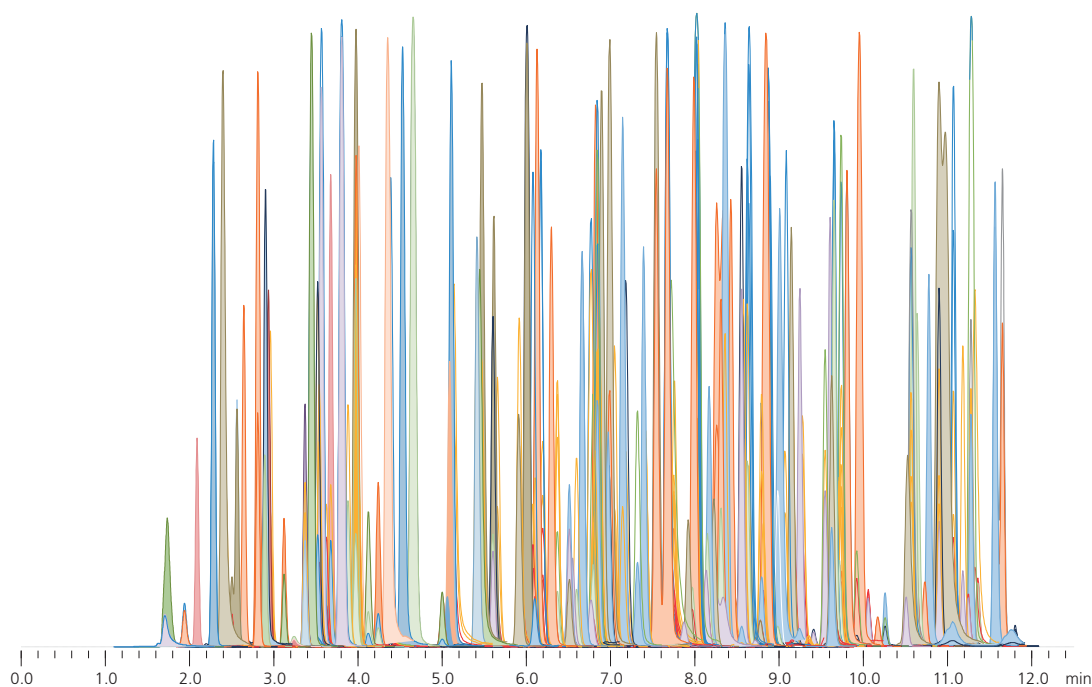


Figure 2. MRM chromatogram for all 193 pesticides spiked at 0.010 mg/kg measured with MRM Spectrum mode. Using this mode 1,291 MRM transitions were measured for 193 pesticides. Despite the high data density acquired with MRM Spectrum mode (for example, 151 MRM transitions were registered in the same time window during the analysis, see Figure 2) sensitivity was not affected by the high data acquisition rate.

Applying 'MRM Spectrum Mode' and Library Searching for Enhanced Reporting Confidence in Routine Pesticide Residue Analysis

On average 7 MRM transitions were applied to each compound, with more than 10 MRM transitions applied to 34 compounds. All MRM transitions were acquired throughout the MRM window without the need for triggering thresholds. The method includes a total of 1,291 MRM transitions for 193 pesticides in a run time

of only 15 minutes. A dwell time of 3 msec was applied to every MRM transition. In order to evaluate the data quality from the MRM Spectrum Mode method, the same method was set up with 2 MRMs applied to each compound (386 MRMs in total) using the same acquisition method (Table 1).

MRM Spectrum based identification

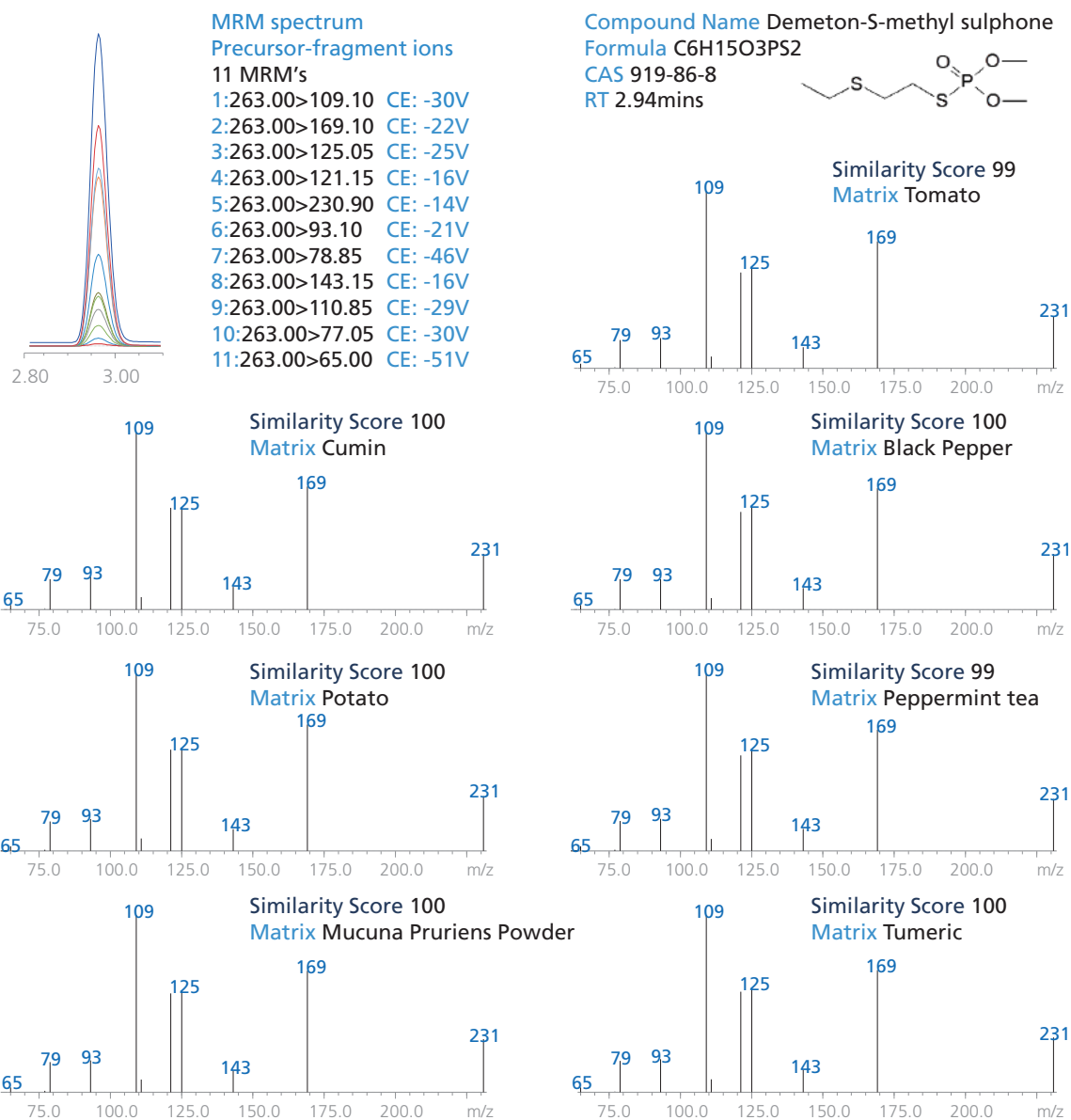


Figure 3. MRM spectrum identification in different matrices for demeton-S-methyl sulphone. In this study, the number of qualifier fragment ion transitions was increased for each pesticide and the combined transitions were used to create a MRM product ion spectrum. The product ion spectrum was used in conventional library matching routines comparing against a reference spectrum to generate a similarity score.

Applying 'MRM Spectrum Mode' and Library Searching for Enhanced Reporting Confidence in Routine Pesticide Residue Analysis

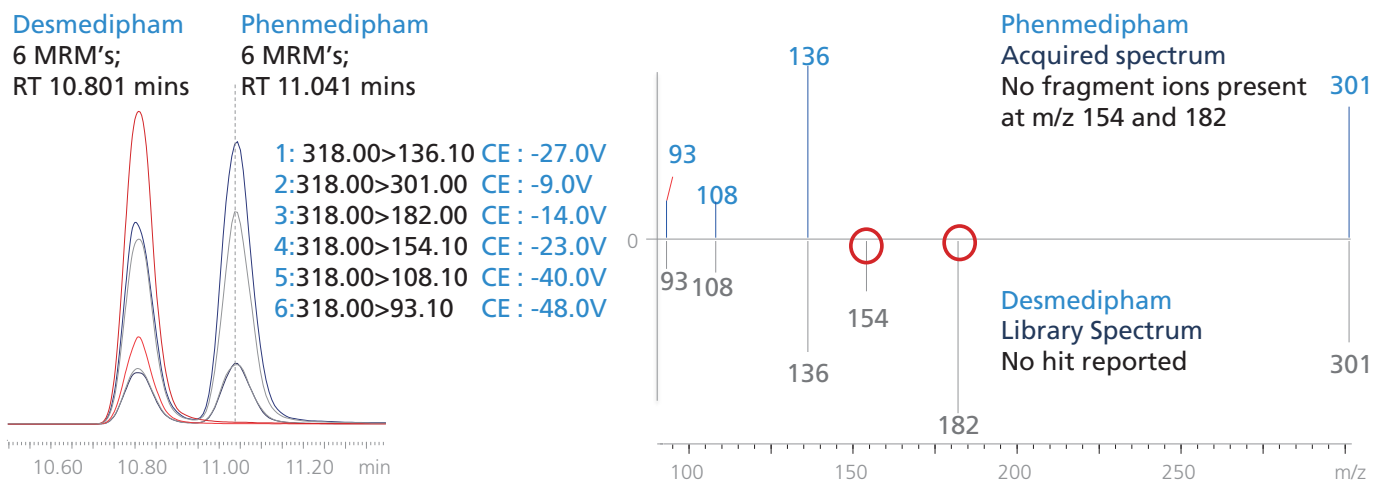


Figure 4. MRM chromatogram for desmedipham and phenmedipham spiked into a cumin extract at 0.1 mg/kg. As phenmedipham shares common transitions and elutes at a similar retention time as desmedipham the MRM spectrum can be used to distinguish between both pesticides to avoid false positive reporting.

Applying 'MRM Spectrum Mode' and Library Searching for Enhanced Reporting Confidence in Routine Pesticide Residue Analysis

MRM Spectrum Quantitation and Library Searching

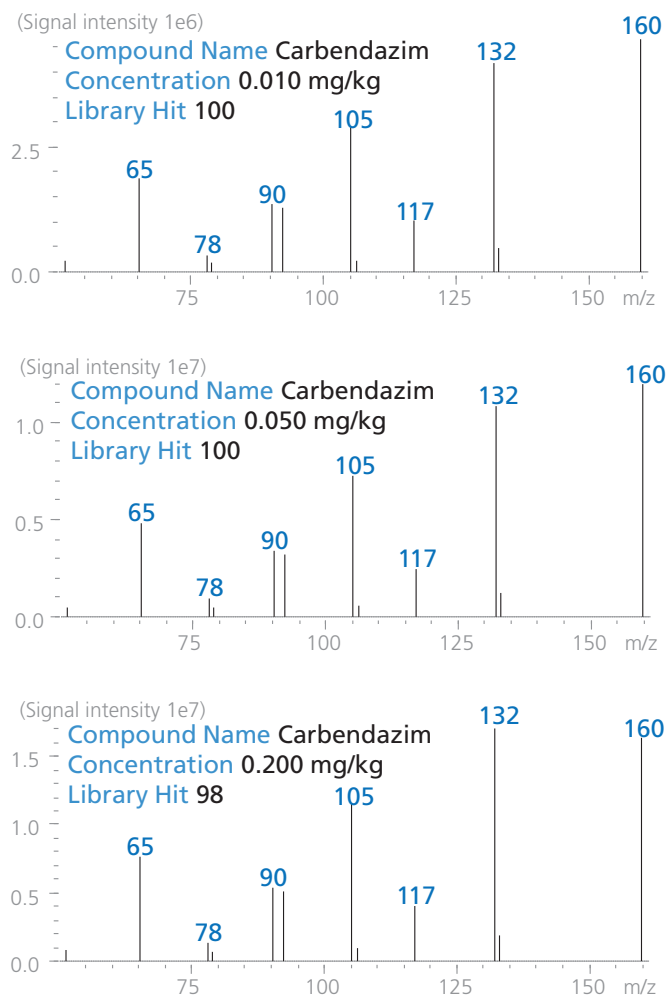
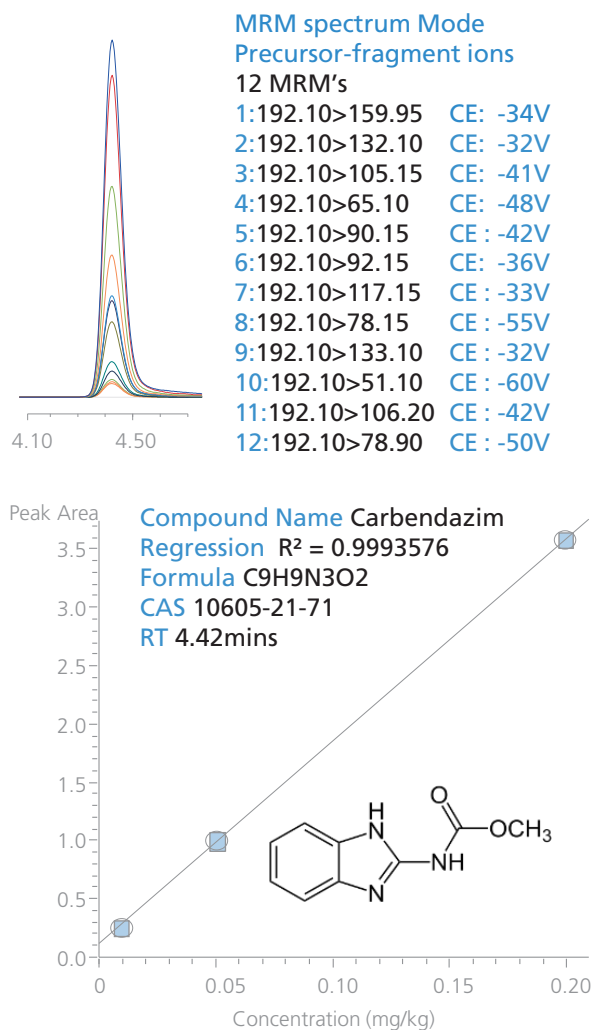


Figure 5. The limit on the number of MRM transitions used to generate a product ion spectrum is dependent on the chemical structure of the pesticide molecule. In the case of carbendazim, several bonds could be broken using collision energies between 10-60V resulting in a product ion spectrum of 12 fragment ions. The product ion spectrum can then be used for library search and analyte confirmation as shown above. For each calibration level ranging from 0.010-0.200mg/kg the library similarity score was greater than 99 confidently confirming the target analyte. The advantage of this technique is that library searchable product ion spectrum data is used in target compound identification without compromising sensitivity, accuracy and robustness in quantitative data reporting.

Applying 'MRM Spectrum Mode' and Library Searching for Enhanced Reporting Confidence in Routine Pesticide Residue Analysis

Conclusions

- False positive results are a major issue for all pesticide residue monitoring laboratories. EU regulations require that retention time and the ion ratio between 2 MRM transitions are within a set threshold. However, even applying this criteria false positives may occur for certain pesticide/commodity combinations.
- We have applied MRM Spectrum mode to identify and quantify 193 target pesticides in a number of different sample matrices. In this workflow the library score is used as an additional identification criterion in order to improve confidence when reporting results.
- Acquisition of the MRM Spectrum method (1,291 MRM transitions) did not compromise data quality when compared to a conventional 2 MRM per compound method (386 MRM transitions) with consistent signal response and repeatability in both methods. The MRM product ion spectrums were consistent across the linear range and between different matrices. The method acquired data in both positive and negative ion modes with a polarity switching time of 5 msec enabling fast cycle times and a high data collection rate.

First Edition: June, 2017



Shimadzu Corporation
www.shimadzu.com/an/

For Research Use Only. Not for use in diagnostic procedures.

This publication may contain references to products that are not available in your country. Please contact us to check the availability of these products in your country.

The content of this publication shall not be reproduced, altered or sold for any commercial purpose without the written approval of Shimadzu. Company names, products/service names and logos used in this publication are trademarks and trade names of Shimadzu Corporation, its subsidiaries or its affiliates, whether or not they are used with trademark symbol "TM" or "®".

Third-party trademarks and trade names may be used in this publication to refer to either the entities or their products/services, whether or not they are used with trademark symbol "TM" or "®".

Shimadzu disclaims any proprietary interest in trademarks and trade names other than its own.

The information contained herein is provided to you "as is" without warranty of any kind including without limitation warranties as to its accuracy or completeness. Shimadzu does not assume any responsibility or liability for any damage, whether direct or indirect, relating to the use of this publication. This publication is based upon the information available to Shimadzu on or before the date of publication, and subject to change without notice.