

# Biomarker Identification in Nonalcoholic Fatty Liver Disease via GC/MS

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## Introduction

- Non-alcoholic fatty liver disease (NAFLD) is a major medical problem in developed countries
- NAFLD is associated with:
  - Increase obesity and insulin resistance
  - Metabolic syndrome
- NAFLD is influenced by lifestyle:
  - Exercise
  - Diet
- Research Samples & Study
  - 168 Rodent samples (original sample set, Pegasus 4D Study)
  - Investigate two critical factors: 1) Increased fructose consumption and 2) Inadequate Cu intake
  - Complementary investigation: Utilize a more sensitive instrument (20 fg) with extended linear dynamic range ( $10^5$ ) to further characterize the samples
- Analysis Objective: Identify potential NAFLD biomarkers in pooled rodent samples using high performance, benchtop GC-TOFMS (Figure 1)



Figure 1. Pegasus® BT GC-TOFMS.

## Workflow & Methods

- Sample preparation
  - Dry: 1) Speed Vac & 2) Lyophilizer
  - Derivatize: 1) MEOX (30  $\mu$ L, 20mg/mL) & 2) MTBSTFA (20  $\mu$ L)
  - (For each step, heat at 60 °C for 1 hr)
- Analyze derivatized samples using high performance GC-TOFMS, ChromaTOF® brand software, and statistical processing software
- Instrument Parameters

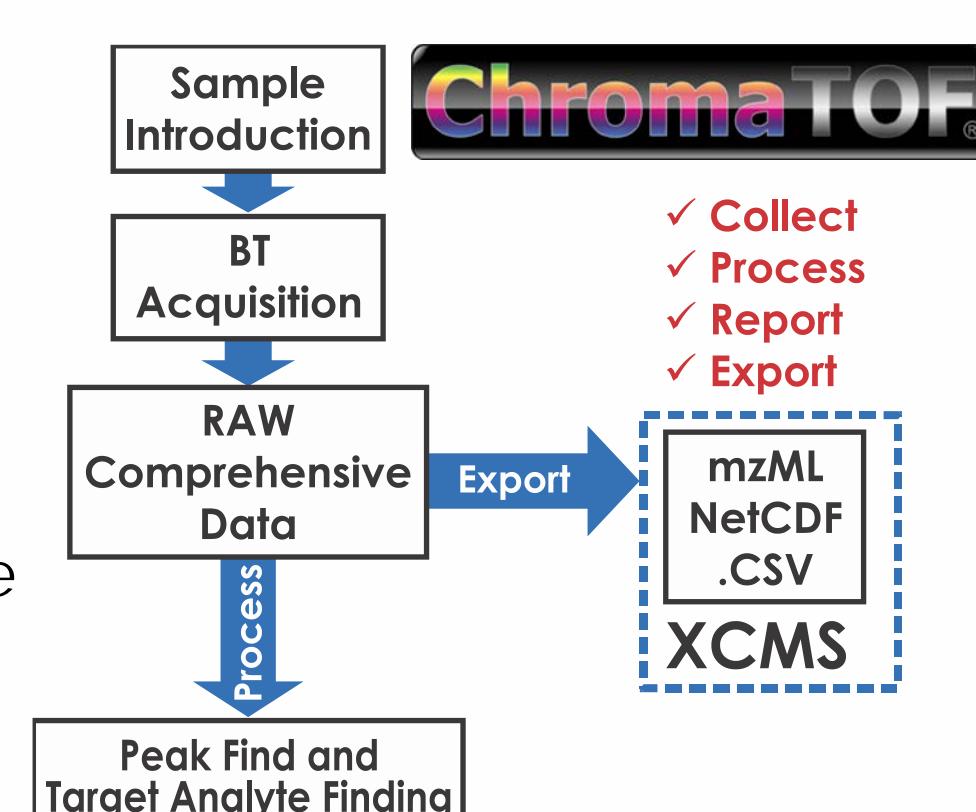


Figure 2. Analysis Workflow.

Table 1. GC-TOFMS Parameters

Gas Chromatograph	Agilent 7890 with LECO L-Pal3 Autosampler
Injection	1 $\mu$ L, Split 20:1; 270 °C
Carrier Gas	He @ 0.8 mL/min, Constant Flow
Column	Rxi 5 Si MS, 20 m x 0.18 mm i.d. x 0.18 $\mu$ m (Restek, Bellefonte, PA, USA)
Temperature Program	60 °C (0.50 min), ramped 36 °C/min to 320 °C (3 min)
Mass Spectrometer	LECO Pegasus BT
Ion Source Temperature	250 °C
Ionization Mode	EI
Mass Range (m/z)	45-600
Acquisition Rate	10 spectra/s

## GC-TOFMS Results

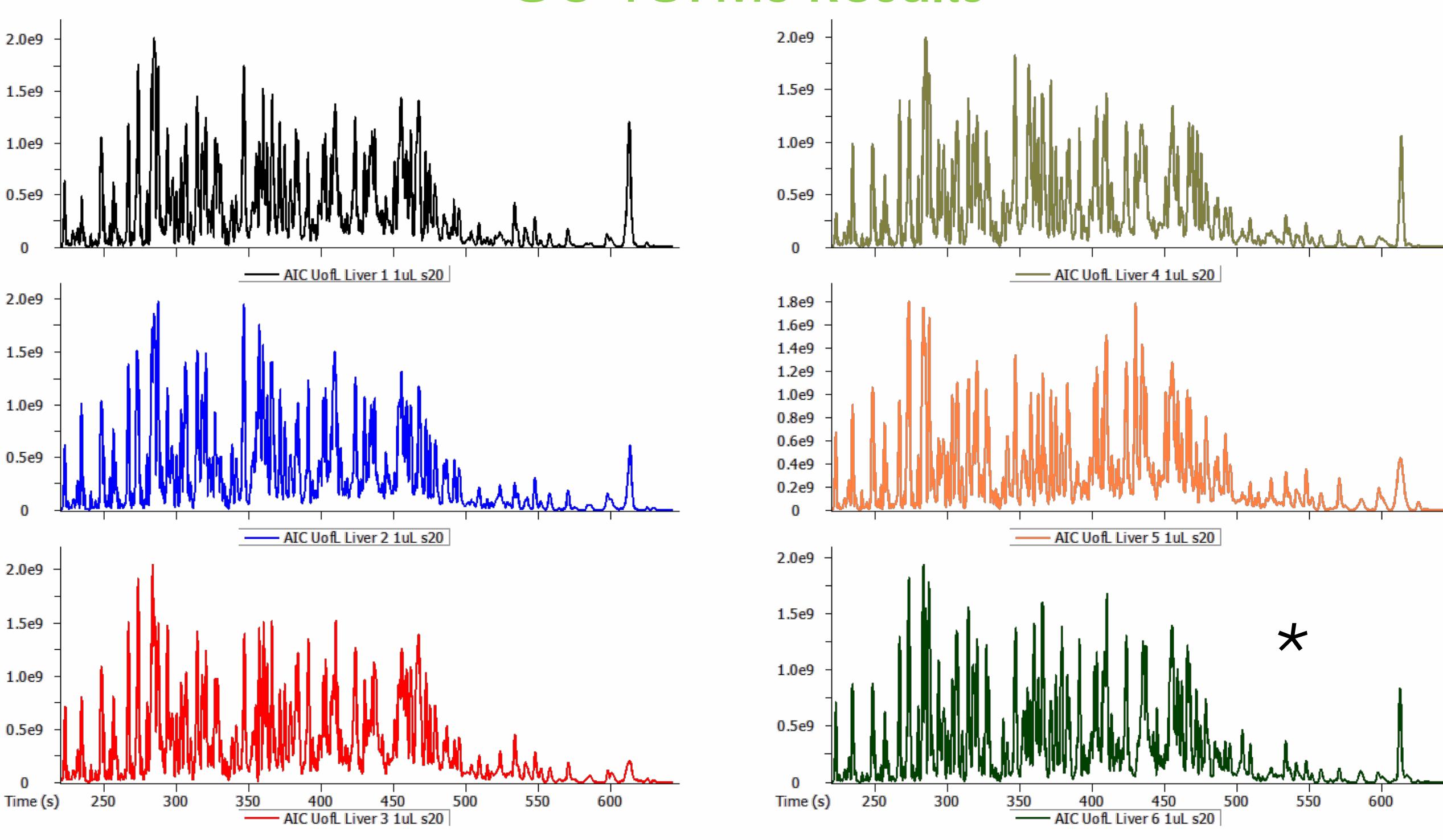


Figure 3. AICs for 6 pooled liver samples.

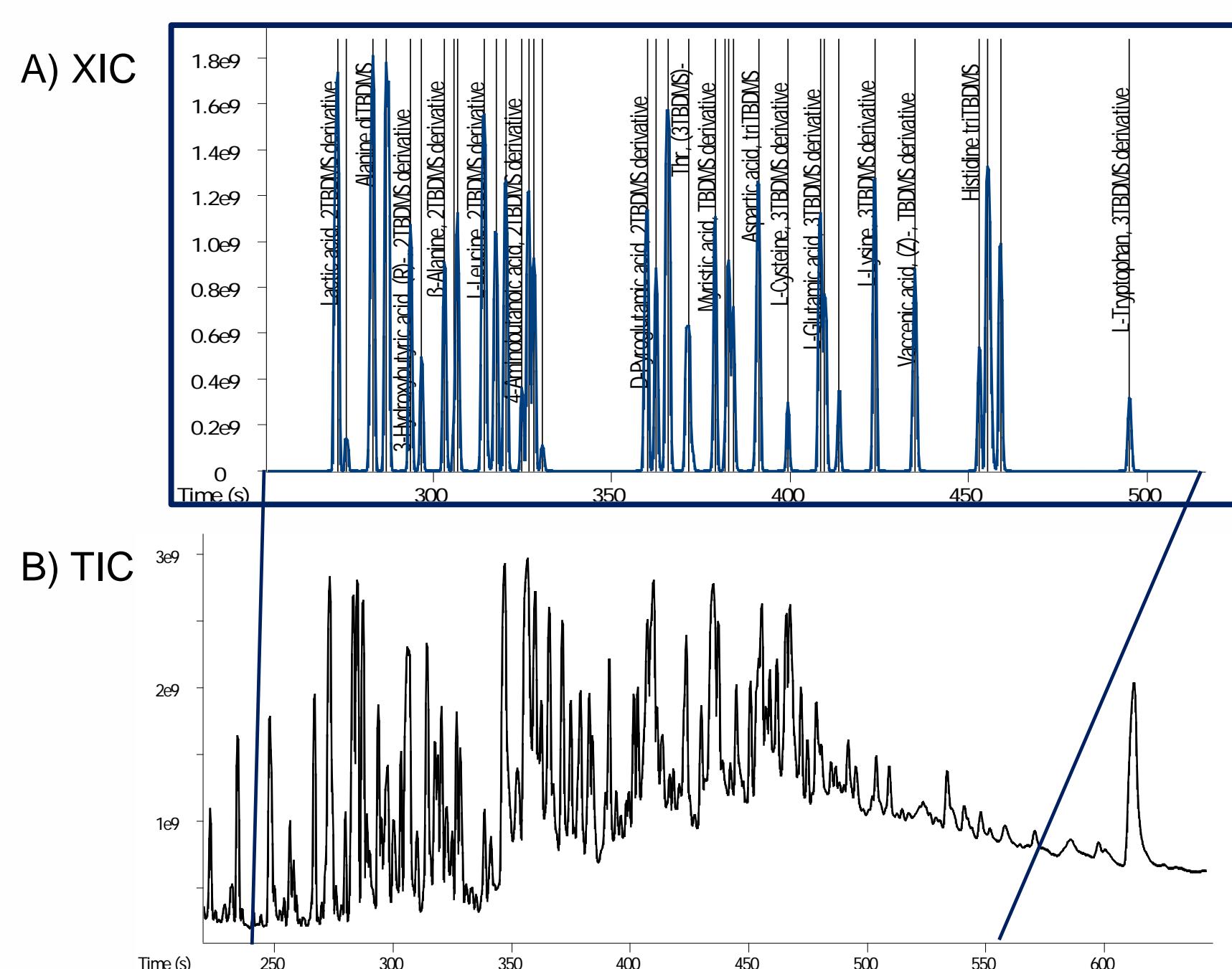


Figure 4. A) eXtracted Ion Chromatogram (XIC) & B) TIC of liver sample 6\*.

Table 2. Representative analytes in liver sample 6

Name	Formula	R.T. (s)	Area	Similarity	Name	Formula	R.T. (s)	Area	Similarity
Lactic acid, 2TBDMs derivative	C <sub>3</sub> H <sub>6</sub> O <sub>2</sub> Si	273	3889358872	865	L-Serine, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	366	1804043501	910
Glycolic acid, 2TBDMs derivative	C <sub>3</sub> H <sub>6</sub> NO <sub>2</sub> Si	276	392513491	865	Threonine, 2TBDMs	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	376	1907273568	870
Alanine, 2TBDMs	C <sub>3</sub> H <sub>6</sub> NO <sub>2</sub> Si	283	2627670348	894	Mycistic acid, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	379	2526194161	906
Glycine, 2TBDMs derivative	C <sub>3</sub> H <sub>6</sub> NO <sub>2</sub> Si	287	2093903802	909	Alanylglucine, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	382	246632471	802
3-Hydroxybutyric acid, (R)-, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	294	1338064233	911	L-Phenylalanine, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	383	1491608293	913
L-2-Aminobutyric acid, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	297	980488165	889	Malic acid, 3TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	384	133022456	852
8-Alanine, 2TBDMs derivative	C <sub>3</sub> H <sub>6</sub> NO <sub>2</sub> Si	303	8117768633	856	Aspartic acid, 3TBDMs	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	391	1444900099	897
Dimethylpentobarital	C <sub>11</sub> H <sub>20</sub> N <sub>2</sub> Si	306	44555160	900	L-Cysteine, 3TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	399	523411147	943
Valine, 2TBDMs	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	307	2085644362	864	L-Glutamic acid, 3TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	408	1539429832	919
L-Leucine, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	314	3168854906	937	Alanine, 3TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	410	1491608293	883
Glutamine, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	318	2512779336	922	Alanylglutamate, 3TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	414	1044773406	873
Isoleucine, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	320	2056141263	943	L-Lysine, 3TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	424	1125874542	917
4-Aminobutanic acid, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	325	460048677	831	Vaccenic acid, (Z), 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	435	4023899325	837
Uracil, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	327	211731605	929	Histidine triTBDMs	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	453	1428415311	895
L-Proline, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	328	2171908625	918	Arachidonic Acid, 3TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	455	1921897209	899
Itaconic acid, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	331	211731605	864	L-Tyrosine, 3TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	459	1990728852	922
D-Pyroglutamic acid, 2TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	360	2010302971	854	Uric acid, 4TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	488	239470552	898
Met, (2TBDMs)	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	363	795167581	868	L-Tryptophan, 3TBDMs derivative	C <sub>5</sub> H <sub>10</sub> NO <sub>2</sub> Si	495	1189857046	866

Ave. = 886/1000

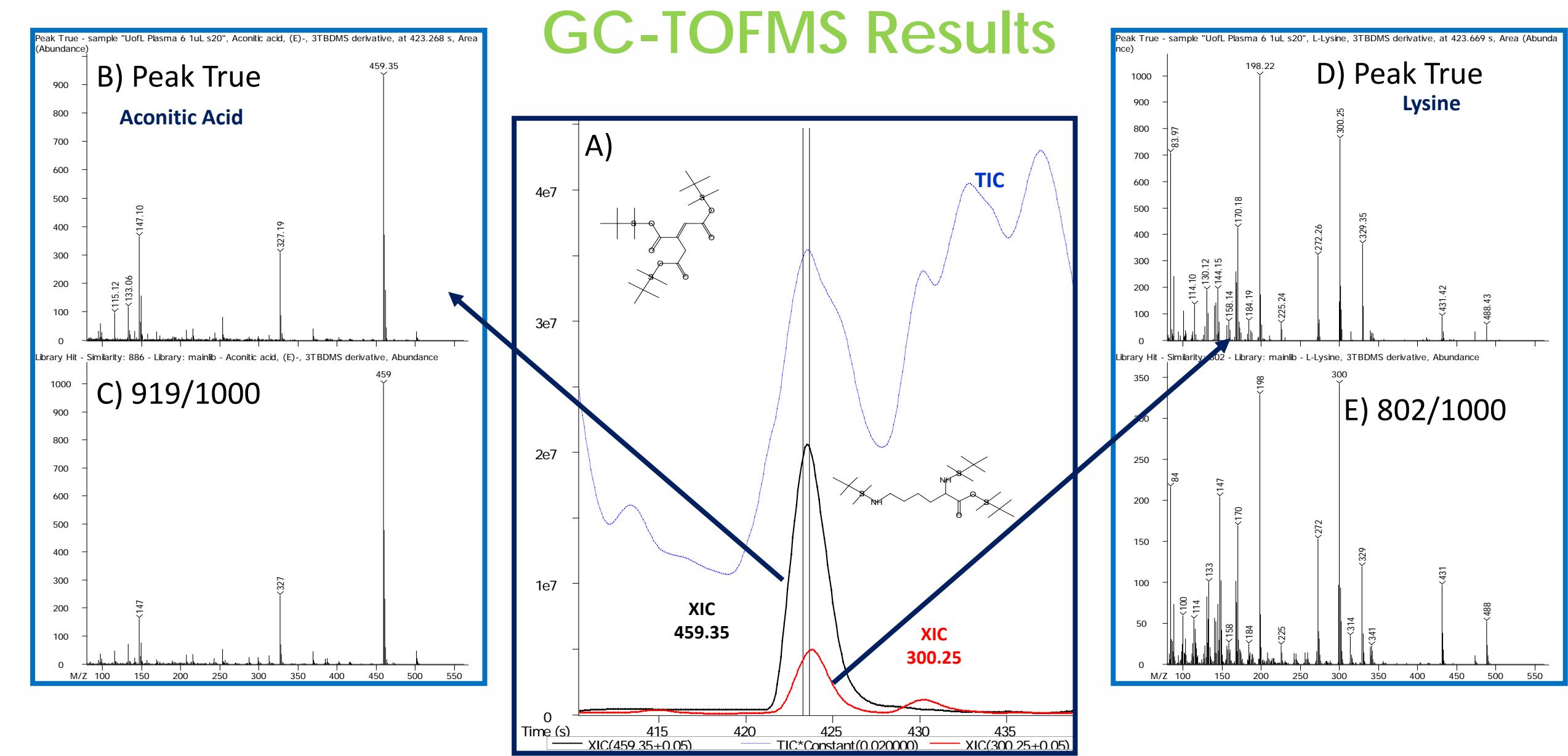


Figure 5. A) ChromaTOF deconvolution results - liver sample 6. Peak True (deconvoluted) and library spectra for aconitic acid (B,C) and lysine (D,E).

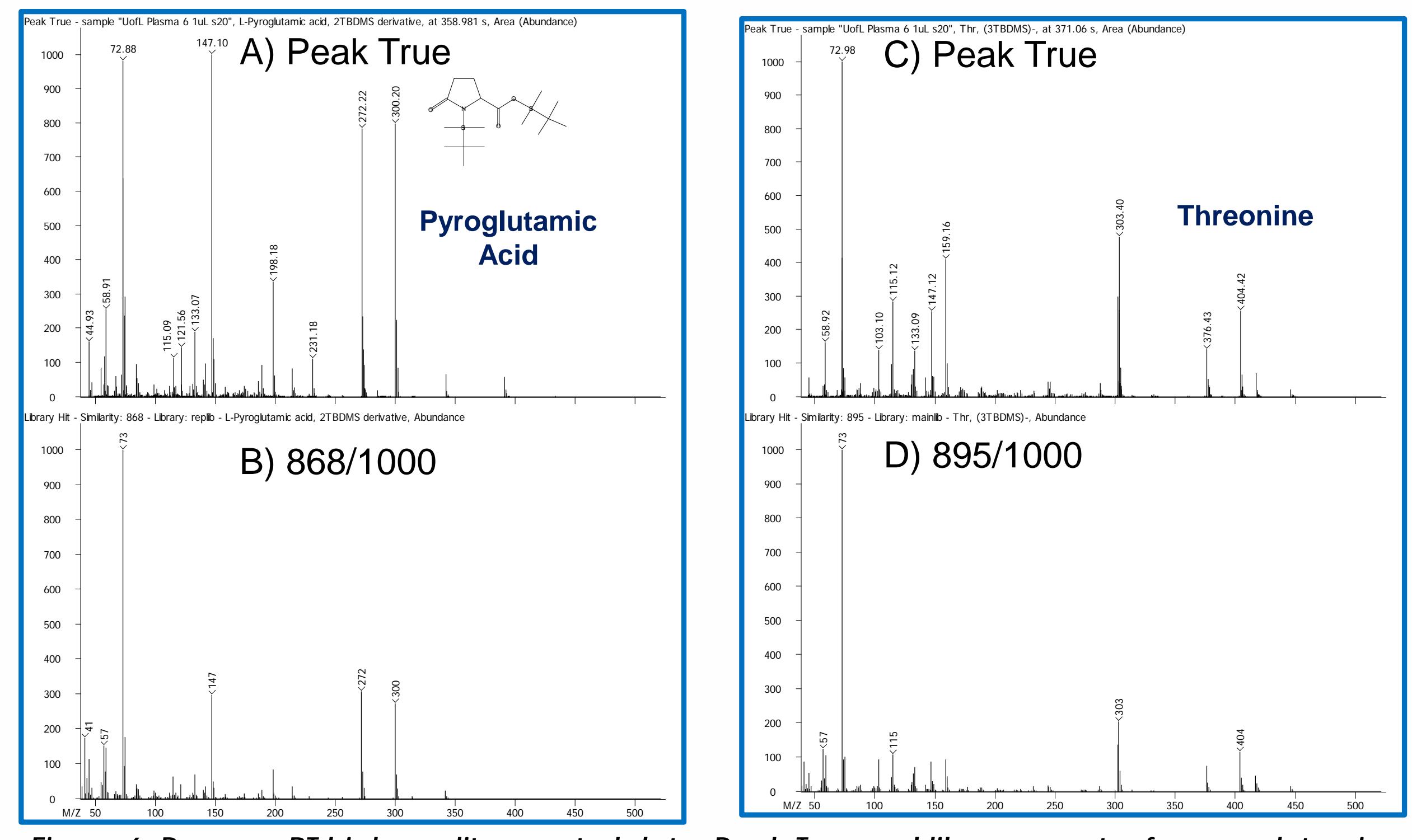


Figure 6. Pegasus BT high quality spectral data: Peak True and library spectra for pyroglutamic acid (A,B) and threonine (C,D).

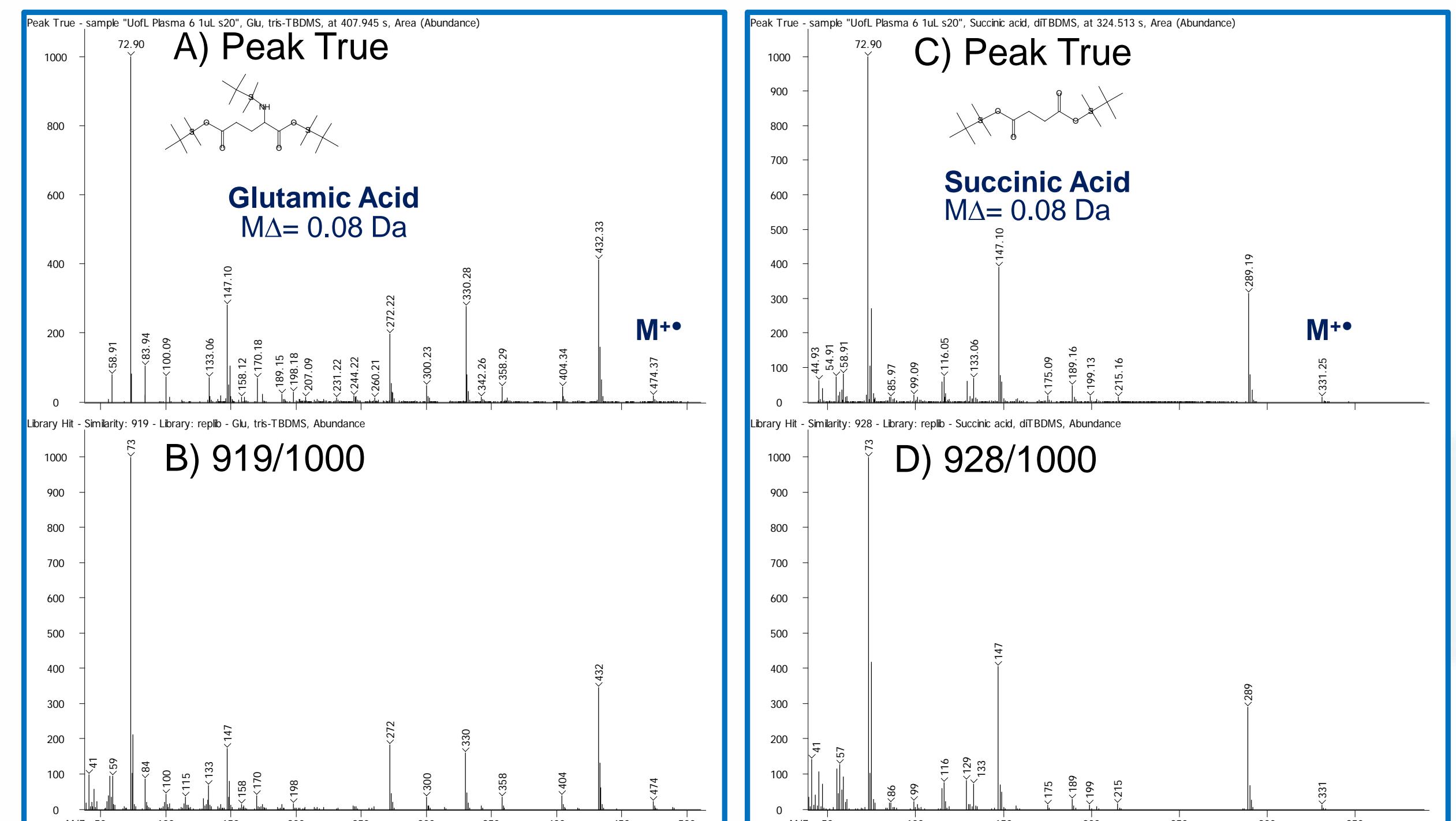


Figure 7. Pegasus BT high performance spectral data with mass delta values <0.10 Da): Peak True and library spectra for glutamic acid (A,B) and succinic acid (C,D).

## Pegasus BT Data Export and Statistical Analysis

- Raw data was exported (netCDF) using ChromaTOF (Figure 8)
- Data was uploaded and processed using XCMS online ([xcmsonline.scripps.edu](http://xcmsonline.scripps.edu))
- PCA and Cloud Plot data (Figure 9) were used to find features that differentiated samples
- Potential biomarker metabolites for NAFLD were confidently identified using ChromaTOF (Figure 10)
- Potential biomarkers characterized included: Amino acids (e.g., histidine, alanine, valine, etc.), fatty acids (e.g., arachidonic acid) and misc. compounds (e.g., uridine, 7-methylxanthine)

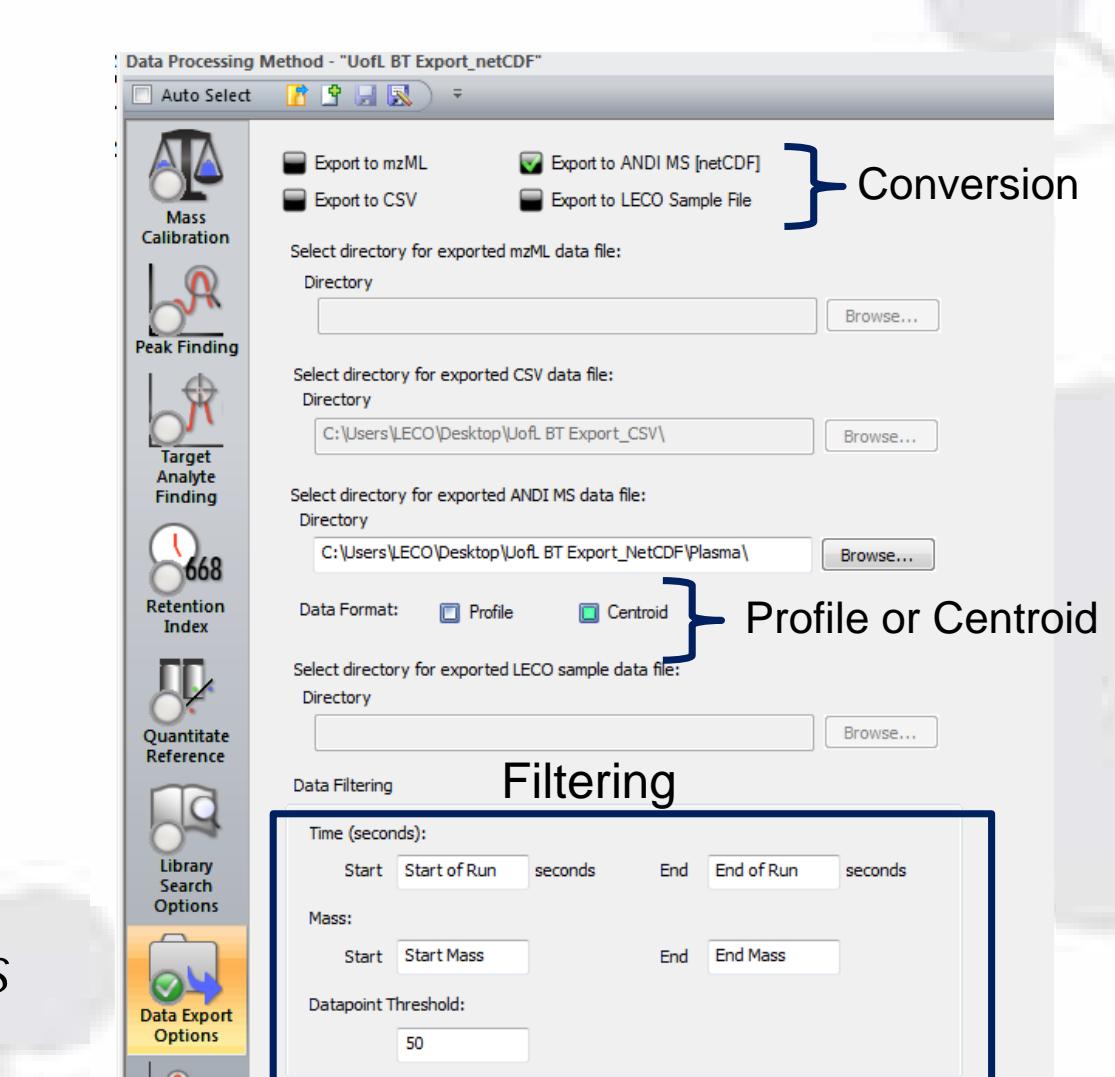


Figure 8. ChromaTOF raw data export feature.



Figure 9. XCMS online Cloud Plot (m/z vs retention time) showing a metabolite feature with a p-value of 4.84e-4, retention time of 7.55 min. and EIC m/z = 440.396.

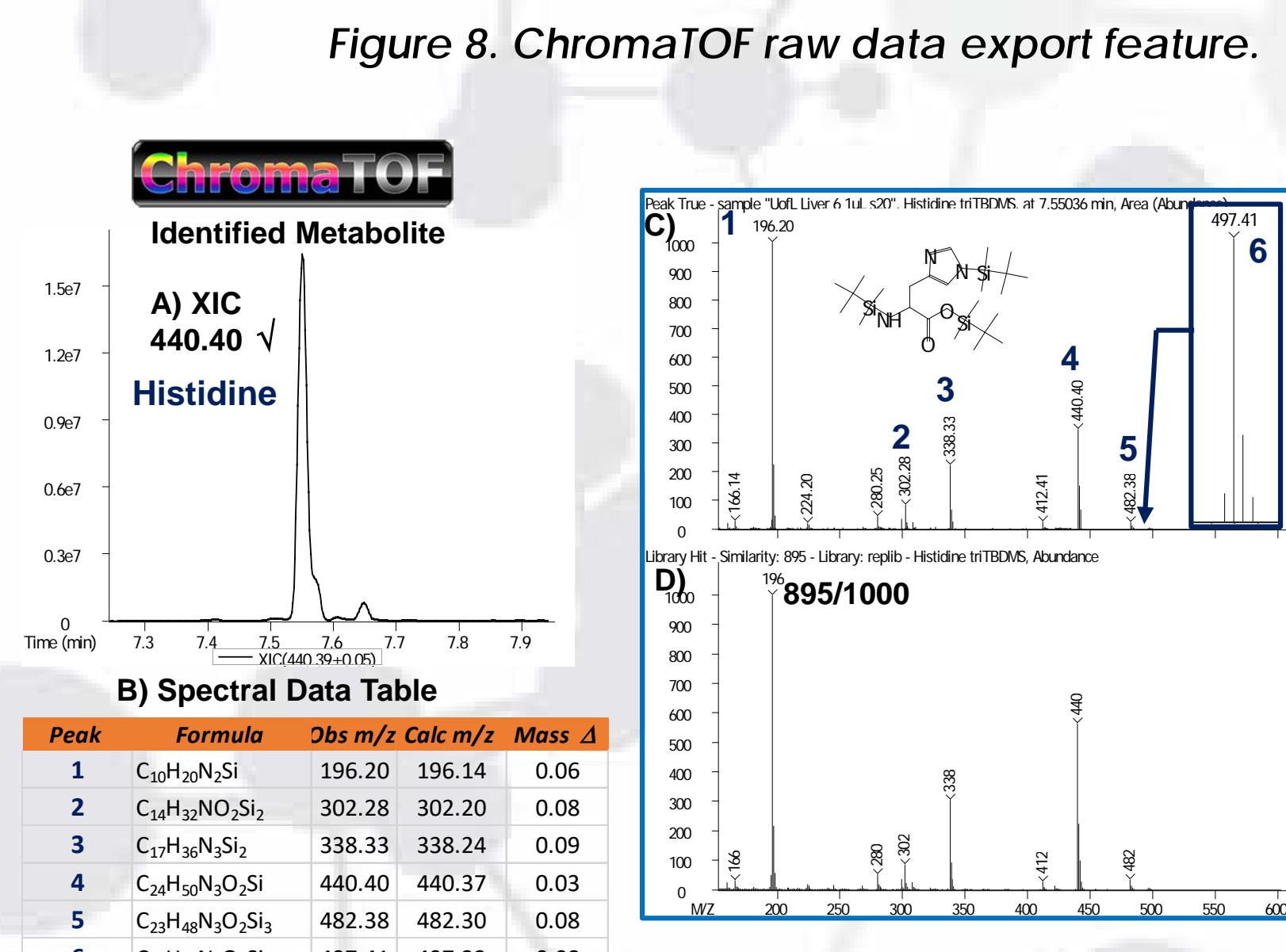


Figure 10. ChromaTOF - A) XIC 440.40, B) Generated formulas for fragment and molecular ions, C,D) Peak True and library match data for histidine.

## Summary

- The Pegasus BT instrument produced high quality data
- This data was exported into statistical processing software for sample differentiation and feature selection
- ChromaTOF brand software was used to fully characterize metabolites using spectral similarity searches against large, well-established databases and formula determinations using data with mass deltas<0.10 Da

