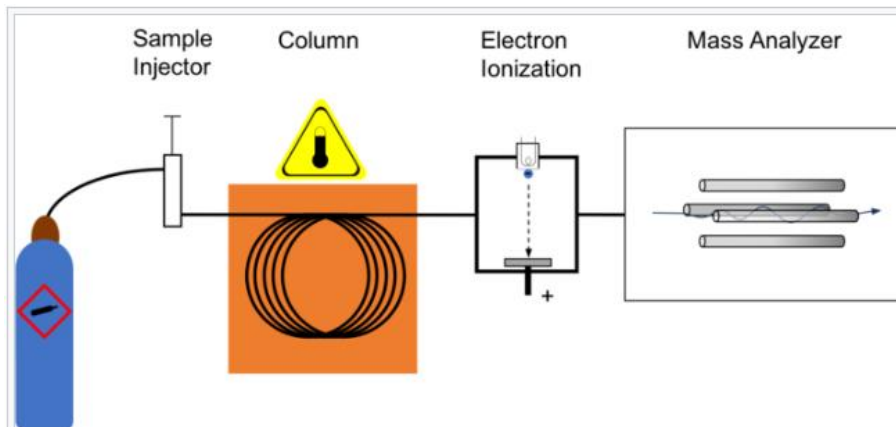


**Free NIST GC-MS Software Lab  
for Universities:  
Part 4: Processing GCMS Data  
with AMDIS**



**James Little**

**August 29, 2023**

**38 years Eastman Chemical Company**

**7 years Mass Spec Interpretation Services**

<https://littlemsandsailing.wpcomstaging.com/>

[Link to GCMS Schematic Above](#)

[Link to University Logos](#)

# Free NIST GC-MS Software Lab for Universities

Part 1: Very Basic Theory of GCMS Analyses

Part 2: Installation of Software

Part 3: Library Searches

Part 4: Processing GCMS Data with AMDIS

Part 5: Understanding EI Fragmentation with MS Interpreter

Part 6: Structure Searches with Input from ChemSketch

Part 7: Creating a User Library

Part 8: Advanced Processing with NIST Software

A "Little" Mass Spec and Sailing  
Organic Mass Spectrometry, NMR, Sailing, Tesla, Duplicate Bridge

Mass Spec Interpretation Services

100  
50  
0

15 16 27 31 39 40 44 43 58

10 20 30 40 50 60

(m/z) Acetone

MASSMAN

569.2876

C<sub>12</sub>H<sub>10</sub>O  
CAN COATING

CAS NO.  
20583873

About Me My Topics Others Links

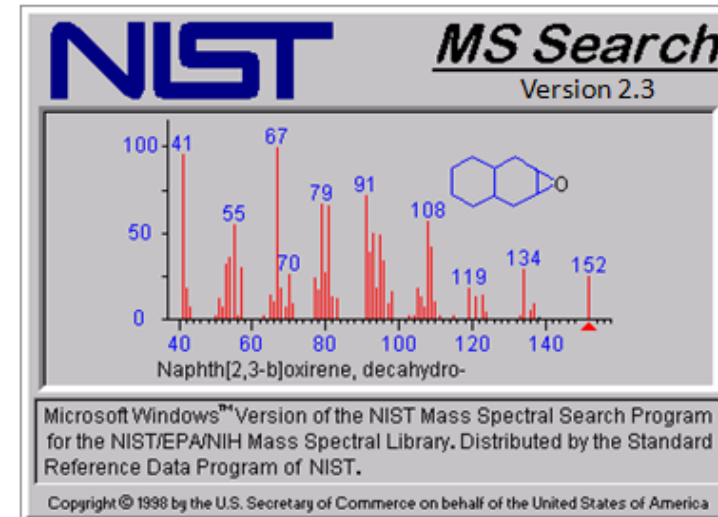
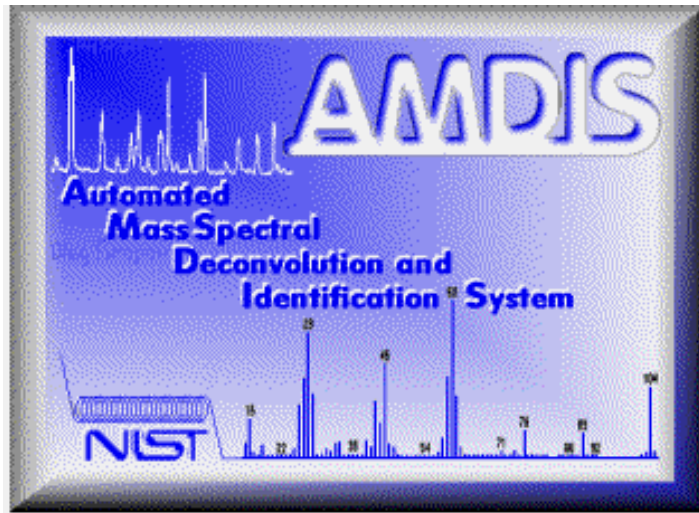
[Link to Training Website](#)

# Modified\* Basic Instructions for Using AMDIS with MS Search

By James L. Little, O. David Sparkman  
Input from Gary Mallard

\*9/6/2020 (Many additional slides on many topics added by JL)

*See AMDIS Manual for Detailed Instructions*



# What is AMDIS?

## Automated Mass spectral Deconvolution and Identification System

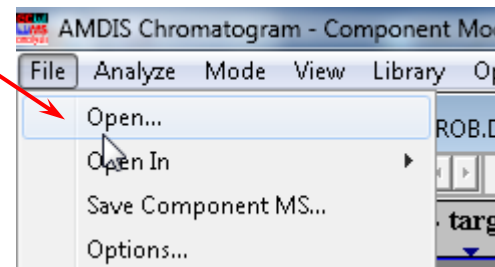
Developed to automatically detect chemicals in violation of Chemical Weapons Convention

- Software to *automatically* separate (deconvolute) chemical background in GC/MS data from signal for sample components
- Deconvoluted spectra can be *sent* to the NIST Mass Spectral Search Program for identification
- If necessary, manual background subtraction performed

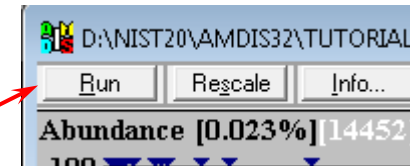
# AMDIS Essentials

- **Must always** open a data file and run deconvolution **before** sending a spectrum to **external** NIST 2.4 Search Program
- **Three ways** to **obtain a spectrum** for searching: automatically, manually by **LMB** on spectrum, or manually with background subtraction
- AMDIS extracts the mass spectra of individual **Components** from chromatograms, these are symbolize with a ▼ on top of the chromatogram at the point of elution

**Must  
always  
first**

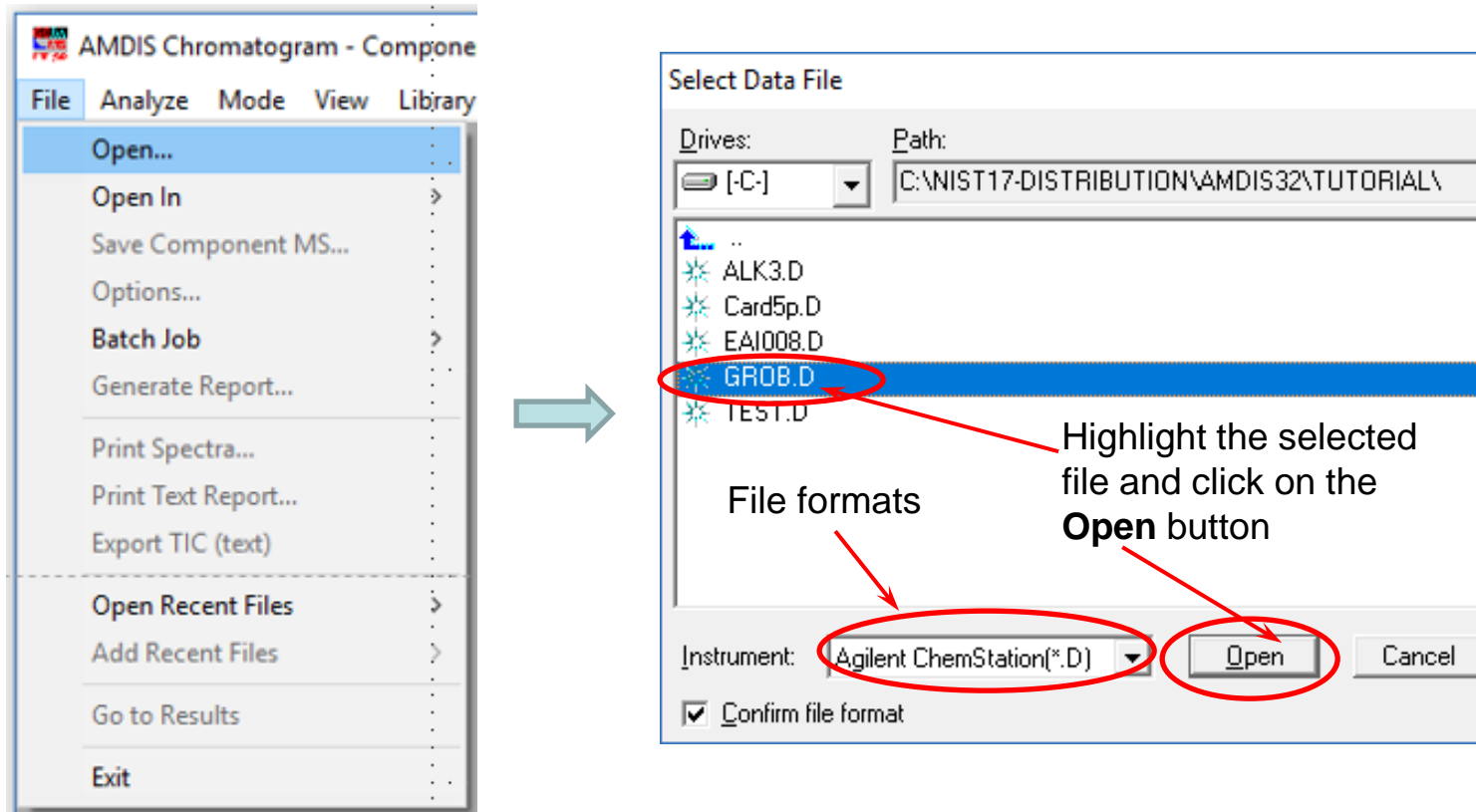


**then**



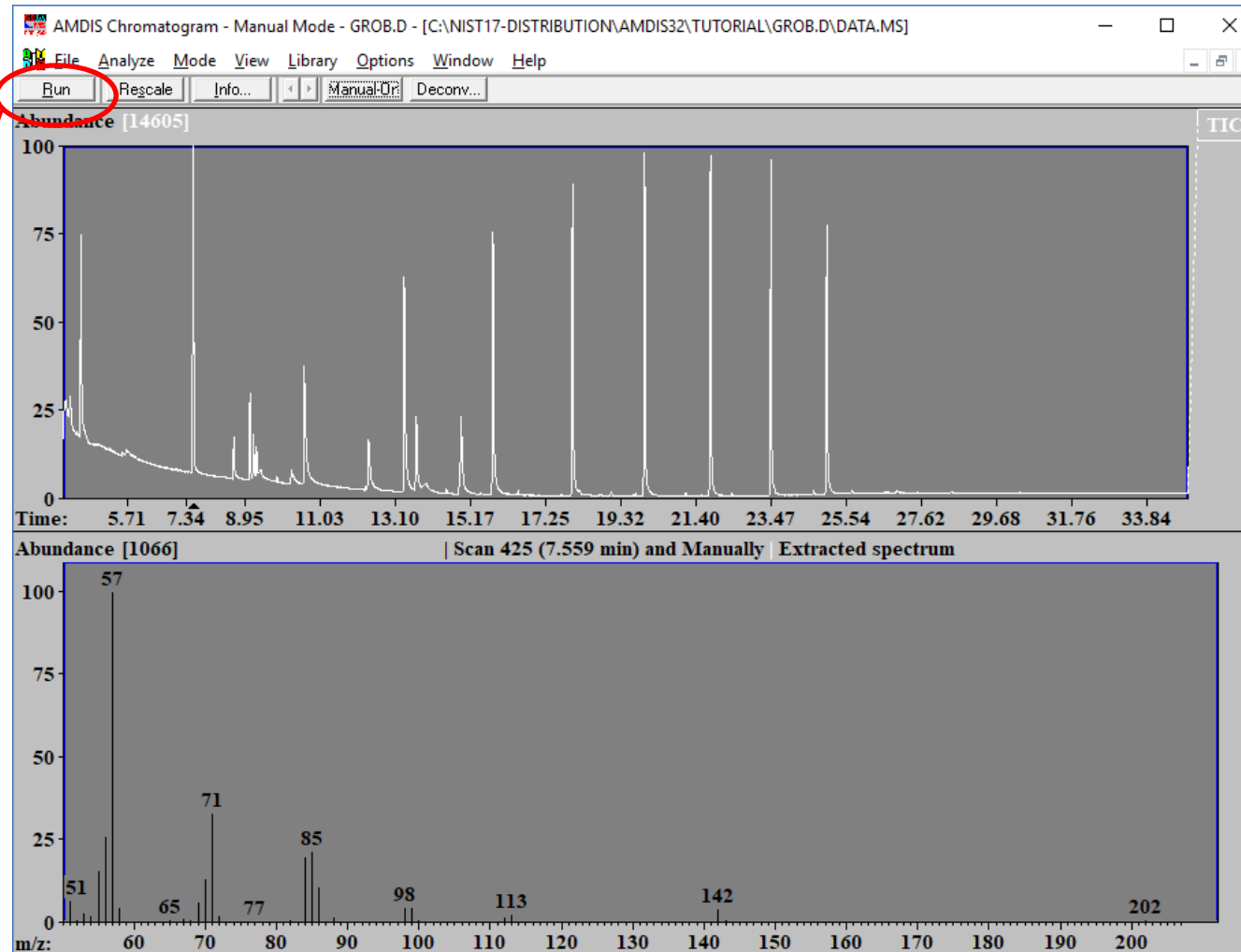
# Opening File with AMDIS

- Can process many different file types with AMDIS including Agilent, netCDF, etc.
- Many manufacturers supply utility programs for conversion of files in their native format to the “standard” netCDF format
- File formats accessed by “pull down” menu
- **Before** sending components to library search, **must** open and run the file to get background corrected spectra



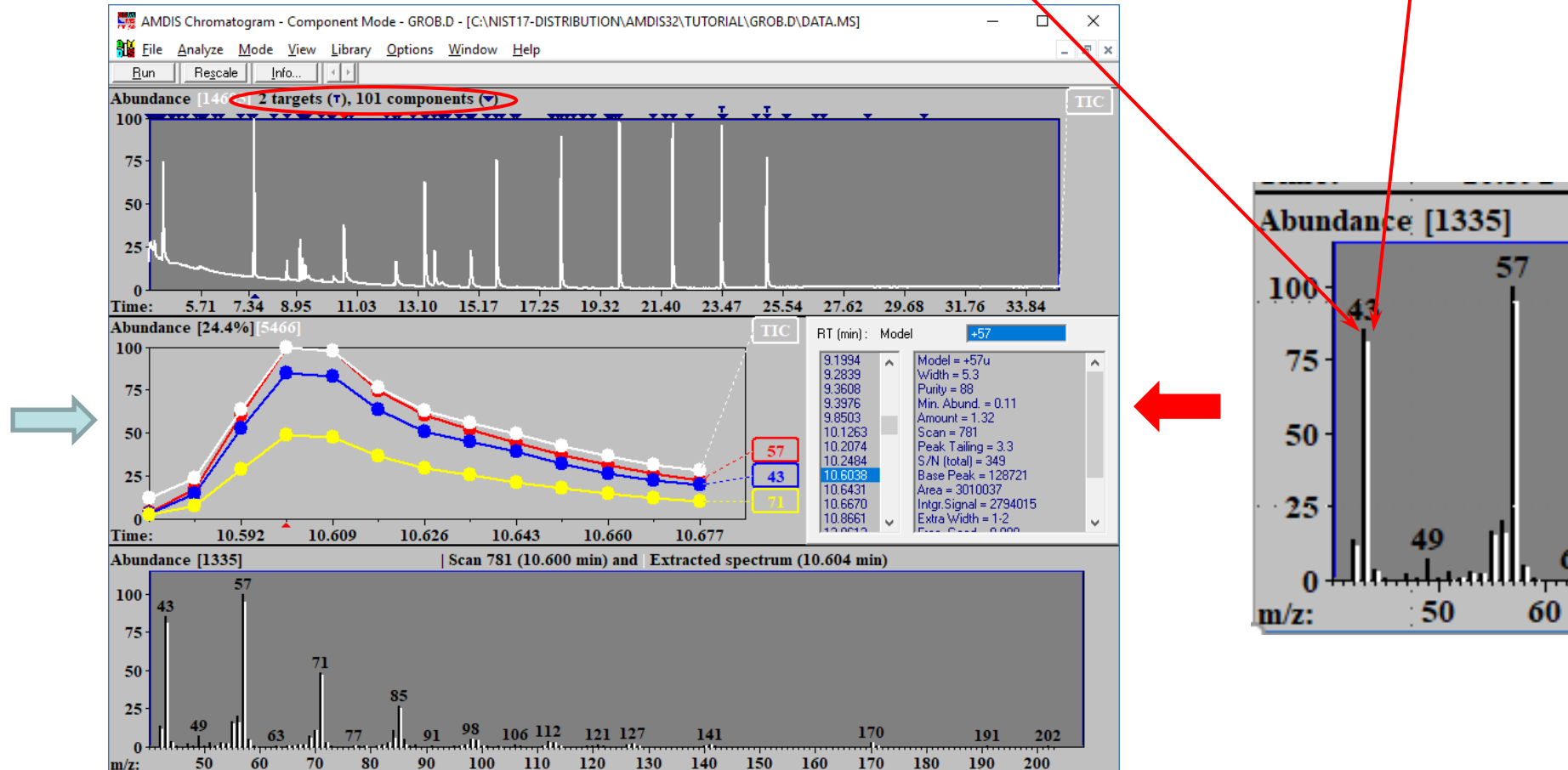
# Deconvoluting Spectra

- First click the **LMB** with the **Pointer** on the **Run** button to deconvolute the file
- The computer plots a chromatogram from every  $m/z$  value in the data file
- Then “looks” at the stacked plots to determine which ions “belong” with each other and subtracts out ions from air, column bleed, other nearby components, etc.



# Evaluating Deconvoluted Results

1. Note the number of **Components** found (101)
2. Note the little blue upside-down triangles (▼), left click on any one to see deconvoluted spectrum
3. After selecting one blue triangle, can step through by using up or down arrows **on your keyboard**
4. The left middle window shows what ions were “modeled” to define your spectrum →
5. The right middle window show you the associated parameters for each peak ←
6. The bottom window shows the unsubtracted spectrum in black and the deconvoluted in white



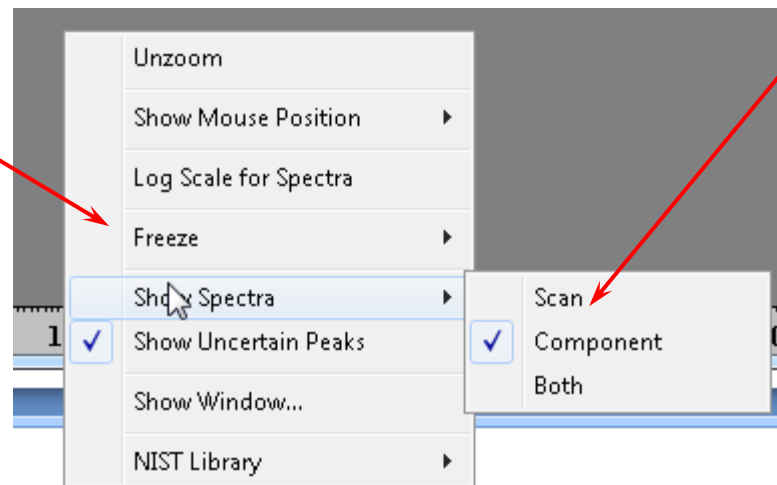


# Evaluating Deconvoluted Results (continued)

- Can just show the **Component** (white peaks), the Scan (black peaks), or Both, but best to get accustomed to looking at both
- When the black matches the white, you probably have a good spectrum of a major **Component**
- For minor **Components**, possibly coeluting with a major **Component**, the white will be different than black and in many cases smaller
- With default “deconvolution parameters”, AMDIS will sometimes ID too many components
- The “deconvolution parameters” need to be adjusted to minimize this
- Very dependent on having a good stable signal from the instrument, but in my experience, just tends to do that without using the appropriate filters for processing (*more on that later*).

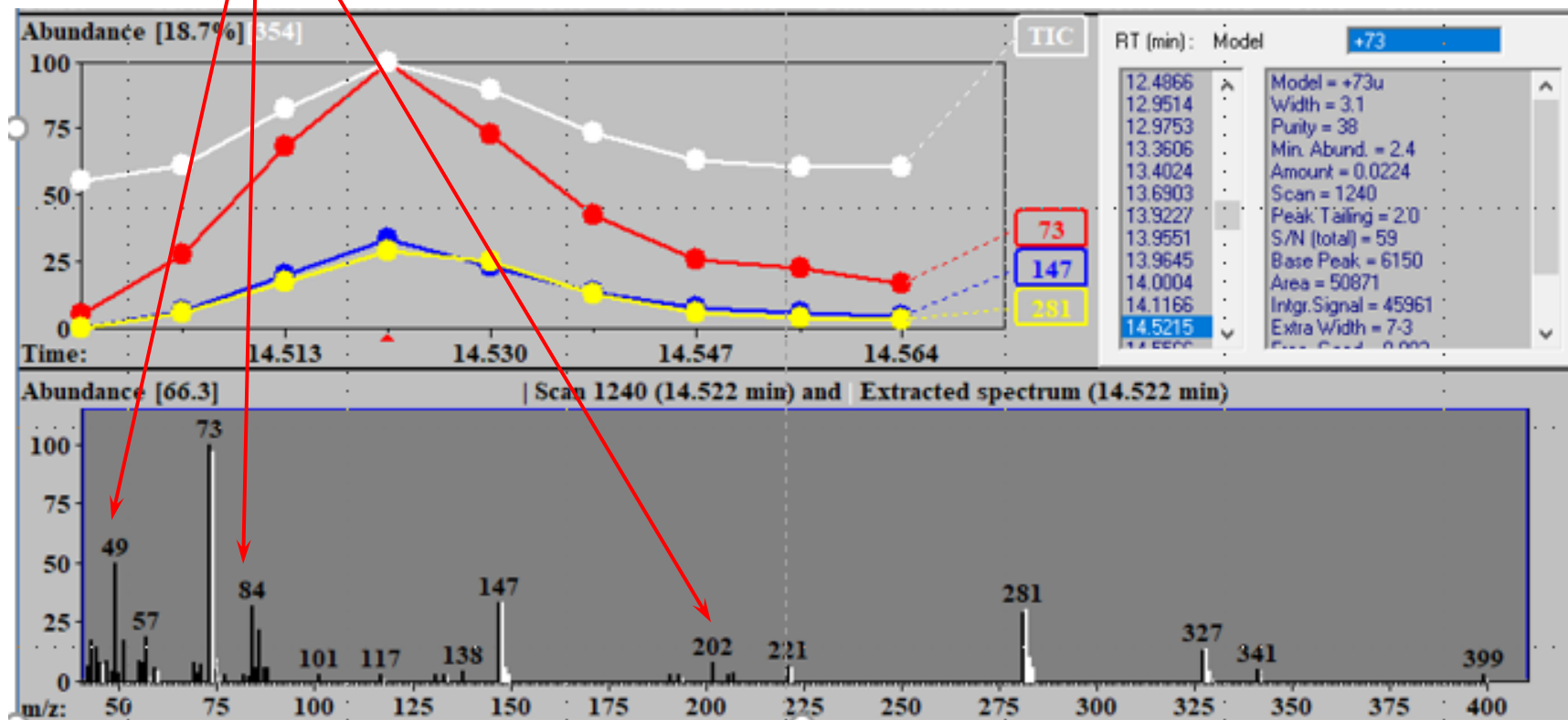
**RMB** Menu displayed by placing the **Pointer** on the Spectrum window and clicking the **RMB**

Which peaks will be displayed in the Spectrum window.



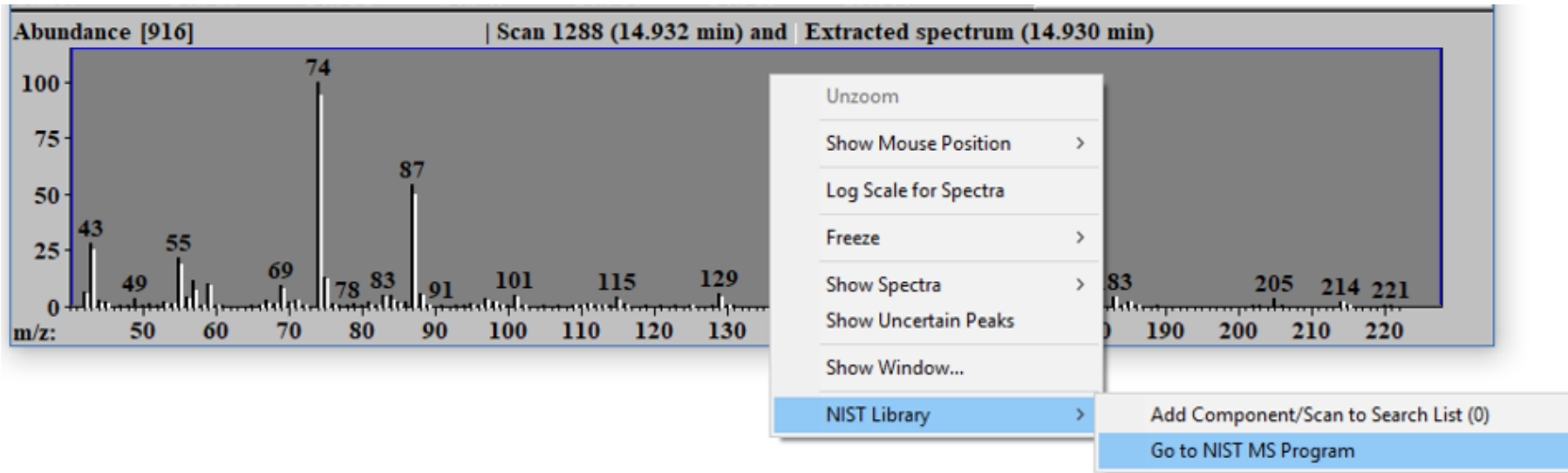
# Evaluating Deconvoluted Results (continued)

- Note black (uncorrected peak with background)
- White is spectrum corrected for back ground and all non tracking ions removed



# Sending Deconvoluted Spectra to NIST Search Program

- To send an individual mass spectrum to the NIST MS Search Program, click the **RMB** with the **Pointer** on the spectrum to display the **RMB** menu
- Select **Go to NIST MS Program**
- The spectrum will be sent to the NIST MS Search Program, if the Program is active; and, if not active, it will be started and the spectrum then sent
- If **Automation** is checked in the **Library Search Option's Search** tab, the search will occur automatically and the results will be displayed in the MS Search Program
- **Tip:** Can just LMB on chromatogram and obtain *manual spectrum* (no background correction) and send to MS Program for searching



# Returning to AMDIS Window after NIST Search

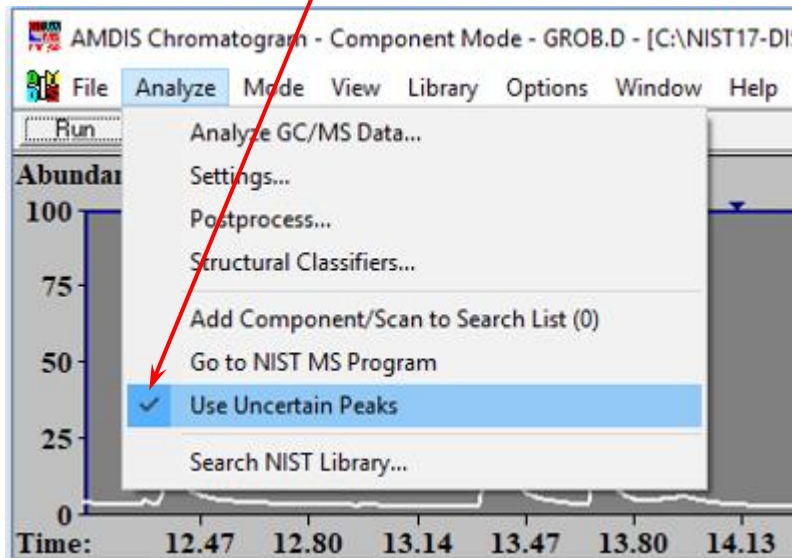
- After NIST search, return to AMDIS window by putting the **Pointer** on “Switch to Caller” button and click the **LMB**.

The screenshot displays the NIST MS Search 2.3 interface. The title bar indicates the search parameters: [Hybrid precursor = 137, Presearch Default - 100 spectra]. The menu bar includes File, Search, View, Tools, Options, Window, and Help. The toolbar contains various icons, including a red arrow pointing to the 'Switch to Caller' button (a left-pointing arrow). Below the toolbar, the main window shows a table of search results with columns for component number, source, and name. The table lists 17 components, including 'Component at scan 628 (9.122 min)' and several chemical names like 'Methyl 2-fluoro-3-[4-(piperidinocarbonyl)phenyl]benzoate'. To the right of the table, there are two spectral plots. The top plot shows a mass spectrum with peaks at m/z 54, 67, and 77. The bottom plot shows a mass spectrum with peaks at m/z 41, 44, 47, and 50. At the bottom of the interface, there is a bar chart showing the distribution of matches across a range of m/z values from 1000 to 500. The bar chart has a red bar at m/z 900 and several blue bars at lower m/z values. The bottom status bar shows the current component selected: 'Component at scan 628'.

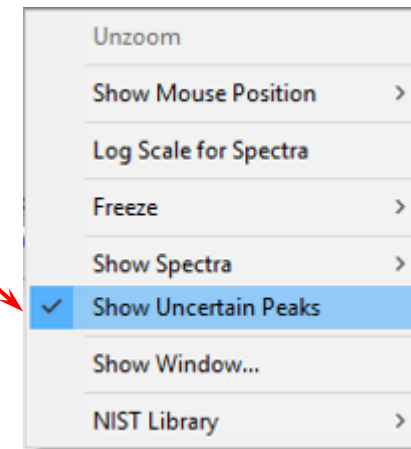
#	Src.	Name
1	A	Component at scan 628 (9.122 min) [Model = +99u] in C:\NIST17\AMDIS32\TUTORIAL\TEST.D\DA...
2	A	Component at scan 628 (9.122 min) [Model = +99u] in C:\NIST17\AMDIS32\TUTORIAL\TEST.D\DA...
3	A	Component at scan 628 (9.122 min) [Model = +99u] in C:\NIST17\AMDIS32\TUTORIAL\TEST.D\DA...
4	A	Component at scan 543 (8.500 min) [Model = +70u] in C:\NIST17\AMDIS32\TUTORIAL\TEST.D\DA...
5	L	Manual Component in C:\NIST17\AMDIS32\TUTORIAL\TEST.D\DATA.MS
6	L	Manual Component in C:\NIST17\AMDIS32\TUTORIAL\TEST.D\DATA.MS
7	L	Component at scan 432 (7.689 min) [Model = +99u] in C:\NIST17\AMDIS32\TUTORIAL\TEST.D\DA...
8	L	Methyl 2-fluoro-3-[4-(piperidinocarbonyl)phenyl]benzoate
9	L	2,4-Dihydroxybenzamide
10	L	2,4-Dihydroxybenzamide, 3TMS
11	L	2,4-Dihydroxybenzamide, 4TMS
12	L	2,4-Dihydroxybenzamide, diacetate
13	L	2,4-Dihydroxybenzamide, N-acetyl-, diacetate
14	L	2,6-Dihydroxy-3,4-dimethylpyridine
15	L	2,6-Dihydroxy-3,4-dimethylpyridine, 2TMS
16	L	2,6-Dihydroxy-3,4-dimethylpyridine, diacetate
17	L	4,4'-Dihydroxy-3,3'-dimethylbiphenyl-4,4'-dicarboxylic acid

# Uncertain Peaks, Dashed Lines, in Deconvoluted Spectrum

- Sometimes the AMDIS “decides” that some peaks “*might*” be associated with the deconvoluted spectrum, but it is not sure; you will need to change the basic settings if you want to use them
- These “*uncertain peaks*” are shown as dashed white lines in the spectrum
- To use them and send them for library searching, the Analyze settings have to be changed
- First, click the **RMB** with the **Pointer** on the spectrum to cause the display of the **RMB** menu and select **Show Uncertain Peaks**. Once selected, this will remain until changed.
- Then go to top of the **Analyze** menu, displayed from the Main Menu, and select **Use Uncertain Peaks**



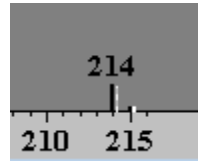
Analyze Menu



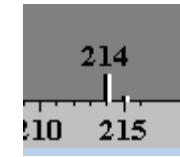
Right Mouse-button Menu  
with Pointer on Spectrum

# Avoiding Uncertain Peaks in a Spectrum

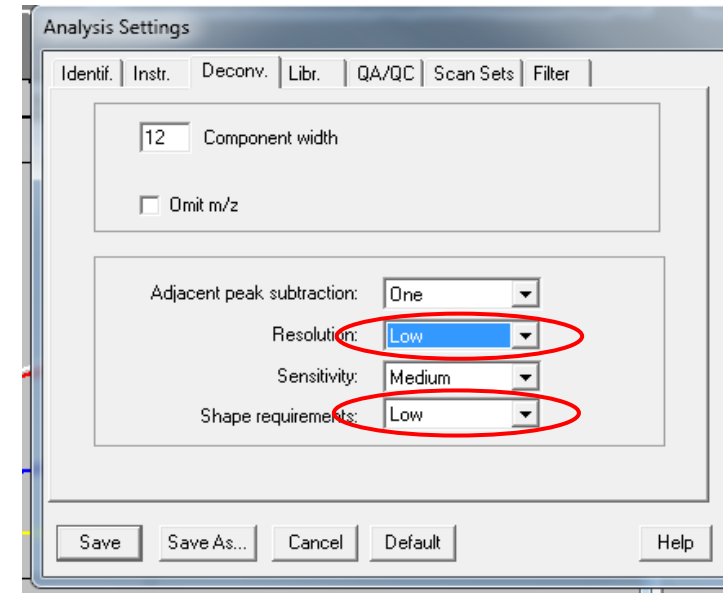
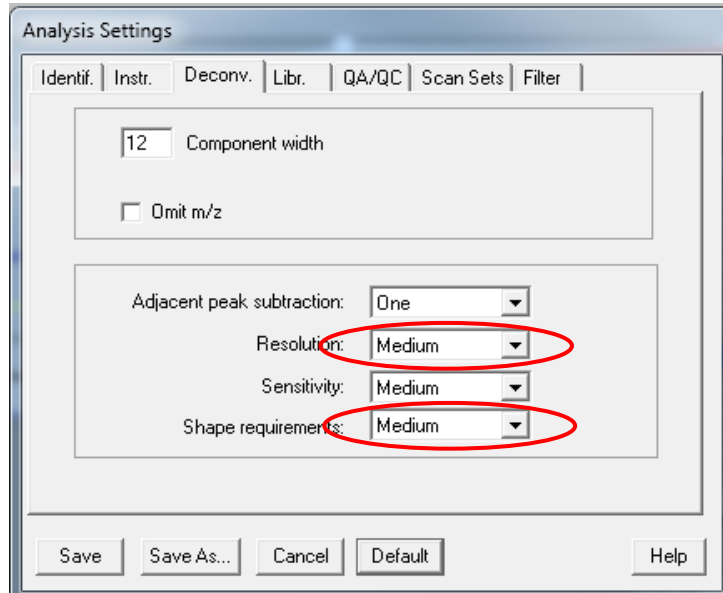
- Often uncertain peaks **can be avoided** by changing the default settings for Resolution and/or Shape Requirements in the **Analysis Settings** menu
- **Alert!** Internal library searches **do not use uncertain peaks**, so best results obtained by avoiding their formation!
- Of course, uncertain peaks **not** a concern with spectra obtained manually



Default Settings

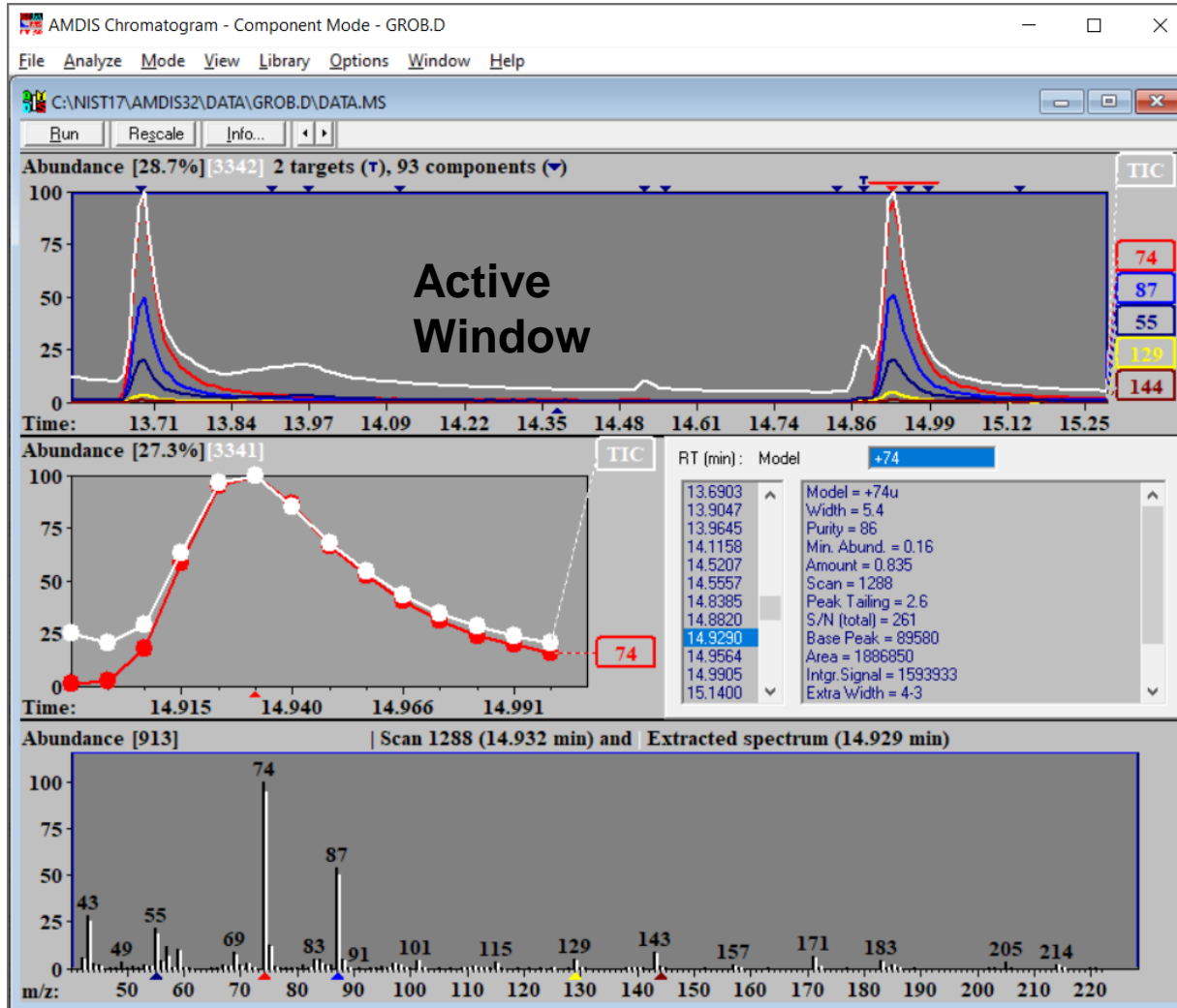


Modified Settings



# Plotting Single (or Extracted) Ion or (Mass) Chromatograms

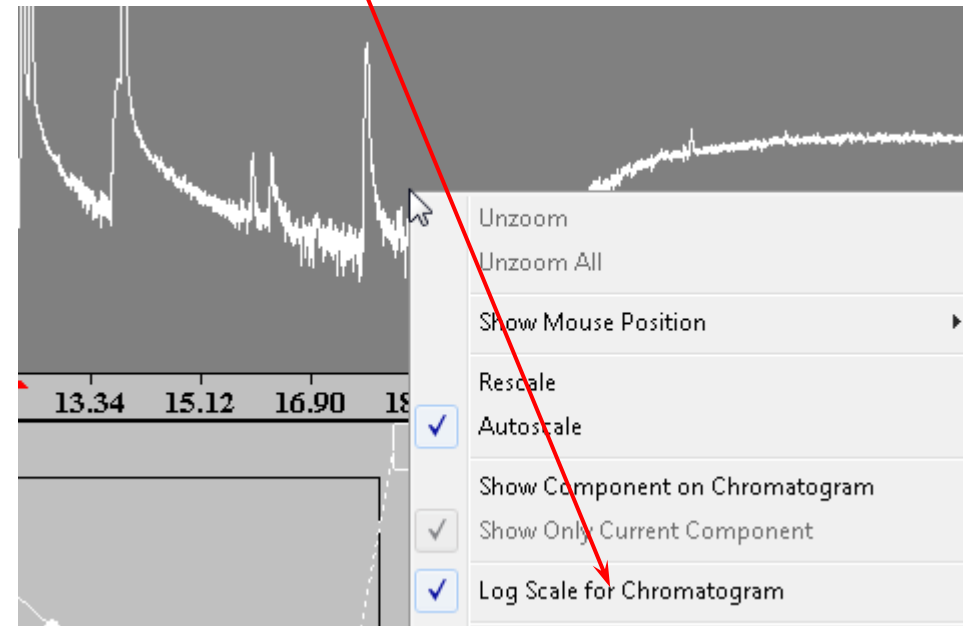
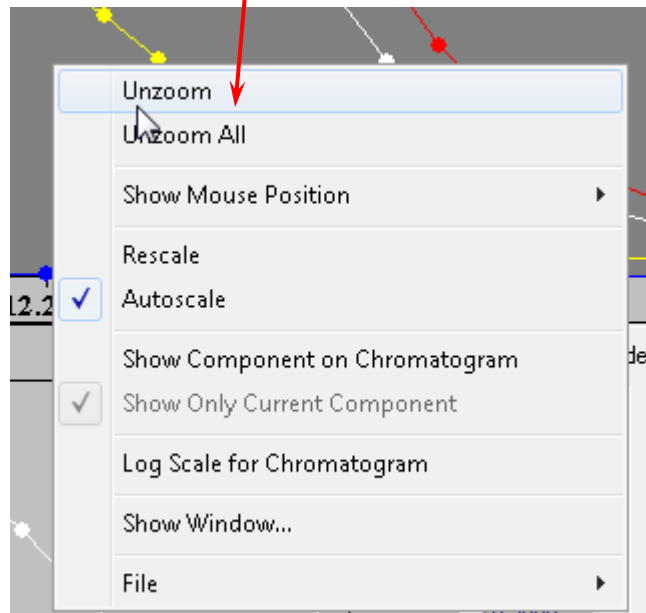
- To plot ion current vs. time (i.e., a mass chromatogram), just click the **LMB** with the **Pointer** on the peak representing the ion in the spectrum window, and the mass chromatogram will **immediately** be displayed in a different color in the active window. The intensity of the peak produced by the 1<sup>st</sup> selected ion is set to be 100%. If a subsequent ion is more abundant than that 1<sup>st</sup> selected ion, its plot will be off scale



- Either the chromatogram (top) window of the model (middle left) window can be the active window
- To select the active window, put the **Pointer** on the bar above the window and click the **LMB**.
- The active window is dark gray
- To delete that mass chromatogram, just click the **LMB** with the **Pointer** on its box to the right of the top chromatogram
- Tip:** the **TIC** (total ion chromatogram) box can be toggled off for easier viewing of low intensity mass chromatograms **or** use log scale as describe on next slide

# Expanding Chromatograms or Plotting in Log Scale to See Small Peaks

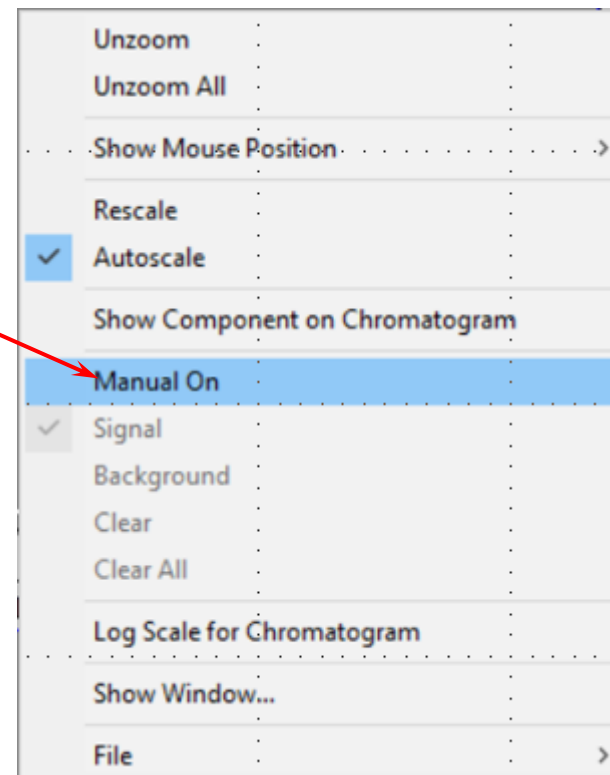
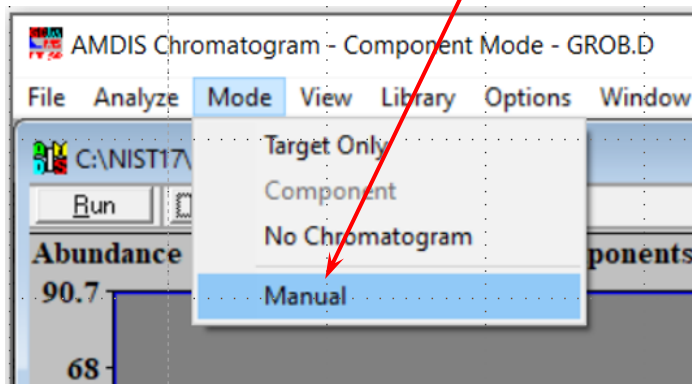
- To expand the chromatogram or spectrum, just hold down the **LMB** and drag (Drag-n-Drop)
- To unzoom, right click in the window and select **Unzoom** or **Unzoom All** from **RMB** menu
- Another way to see small peaks is to put Mouse-pointer on the chromatogram (or spectrum) window, click the **RMB**, and select **Log Scale for Chromatogram** or **Log Scale for Spectra** from the **RMB** menu





# Manually Processing File in AMDIS

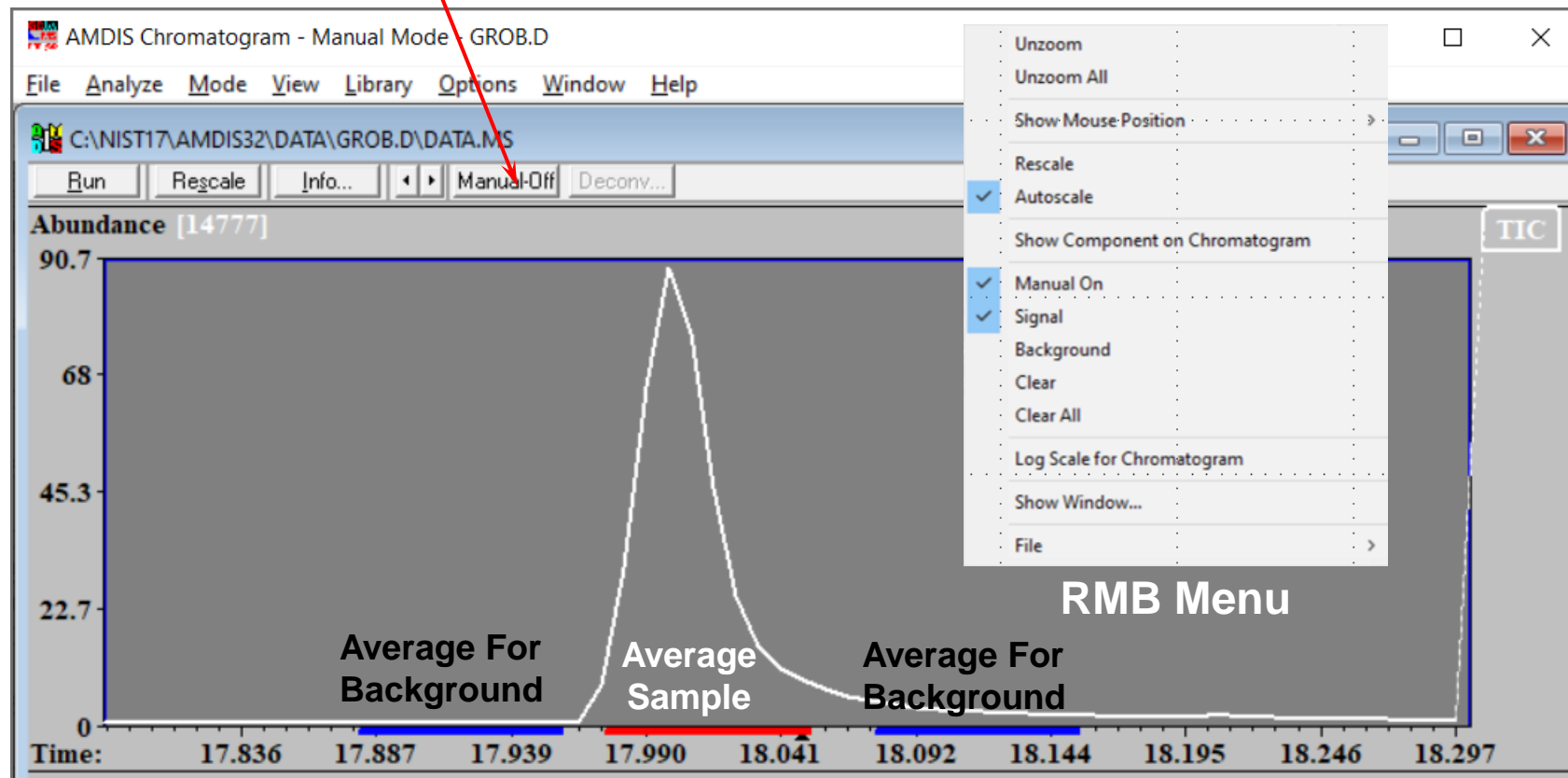
- If you just want a non-deconvoluted (uncorrected) spectrum of the background, click the **LMB** with the **Pointer** on the scan of interest, it can then be sent to the MS Search Program and searched against the NIST and/or other libraries
- AMDIS can produce a manual background-subtracted spectrum, typical of other MS software
- Often helpful for broad or peaks with excessive tailing
- First, go to top bar and select **Manual** from the **Mode** menu on the Main Menu bar
- Second, display the **RMB** menu and select **Manual On**



**RMB** Menu displayed by putting **Pointer** on Chromatogram window and clicking the **RMB**

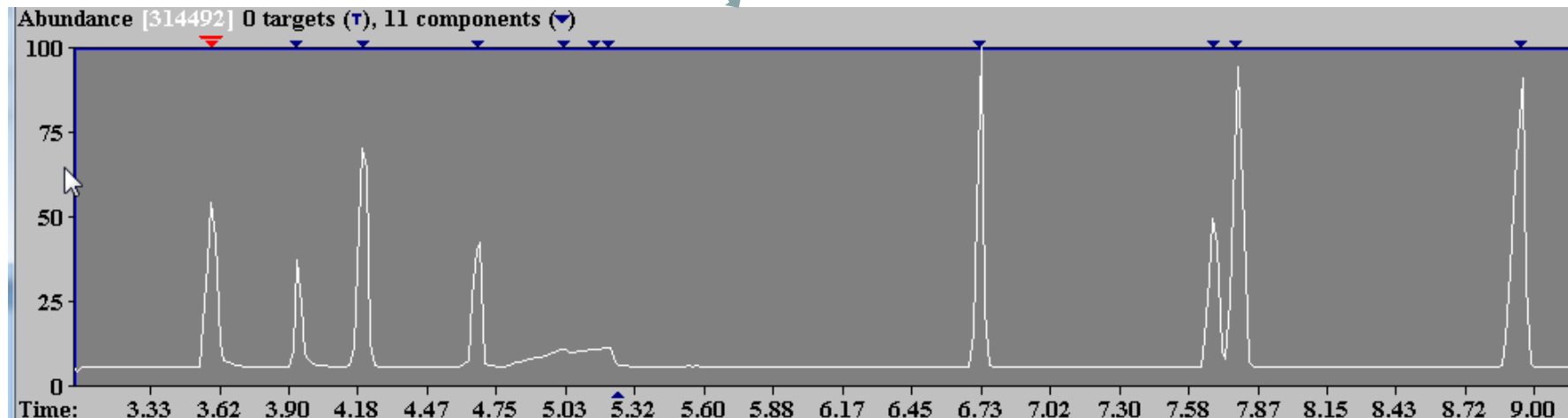
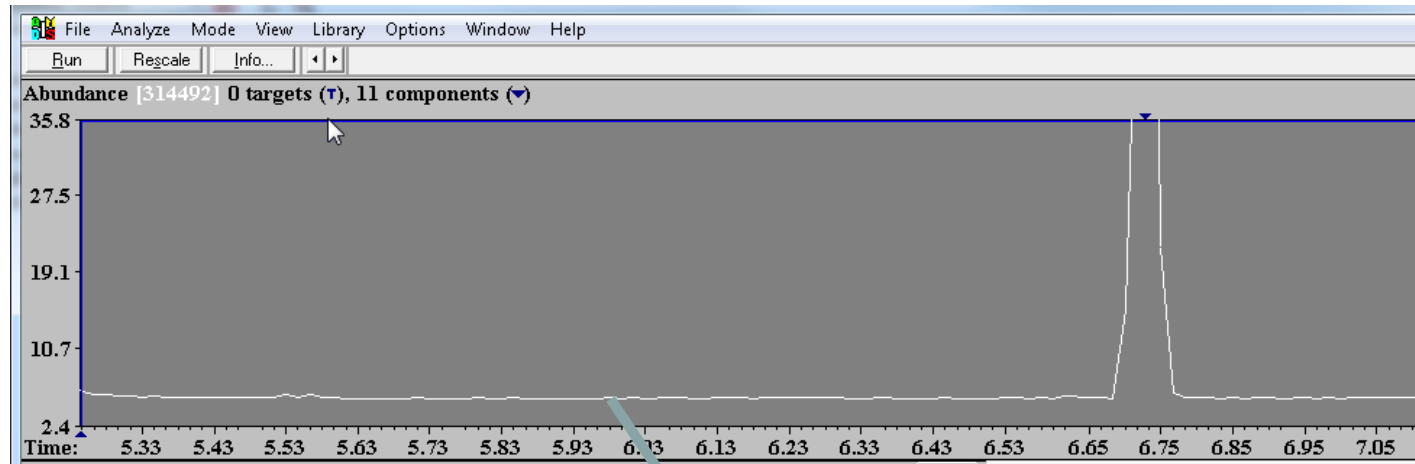
# Manually Processing File in AMDIS (continued)

- From **RMB** menu displayed with **Pointer** on the chromatogram window select (**one at a time**) in a sequence, **Signal** (one or more ranges to average) and **background** (one or more ranges)
- The manually background spectrum is shown in the spectrum window, (bottom of the two displayed windows; the model window (middle), used in deconvolution, is no longer present)
- The chromatogram window can be unzoomed using the **RMB** menu; but, to zoom requires **LMB** clicking on the **Manual Off** button above the chromatogram turning it to **Manual On**
- The spectrum obtained can be sent to MS Search using the **RMB** menu



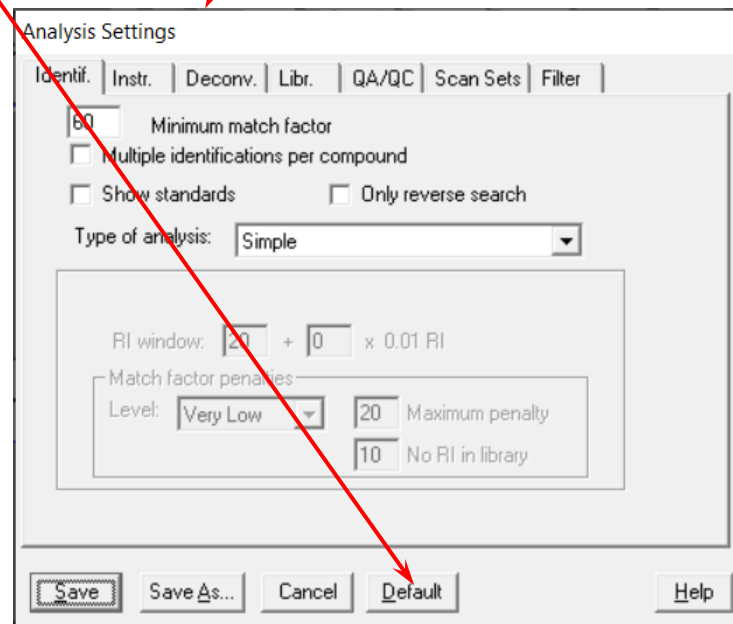
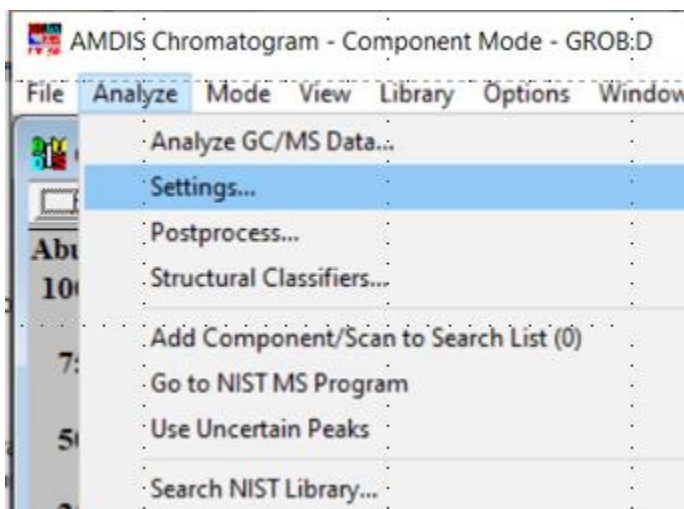
## Adjusting Parameters for Optimized Peak Detection

- When trying to determine proper parameters, expand the chromatogram to only show the most difficult areas
- Change to parameter and only the area shown will be reanalyzed
- After getting all the parameters as desired, then show the whole chromatogram and **Run** (Reanalyze) again
- This will greatly speed the process!
- **Tip:** NIST wrote 3 part series on suggested parameters for deconvolution<sup>23</sup>



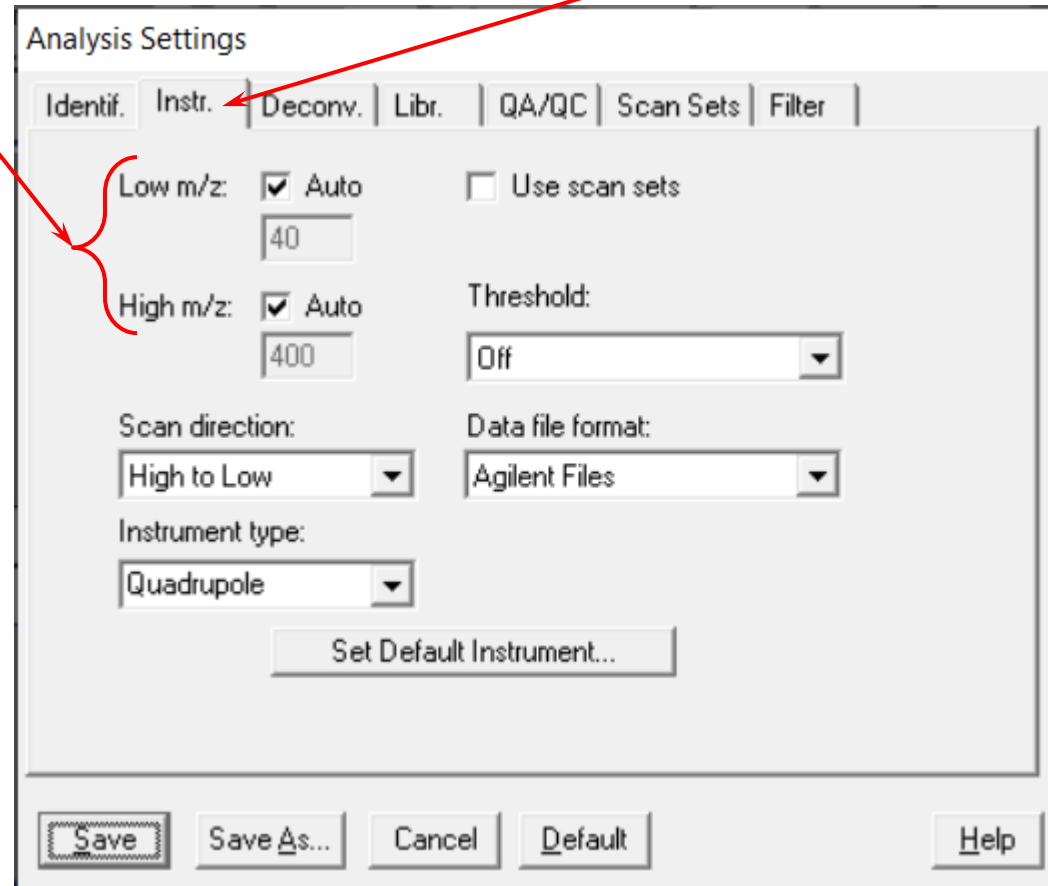
# Minimizing Marking Components in Chromatogram

- The “multi-marking” of **Components** due to noise or instrument scanning irregularities can be annoying
- Almost all instruments under varying conditions tend to have this problem
- This can be minimized by adjusting some parameters in the **Analysis Settings** dialog box
- Note that there are multiple tabs with many parameters in this dialog box
- It is easy to restore the program's **Default** settings



# Minimizing Marking Components in Chromatogram (cont'd)

- Setup the processing parameters based on the instrument and its scan function (**Instr** tab)
- Can set the low and high m/z manually, or just automatically use those determined by AMDIS from the file



The screenshot shows the 'Analysis Settings' dialog box with the 'Instr.' tab selected. The 'Instr.' tab is highlighted with a red arrow from the text 'Instr tab' in the list above. A red bracket highlights the 'Low m/z' and 'High m/z' settings, with a red arrow pointing from the text 'Can set the low and high m/z manually' in the list above. The 'Low m/z' is set to 'Auto' with a value of 40. The 'High m/z' is set to 'Auto' with a value of 400. Other settings include 'Use scan sets' (unchecked), 'Threshold' (Off), 'Scan direction' (High to Low), 'Data file format' (Agilent Files), and 'Instrument type' (Quadrupole). A 'Set Default Instrument...' button is located below the 'Instrument type' dropdown. At the bottom of the dialog are buttons for 'Save', 'Save As...', 'Cancel', 'Default', and 'Help'.

Analysis Settings

Identif. **Instr.** Deconv. Libr. QA/QC Scan Sets Filter

Low m/z:  Auto  Use scan sets  
40

High m/z:  Auto Threshold:  
400 Off

Scan direction: Data file format:  
High to Low Agilent Files

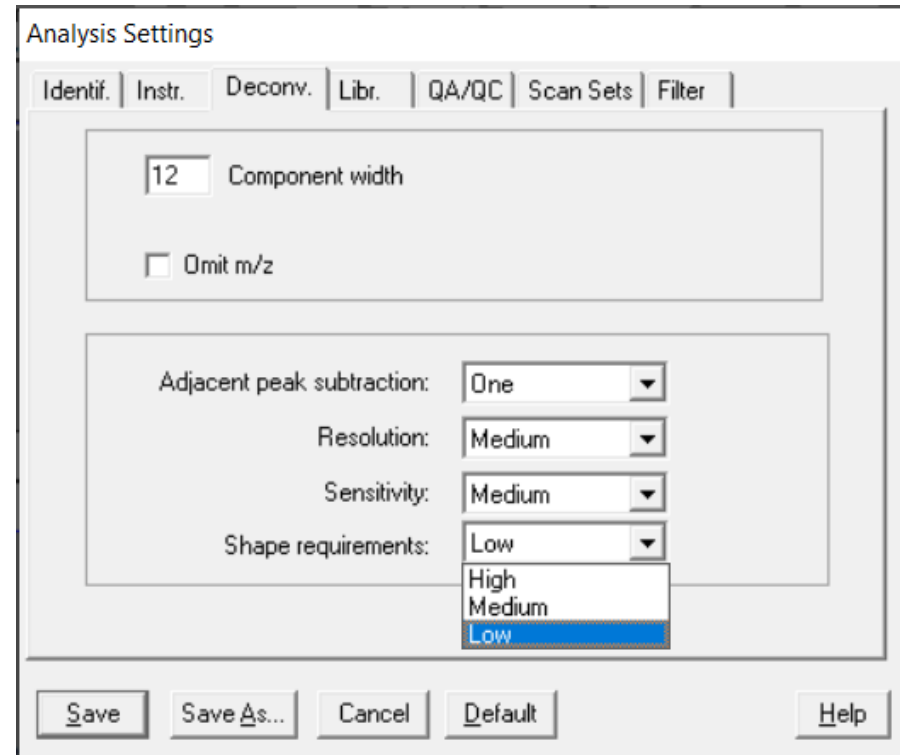
Instrument type:  
Quadrupole

Set Default Instrument...

Save Save As... Cancel Default Help

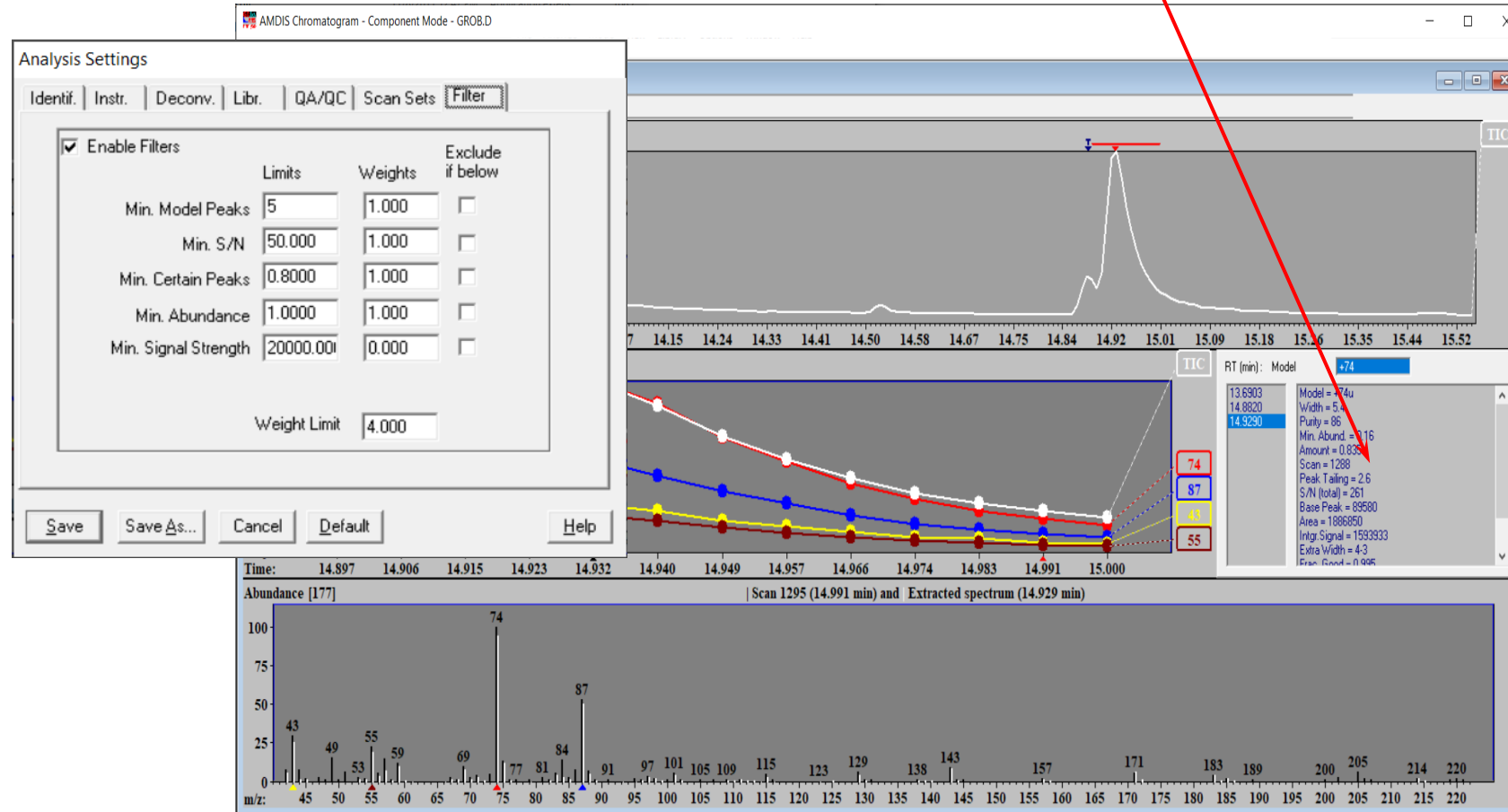
# Minimizing Marking Components in Chromatogram (cont'd)

- The **Deconvolution** tab can be set to get rid of some peaks
- In particular, for tailing peaks, might want to set the **Shape** requirements to Low
- The default for **Shape** requirements is Medium
- The values show below are the **Defaults**
- In general, the **Filter** tab (see slides 36-37 of this handout) usually minimizes the multi-marking of **Components**



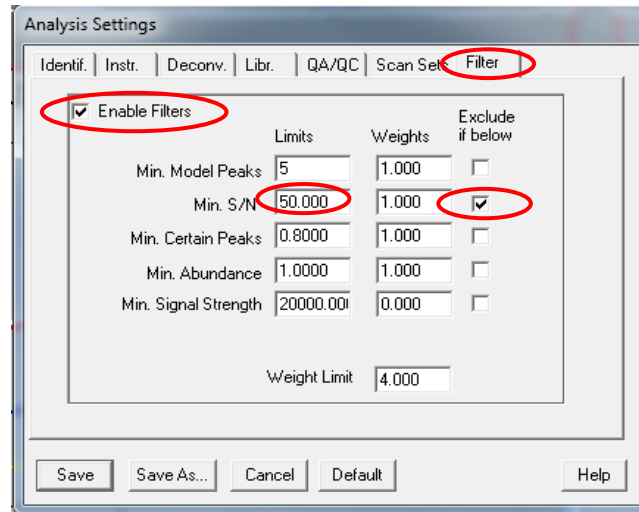
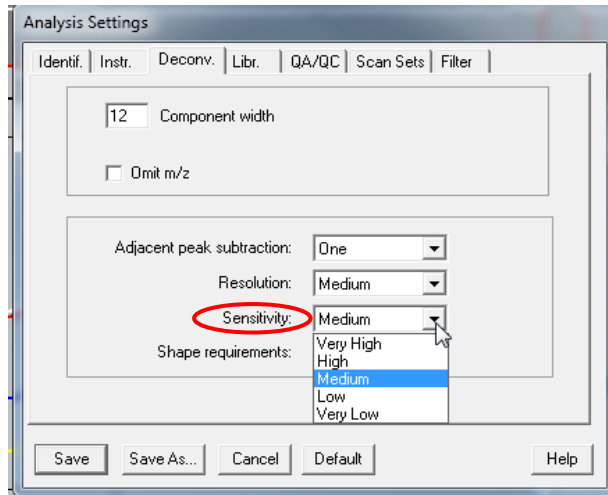
## Minimizing Marking Components in Chromatogram (cont'd)

- The default filter settings are shown below; the default settings do NOT have the **Enable Filters** check box selected; unless checked, the fields are grayed
- The values associated with a particular **Component** can be viewed in the window next to the Model (middle-left) window
- Looking at these values gives an idea of how to limit parameters to minimize the marking of **Components**



# Minimizing Number of Components Detected

- The number of components detected can be minimized by many approaches
- One approach is to change the **Sensitivity** setting in the **Analysis Settings**
- Another approach is to use the filter settings and **Exclude if below** a specified **Min S/N**
- To optimized, expand the chromatogram and find a suitably sized “small” peak to find an appropriate S/N to mark the number of components detected



16.618	16.626	16.634	16.643
RT (min): 1 of 2 models +159, -253			
15.1713	Model = +159u, -253u		
15.3064	Width = 3.7		
15.3608	Purity = 27		
15.4333	Min. Abund. = 0.95		
15.4621	Amount = 0.0938		
15.4885	Scan = 788		
15.7076	Peak Tailing = 1.2		
15.7768	S/N (total) = 185		
15.8065	Base Peak = 9045		
15.8090	Area = 651915		
15.8584	Intgr. Signal = 593783		
15.9119	Extra Width = 2.0		
15.9367	Frac. Good = 0.930		
15.9605	Models 6: 159 241 173 128 256 242		



## Problem No Chromatogram/Spectrum Window!

1. AMDIS initially designed to only yield report
2. If you inadvertently select “Go to Results”
3. Then “Analyze”
4. You could see “NO TARGET CHEMICALS FOUND!”
5. Select “Confirm” button to return to chromatogram/spectrum window normally used for qualitative analyses

