

CHROMATOGRAPHIC SEPARATION AND PHOTODIODE ARRAY IDENTIFICATION OF SYNTHETIC INDUSTRIAL DYES IN FOODS, BEVERAGES, OVER THE COUNTER (OTC) DRUGS, AND COSMETICS

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INTRODUCTION

Food dye ingredients are often in the news due to concerns regarding neurobehavioral impacts on children, particularly hyperactivity, inattentiveness, restlessness, and other behavioral problems. As a result, some US states, such as California, prohibit the sale of products that contain synthetic food dyes in schools, while other states, such as West Virginia, have moved a step further by prohibiting synthetic dye containing food products throughout the state. In addition to neurobehavioral concerns, Red # 3, also known as erythrosine, was shown in a 2022 US Food and Drug Administration (FDA) petition to cause cancer in male laboratory rats. With this data, the FDA revoked the authorization for use of Red #3 in consumer products, issuing a complete phase out of the ingredient across the US by 2028. The FDA works to continuously monitor adulterated or misbranded products, and notably, color additive violations are a common reason for import refusals of food and cosmetic products offered for entry to the US.

In this poster, the Alliance™ iS HPLC System with Photodiode Array (PDA) Detector is shown to separate synthetic food dyes formulated in a variety of foods, beverages, OTC drugs and cosmetics. Three individual wavelengths were extracted from one HPLC injection to quantify yellow, red, blue, and green synthetic dyes. The PDA software determined chromatographic peak purity by UV spectral analysis, and identified dyes according to an Empower™ 3 Software, PDA Library.

METHODS

LC System:	Alliance™ iS HPLC System with Photodiode Array (PDA) Detector, Software version 1.4.0.
Column:	XBridge™ Premier BEH™ C18 2.5 µm, 4.6 x 150 mm, p/n: 186009849
Column temperature:	40 °C
Sample temperature:	20 °C
Injection volume:	30 µL
Flow rate:	1.6 mL/min
Needle wash:	50/50 Methanol/Water
3D Wavelengths:	200 – 800 nm
Resolution:	1 nm
Data rate:	10 Hz
Extracted channels:	455 nm (Yellow dyes) 520 nm (Red dyes) 628 nm (Blue and Green dyes)
CDS:	Empower™ Software, Version 3.8.0
Mobile phase A:	10 mM Ammonium acetate pH 7.0
Mobile phase B:	Methanol
Mobile phase C:	Acetonitrile

**Sample preparation:**  
Yellow #5, Yellow #6, Blue #2, Red #40, Green S, Carmoisine, Green #3, Blue #1, Red #3, and Patent Blue V food dye certified reference standards were obtained from Sigma-Aldrich. Standards were solubilized in water at 1.0 mg/mL. A standard stock mixture was prepared to contain each dye at 0.1 mg/mL. From the mixture, serial dilutions were prepared in dye-free sports drink to equal HPLC column sample loads between 0.003 ug to 0.375 ug of each dye. From the linearity injections, a PDA Library was created in the Empower 3 Software. For solid samples, one serving was dissolved in a respective volume of water (i.e. 10 – 200 mL). Each extraction was sonicated or stirred until the matrices were dissolved. The extractions were centrifuged for 10 minutes at 15,000 RPM to separate insoluble material, and the supernatant used for analysis. For analysis of sports drinks, each beverage was diluted 1:10 in water prior to HPLC injection to reduce turbidity.

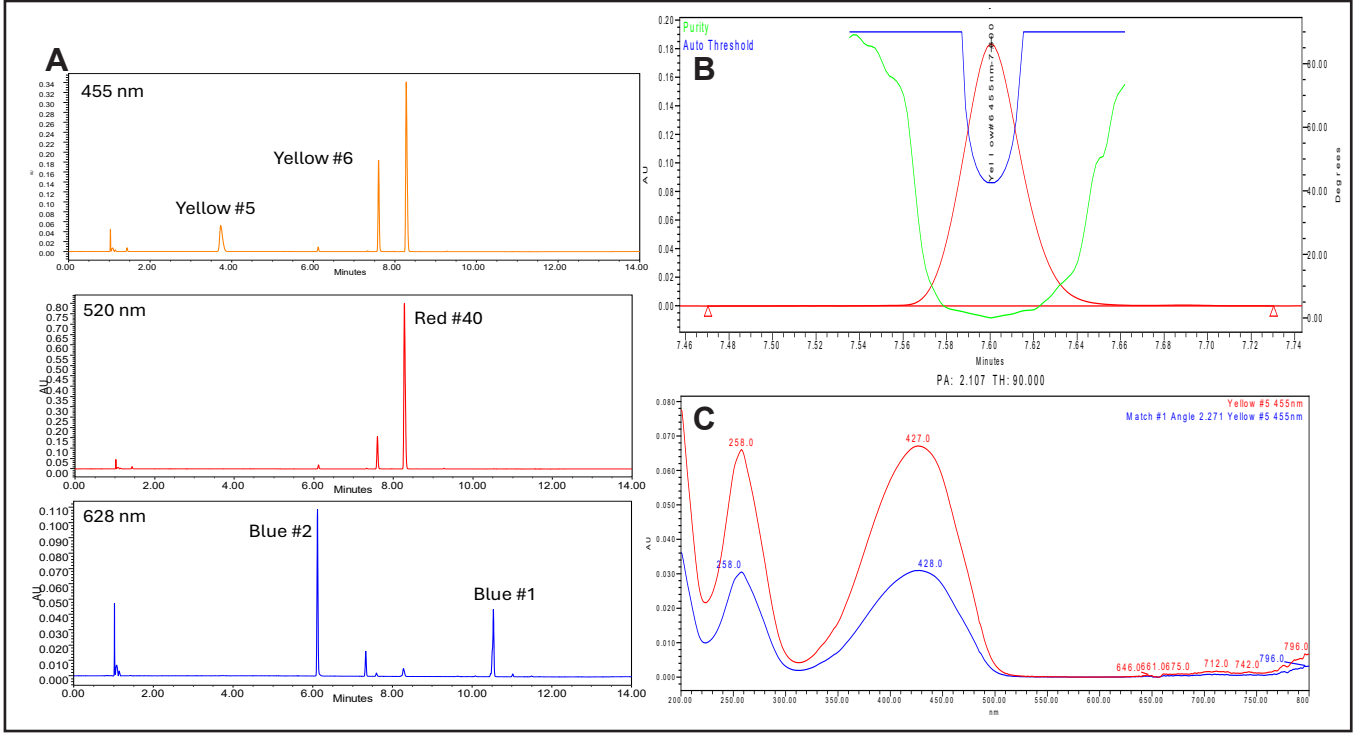
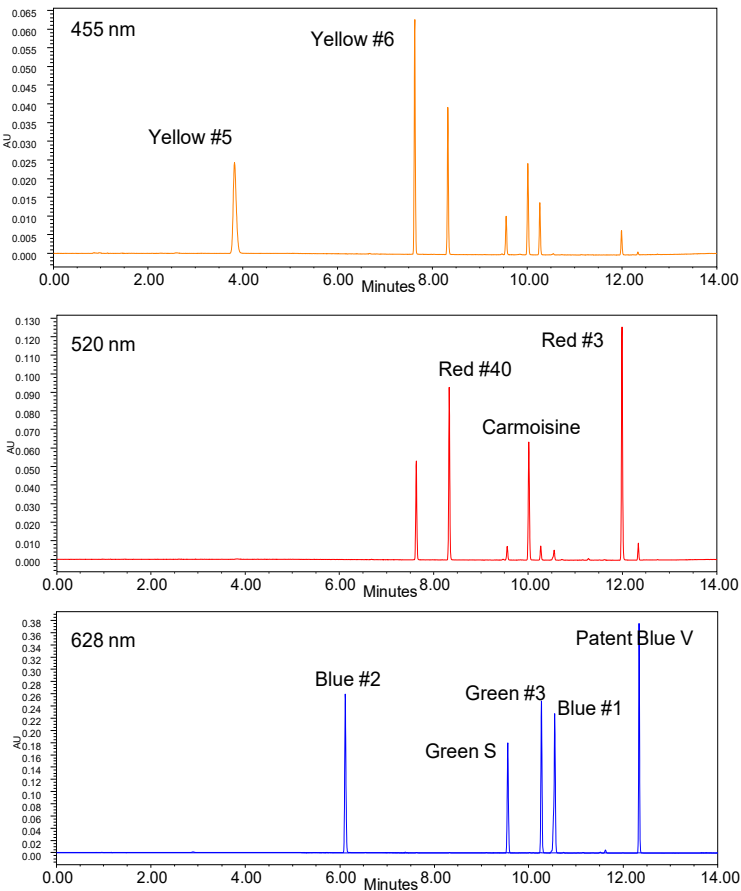
RESULTS AND DISCUSSION

The HPLC method provided baseline separation of all reference standards in the dye mixture. Using the Empower 3 Software Method Set chromatograms at various wavelengths were visualized, and peaks quantified from the 3-D, 200-800 nm chromatogram. Yellow dyes were visualized by extracting a UV channel at 455 nm, while 520 nm was utilized for red dyes, and 628 nm utilized for both green and blue dyes (**Figure 1**). A PDA Library was created from the reference standard mixture injections after wavelength extraction. The chromatographic separation method showed linearity between 0.003 ug and 0.750 ug, mass on column (data not shown).

Peak purity analysis was performed for each dye peak in the sample solutions using the Empower 3 Software, Processing Method. As a representative example, dyes in fruit flavored candy were visualized at the respective wavelengths, as shown in **Figure 2, A**. Peak purity for dye peaks was determined by the software through automated PDA spectral Purity Angle and Purity Threshold. In the fruit flavored candy, the Purity Angle was below the spectral Purity Threshold of a selected matrix-only region of the baseline, therefore chromatographic peaks were spectrally pure and absent of co-eluting constituents (**Figure A, B**). Dyes present in the fruit flavored candy were identified as Yellow #5, Yellow #6, Red #40, Blue #1 and Blue #2 using the Match Angle of reference spectra stored in the PDA Library (**Figure 2, C**).

Samples were ranked according to the total quantity of synthetic food dye per serving (**Table 1**). The highest amount was found in the spicy hot snack sticks. The product contained primarily Yellow #6, combined with a relatively low quantity of Red #40.

**Figure 1: Chromatograms extracted to visualize at yellow dyes), 520nm (red dyes), and 628nm (blue dyes). In the reference standard mixture.**



**Figure 2: A) Extracted chromatograms for yellow, red, and blue dyes in mixed fruit candy. For Yellow #5, B) Peak purity assessment and C) PDA library match for peak identification.**

The visual appearance, a deep yellow, flour derived product heavily coated in red powder, correlated with the dye ratio reflected by the chromatographic results. Additionally, upon extraction, the product showed the most intense color by visual inspection when compared with other samples.

The second highest dye content per serving was observed in beverages. The single serving size of sports drinks was 360 mL. As a result, the mass of dye per serving was relatively high compared to small serving size products. Chewy fruit candy, assorted coated chocolate, and jellybeans, packaged in approximately 2 oz bags per serving, showed the highest combination of different dyes (i.e. Red #40, Blue #1, Blue #2, Yellow #5 and Yellow #6) when compared to other samples. The dye content in the candy ranged between 4.8 mg and 18.6 mg per serving.

The quantity of dye per serving in individually wrapped, single flavor, sour hard candy ranged between 0.1 mg and 0.6 mg per serving. Each individually wrapped, sour hard candy weighed approximately 3.0 g, and was 1.0 cm<sup>2</sup> in diameter, which is a relatively small serving size. OTC drugs and cosmetics were the lowest of the samples tested, again due to the relatively small serving size. Over the counter syrup, gelcaps and tablets, contained 0.5 mg of dye or less per serving, which was equivalent to mouthwashes and toothpastes, which are not intended for ingestion by the consumer.

**Table 1: Highest synthetic dye per serving for common products tested.**

Type	Product	Red #40 (520nm)	Red #3 (520nm)	Green #3 (628nm)	Blue #1 (628nm)	Blue #2 (628nm)	Yellow #5 (455nm)	Yellow #6 (455nm)	Dye per Serving (mg)
Snack Food	Spicy Hot Snack Sticks	0.6	-	-	-	-	-	26.7	27.3
Sports Drink	Apple Beverage	-	-	-	0.8	-	19.7	-	20.5
Sports Drink	Fruit Punch Beverage	20.1	-	-	-	-	-	-	20.1
Candy	Mixed Fruit Candy	4.1	-	-	0.1	12.7	0.7	1.1	18.7
Candy	Coated Chocolate	3.0	-	-	0.9	-	1.8	7.1	12.8
Sports Drink	Orange Beverage	-	-	-	-	-	-	11.2	11.2
Breakfast Cereal	Mixed Berry Flakes	1.1	-	-	0.5	-	2.3	5.8	9.7
Breakfast Cereal	Fruit Favored Rice	2.2	-	-	0.2	-	3.9	1.9	8.2
Candy	Assorted Jelly Beans	1.1	-	-	0.2	2.3	0.4	0.8	4.8
Breakfast Cereal	Fruit Wheels	1.1	-	-	0.5	-	2.3	0.6	4.5
Sports Drink	Blue Cherry Beverage	-	-	-	3.1	-	-	-	3.1
Breakfast Cereal	Marshmallows and Grains	1.0	-	-	0.2	-	1.7	0.2	3.1
Sports Drink	Lemon-Lime Beverage	-	-	-	-	-	1.8	-	1.8
Candy	Sour Cherry Hard Candy	0.8	-	-	0.4	-	-	-	1.2

CONCLUSIONS

In the work presented here, we showed a single chromatographic method for separation of ten synthetic food dyes in a variety of consumer products. Empower 3 Software peak purity analysis confirmed that the Alliance iS HPLC System with PDA Detector successfully resolved dyes from matrix constituents. Detector linearity and peak purity analysis provided accurate dye quantification, while PDA Matching identified dyes against reference spectra stored in the PDA Library. With the separation method, synthetic dyes in a variety of matrices can be quantified and identified.

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