

AN1308: SEC-MALS to determine chitosan molar mass

Introduction

Chitin is one of the most abundant biopolymers on earth (the other is cellulose). Chitin, or poly-N-acetyl-glucosamine, is the major polymer in the exoskeleton of marine arthropods and can also be found in fungi and yeasts. It is used in water treatment, photographic emulsions and dyeing improvement of synthetic fibers and fabrics

Chitosan is deacetylated chitin. It can be obtained from shrimp or crab shells. Its applications vary from the therapeutic, such as wound healing, to cosmetics and dietary supplements.

For many of these applications, it is useful to fully characterize the molar mass moments and distributions of the chitosan products. Size-exclusion chromatography in combination with multi-angle light scattering detection (SEC-MALS) provides an easy method to obtain these properties in an absolute manner, free of molecular references. In this note, we describe the results for two chitosan samples analyzed by SEC-MALS.

Materials and Methods

A Wyatt DAWN[®] MALS detector and Optilab[®] differential refractive index (dRI) detector were plumbed downstream of the GPC column. Data collection and analysis were performed in the ASTRA[®] software using empirically-determined differential refractive index increments (dn/dc). Polymer molar mass M was calculated at each elution volume using signals from the two detectors.

The differential refractive index is a property of the polymer/solvent system. It is measured by injecting a series of known concentrations into the Optilab, using solutions that are often prepared by the dry weight method for accuracy, and the SEC mobile phase as solvent. ASTRA collects and analyzes the results to determine dn/dc.

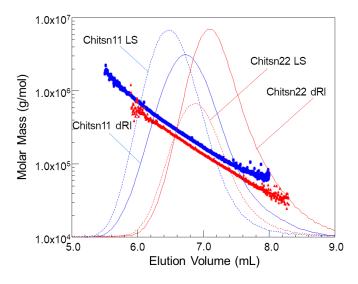


Figure 1. Molar Mass vs. elution volume plots superimposed over chromatograms for two chitosans.

Results and Discussion

The molar masses of two chitosan samples were plotted as a function of elution volume in Figure 1. The molar mass decreases logarithmically, indicating that chromatographic conditions were optimal. The offset in elution volume for a given molar mass may be a result of conformation (short-chain branching, SCB) or non-ideal analyte-column interaction. SCB may be further investigated by making use of simultaneous size (radius of gyration, R_a) and molar mass analysis by MALS.

A cumulative molar mass distribution plot, depicted in Figure 2, clearly differentiates the two chitosan samples. ASTRA software can also report weight fractions above, below, or between the molar masses of interest. As an example, the weight fraction of molar masses below 50 kDa and above 500 kDa for these samples are given in the table in Figure 2. These calculations and the cumulative molar mass distribution plot are ideal for quality control applications.

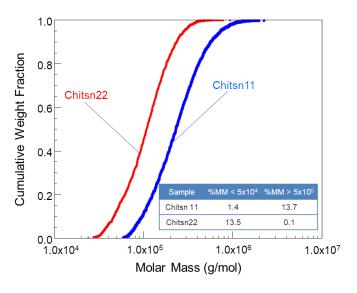
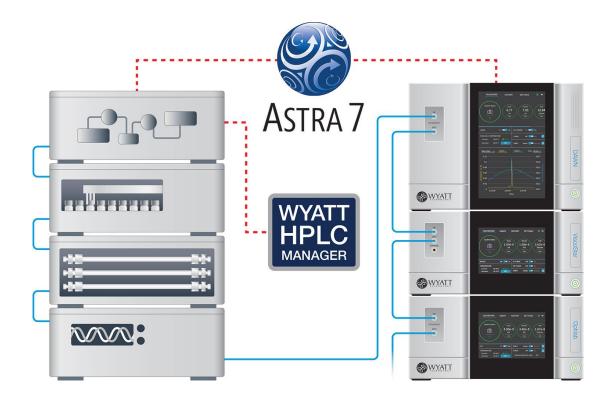


Figure 2. Cumulative molar mass distribution plot of two chitosan samples with quite different spans.

Conclusions

The results described herein show that MALS detection combined with SEC provides an indispensable tool for biopolymer characterization. Absolute molar mass and molar mass distributions can be readily obtained without the use of any standards or empirical relations. For chitosan-based products SEC-MALS simplifies QC and routine analyses while enhancing in-depth analysis with absolute measurements of size and conformation as well as molar mass.





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