



CLAM[™]-2030 Fully Automated Sample Preparation Module for LCMS LCMS-8060 Liquid Chromatograph Mass Spectrometer

Measuring Choline and its Metabolites in Human Plasma Using an LC/MS/MS System with Fully Automated Sample Preparation Module

E. Imoto and D. Kawakami

User Benefits

- Capable of quantifying choline, TMA, and TMAO levels in biological samples
- Performs time-consuming sample preparation automatically.
- Eliminates the need for manual sample preparation and reduces variability in quantitative results due to manual operations.

Introduction

It is known that some of the choline ingested in food is partly metabolized to trimethylamine (TMA) by intestinal bacteria and subsequently metabolized to trimethylamine N-oxide (TMAO) by enzymes in the liver. Reports have noted that TMAO is associated with cardiovascular diseases such as arteriosclerosis¹⁾ and that it is useful in predicting the prognosis of heart failure²⁾, As a result, TMAO is now attracting interest as a new biomarker. This article describes a system for simultaneous analysis of choline and choline metabolites in human plasma using an LC/MS/MS system with fully automated sample preparation module (Fig. 1) that automatically performs deproteination of biological samples and adds internal standards.



Fig. 1 LC/MS/MS System with Fully Automated Sample Preparation Module (CLAM[™]-2030 + LCMS-8060)

■ Fully Automated Sample Preparation of Choline and Choline Metabolites in Plasma

The CLAM-2030 performs deproteination and other sample preparation steps automatically by simply setting the blood collection tubes directly into the system. Fig. 2 shows the sample preparation process using the CLAM-2030. Sample analysis by the LC/MS/MS system and preparation of the next sample are performed in parallel. Hence the time required per sample can be reduced substantially. The analysis cycle time from plasma sample preparation to simultaneous analysis of choline and choline metabolites by LC/MS/MS was about 5 min per sample.

■ Sample Preparation

Calibration curves were created from standard samples prepared by serial dilution of choline (1 to 1000 nmol/L), TMA (10 to 1000 nmol/L), and TMAO (10 to 10000 nmol/L) with water. Water was also used to prepare 2 μ mol/L samples of choline-d13, TMA-d9, and TMAO-d9 to use as ISTDs. Choline, TMA, and TMAO levels in plasma were quantified using commercially available human plasma.

The samples were pretreated with the CLAM-2030, analyzed automatically by the LCMS-8060, then calibration curves were created, and analyte levels in the human plasma were determined.

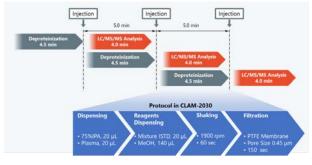


Fig. 2 Preparation of Plasma Samples Using CLAM-2030

Analytical Conditions

HPLC and MS analytical conditions are shown in Table 1. The MRM transitions for each compound are shown in Table 2.

| Table 1 HPLC and MS Analytical Conditions |
|---|
|---|

| Liquid Chromatograph | |
|----------------------|--|
| System: | Nexera [™] X2 |
| Column: | Shim-pack Velox HILIC (50 mm x 2.1 mm I.D., 2.7 μm) |
| Temperature | 40 °C |
| Injection Volume: | 3 µL |
| Mobile Phase A: | 50 mM ammonium formate +0.1 % formic acid—Water |
| Mobile Phase B: | Acetonitrile |
| Flowrate: | 0.4 mL/min |
| Gradient Program | 70 % (0.0 min) ➡ 30 % (2.40 min) ➡ |
| (B conc%): | 5 % (2.41 – 3.00 min) ➡ 70 % (3.41 – 4.00 min) |
| | |

Mass Spectrometer

LCMS-8060 System: ESI (Positive) Ionization: Nebulizing Gas: 31/min Drying Gas: 10 L/min Heating Gas: 10 L/min DL Temp.: 250 °C Heat Block Temp.: 400 °C Interface Temp.: OFF

| Table 2 MRM Transitions for Choline and Choline Metabolite |
|--|
|--|

| Compound Name | lon | Precursor ion (<i>m/z</i>) | Product ion (<i>m/z</i>) | |
|------------------|------------------|---------------------------------|----------------------------|--|
| Chalina | Quantitative lon | 104.10 | 60.10 | |
| Choline | Reference ion | 104.10 | 45.10 | |
| TAAA | Quantitative lon | 60.20 | 44.15 | |
| TMA | Reference ion | 60.20 | 45.05 | |
| ТМАО | Quantitative lon | 76.00 | 58.20 | |
| | Reference ion | 76.00 | 59.10 | |
| Choline-d13 | Quantitative lon | 117.25 | 66.05 | |
| TMA-d9 | Quantitative lon | 68.90 | 49.15 | |
| | Reference ion | 68.90 | 51.15 | |
| TMAO-d9 | Quantitative lon | 85.20 | 66.00 | |
| | Reference ion | 85.20 | 68.25 | |

Checking Calibration Curve Linearity and Reproducibility

Linearity was confirmed after calibration curve standard samples were analyzed three times. The calibration curve range, coefficient of determination (R²), and reproducibility (concentration %RSD) and accuracy of the lowest calibration curve sample concentration are shown for each compound in Table 3. Fig. 3 also shows the calibration curves for each compound and Fig. 4 shows mass chromatograms for standard samples of each compound.

| 14 | Die 5 Analysis Nesul | is nom Standar | u Samples (II – |) | | | |
|------------------|--|---|-----------------------|--------------|--|--|--|
| Compound Name | Calibration Curve Range (nmol/L) | Coefficient of Determination R ² | Concentration %RSD | Accuracy (%) | | | |
| Choline | 1-1000 0.999 14 % 86 % | | | | | | |
| TMA | 10-1000 0.997 12 % 95 % | | | | | | |
| TMAO | 10-10000 | 0.999 | 2 % | 96 % | | | |
| Choline | Avis Ratio 0.7 - 0.0001764146 + 0.000227900 0.4 - 0.99533 & 0.099516 0.5 - 0.001764146 + 0.000227900 0.5 - 0.001764446 + 0.000227900 0.5 - 0.00176446 + 0.00022790 0.5 - 0.00176446 + 0.00022790 0.5 - 0.0017647 + 0.0017646 0.5 - 0.0017647 + 0.0017647 0.5 - 0.0017647 + | | | | | | |
| тма | 0 0 0 10 10 200 10 10 200 10 10 10 10 10 10 10 10 10 10 10 10 10 1 | 0 100 200 300 400 500 600 700 800 900 1000 000 100 100 100 100 100 100 100 100 100 | | | | | |
| | Fig. 3 Calibration | | Conc. | Ratio (nM) | | | |

CLAM and Nexera are trademarks of Shimadzu Corporation in Japan and other countries.

Top: Choline 1 to 1000 nmol/L, Middle: TMA 10 to 1000 nmol/L,

Bottom: TMAO 10 to 10000 nmol/L

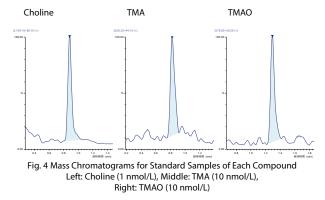
01-00238-EN First Edition: Mar. 2022



. without notice.

Shimadzu Corporation

Analytical & Measuring Instruments Division **Global Application Development Center**



Measuring Choline, TMA, and TMAO Levels in Human Plasma

Choline, TMA, and TMAO concentrations measured in human plasma samples (n = 3) prepared by CLAM-2030 are shown in Table 4, and the mass chromatograms of each component are shown in Fig. 5. Good reproducibility was obtained with human plasma.

Table 4 Quantitative Results from Human Plasma (n = 3)

| 14510 | Quantita | active nes | | lamanı | lasina (n – 5 | / |
|------------------------------------|----------------|------------------------------------|-------------|---------------|-----------------------|-------------------------|
| Compound Name | | Measured Concentration (nmol/L) | | | Concentration %RSD | |
| Choline | | 938 2.8 % | | | % | |
| TMA | TMA 685 1.1 % | | % | | | |
| TMAO | TMAO 655 4.4 % | | % | | | |
| Choline | | ТМА | | | ТМАО | |
| 0 (164.16-40.10 (+) 100.00 s | 100 | \$220-44.15(+) | | 2 Q 76.00 <51 | 8.22 (-) | 17865 |
| 0.00 0.4 0.6 0.8 1.0 | 1.2 RT | 0.2 0.4 | 0.6 0.8 1.0 | 1.2 RT | 0.8 1.0 1.2 | 1.4 1.6 1.8 RT (min) |

Table 3 Analysis Results from Standard Samples (n = 3)

Fig. 5 Mass Chromatograms of Choline, TMA, and TMAO in Human Plasma

Conclusion

Sample preparation was performed using the CLAM-2030, which automatically performs deproteination of biological samples and adds internal standards. Choline, TMA, and TMAO levels in human plasma were quantified using LCMS-8060. The calibration curves for each compound showed good linearity and no reproducibility problems were encountered with human plasma samples, indicating how highly useful this analysis system is.

<References>

1) Wang Z, Stanley L Hazen et al., "Gut flora metabolism of

phosphatidylcholine promotes cardiovascular disease", Nature 472 (2011) 57-63

2) Yazaki Y, Aizawa K et al., "Ethnic difference in association of outcomes with trimethylamine N-oxide in acute heart failure patients", ESC Heart Fail 7(5) (2020) 2373-2378.

<Acknowledaments>

are used with trademark symbol "TM" or "@". Shimadzu disclaims any proprietary interest in trademarks and trade names other than its own

The research described in this document was undertaken with considerable help from Kenichi Aizawa, Clinical Pharmacology Center, Jichi Medical University Hospital.

The information contained herein is provided to you "as is" without warranty of any kind including without limitation warranties as to its accuracy or completeness. Shimadzu does not assume any responsibility or liability for any damage, whether direct or indirect, relating to the use of this publication. This publication is based upon the information available to Shimadzu on or before the date of publication, and subject to change

For Research Use Only. Not for use in diagnostic procedure. This publication may contain references to products that are not available in your country. Please contact us to check the availability of these products in your country.